



Recent Progress in Development of Polo Like Kinase 1 Inhibitors: Efforts So Far

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Abstract

Polo-like kinase 1 (Plk1) plays an essential role in inhibiting cell proliferation and comes under the family of serine/threonine-protein kinase, which is a particular target for cancer therapy. In some clinical studies, Plk1 has been identified as a target for cancer. Currently, so many scientists are developing the Plk1 inhibitors, and they are thinking about working on them. A recent strategy for Plk1 inhibition is the development of small-molecule inhibitors, which will inhibit the Plk1 through the ATP-binding site of the Plk1. Now new generation Plk1 inhibitors being tested clinically, which are targeting the polo box domain. This review highlights the recent progress made in the development of Plk1 inhibitors as anticancer agents.

Introduction

Polo-like kinases (Plks) belong to the family of serine/threonine protein kinases and play a prominent role in regulating the cell cycle by regulating initiation, maintenance and completion of the mitosis phase.¹ The polo-like kinase was identified firstly in 1988 with a chain of 577 amino acids from the genetic screens of *Drosophila* larval neuroblast.² Five types of Plks have been identified in the human species to date, i.e., Plk1, Plk2, Plk3, Plk4 and Plk5 and Plk1 is the most studied member from the Plks family due to its regulatory role in the cell division and genomic stability.³ It is a specific target for cancer therapy because its high expression in the cell's proliferation is responsible for tumorigenesis.⁴ A decrease in Plk1 expression activity results in inhibition of cell proliferation of cancer cell lines and xenografts.⁵⁻⁷ Plk2 plays its role in centriole duplication during the G1 to S transition of the cell cycle and plays a vital role in DNA damage checkpoint.⁸⁻¹⁰ In contrast, Plk3 is important in S-phase entry and depletion arrest in the process of cell proliferation.¹¹ The remaining Plk4 is required to maintain cell viability, and Plk5 has been implicated in the G1 phase of cell cycle arrest and subsequent apoptosis.^{12,13} Polo-like kinases consist of C-terminal, known as polo box domain (PBD) and N-terminal, a kinase domain (KD).¹⁴ The PDB is helpful for protein interactions, and the kinase domain plays an essential role in forming a binding pocket that is essential for the kinases.^{15,16}

Polo-like kinases play a promising role in regulating the cell cycle; therefore, Plks inhibition is the best strategy for

preventing tumorigenesis. However, Plk1 shows its action in regulating various cell cycle steps like centrosome maturation, mitotic entry, cytokinesis, mitotic exit, chromatin segregation, and spindle assembly.¹⁷ In the last few years, some small molecule inhibitors targeting the kinase domain are undergoing clinical trials.¹⁸ Kinase domain is the specific target for attachment and which is helpful in the inhibition of enzyme.¹⁹

The current review provides an update of various Plk1 inhibitors reported in recent years and their structure-activity relationship (SAR), underlining important structural attributes required to design specific Plk1 inhibitors and guide medicinal chemists towards them.

Plk1 Inhibitors

Protein kinases have grabbed much attention in the last several years as potential targets in treating cancer.^{20,21} In an attempt to understand the molecular mechanism that forms structural requirements for inhibition of Plk1 action, the kinase domain of the complex resembled the previously determined isolated KD structures and consists of residues (18-116) forming an N-terminal lobe and C terminal lobe formed by residues (295-311). These both lobes are connected by the short hinge region formed by the residues 117-123. The kinase domain contact area comprises C-terminal 295-311 residues, alpha-helix from the C-terminal lobe with hinge area, and an N-terminal beta-strand. In contrast, PBD has two polo boxes, namely, PB1 and PB2, connected by two connecting linkers. One

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is the short linker L1, formed by residues 392-402, and the other is secondary linker L2, having residues 479-490. These two linkers, L1 and L2, include the KD binding domain of PBD (Figure 1).

Currently, more than 51 kinase inhibitors are approved for the management of cancer.²² Some Plk1 inhibitors which are mentioned in Table 1. However, designing selective kinase inhibitors is challenging as most of the kinases share the structural similarity in their ATP binding cleft, which targets most kinase inhibitors. Developing competitive ATP inhibitors has been explored so far. However, a series

of peptide-based inhibitors targeting PBD of Plk1 has also been reported to display high target specificity.²³

In 2015, Chen *et al.* designed, synthesized, and evaluated novel benzimidazole and indole-based molecules as potential Plk1 PBD inhibitors. The study focused on the logical designing of Plk1 PBD inhibitors in terms of the substrate-binding site of PDB. In their study, the authors performed the screening of an in-house database followed by molecular docking to screen CJ-032 as a lead. Furthermore, they modified the structure with different substitutions, synthesized and performed *in vitro*

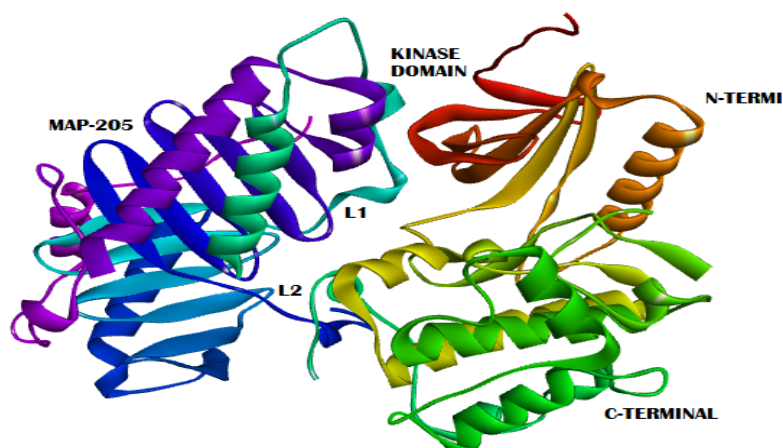


Figure 1. Structure of Plk1 showing N-lobe (orange), C-lobe (green) and kinase domain (red) joined by linkers, L1 and L2 to PBD (PDB ID 2J5F) (purple).

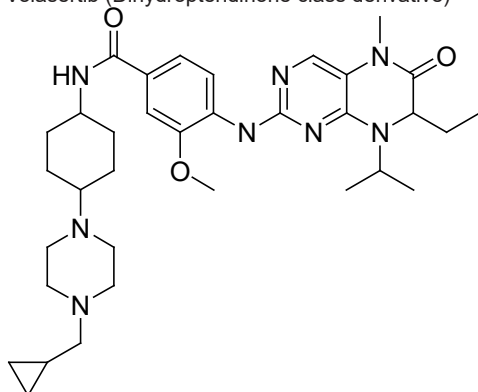
Table 1. PLK-1 kinase domain-targeted inhibitors.

Structure	Mechanism of action	IC ₅₀ values for PLK1	Selectivity
Rigosertib (ON 01910.Na) (benzylstyryl sulphone) 	A-non-ATP competitive Plk1 inhibitor; Affects microtubule dynamics. ²⁴⁻²⁷	9-10 nM	Also inhibits PDGFR,ABL, FLT1, CDK-2, PLK-2,Src, and Fyn. Efficacious both as a single agent and in combination with cytotoxic drugs in xenograft models.
Dihydropteridinone derivative (BI 2536) 	ATP-competitive inhibitor. ^{24,28-30}	0.83 nM	(i) Exhibited 1,000-fold selectivity against a wide panel of tyrosine and serine /threonine kinases (ii) PLK2b IC ₅₀ = 3.5 nM (iii) PLK3c IC ₅₀ = 9.0 nM (iv) EC50 = 2–25 nM

Table 1 Continued.

BI 6727

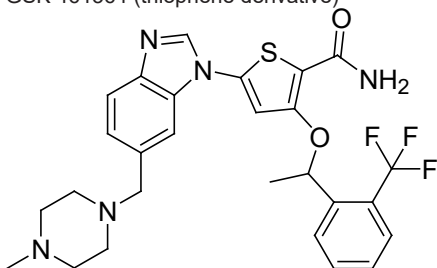
Volasertib (Dihydropteridinone class derivative)

ATP-competitive kinase inhibitor.^{31,32}

0.87 nM

(i) No inhibitory activity against a wide panel of more than 50 protein kinases
 (ii) PLK2 IC_{50} = 5 nM
 (iii) PLK3 IC_{50} = 56 nM
 (iv) EC₅₀ = 11–37 nM

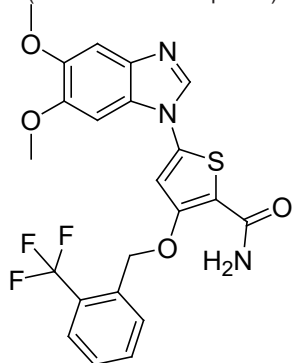
GSK 461364 (thiophene derivative)

ATP-competitive inhibitor.³³⁻³⁷

2 nM

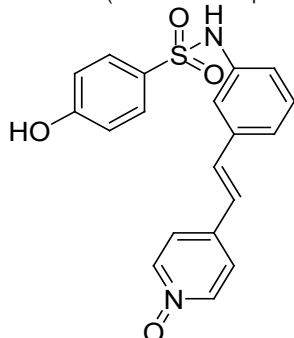
(i) No inhibitory activity against a wide panel of more than 50 protein kinases
 (ii) PLK2 IC_{50} = 5 nM
 (iii) PLK3 IC_{50} = 56 nM
 (iv) EC₅₀ = 11–37 nM

GW 843682 (benzimidazole thiophene)

ATP-competitive inhibitor.³³⁻³⁷

2.2 nM

HMN-176 (Stilbazole compound)

ATP-competitive Inhibitor.³⁸⁻⁴⁰

118 nM

Shows potent antitumor activity in gastric, breast, and lung human tumor xenografts and so forth. Better activity compared to known drugs such as cisplatin, doxorubicin, vincristine, and tegafur-uracil. Inhibits the expression of NF- κ B and induces the cell cycle arrest.

Table 1 Continued.

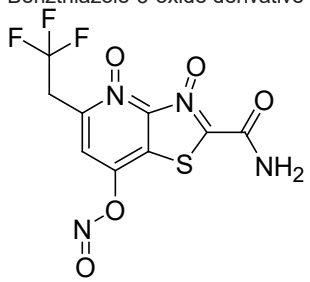
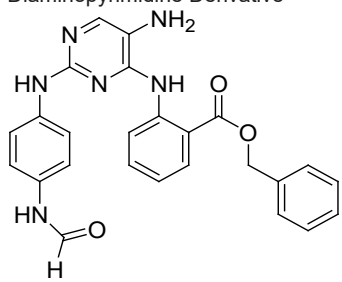
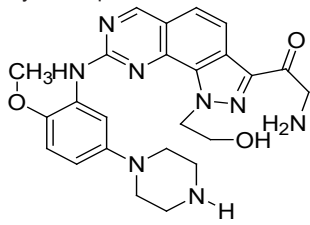
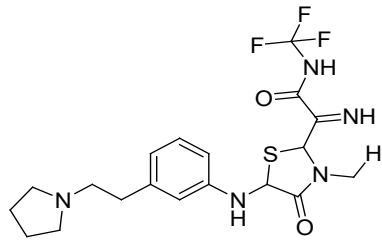
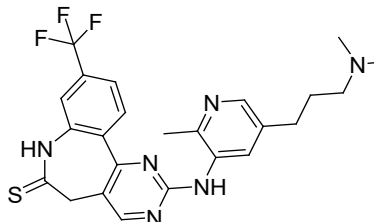
<p>Cyclapolin 1 Benzthiazole-3-oxide derivative</p> 	Non-competitive with respect to ATP. ⁴¹	20 nM	Inhibits PLK1; other family members were not determined Inhibits C terminal Src kinase; IC ₅₀ ~100 μM Cell cycle may also be affected in G1/S
<p>DAP-81 Diaminopyrimidine Derivative</p> 	Predicted to target the nucleotide pocket. ⁴²	0.9 nM	Destabilized kinetochore microtubules. Dose-dependent reduction of CDC25C phosphorylation in cells and recapitulation of key aspects of the loss-of-function phenotype for PLK1
<p>NMS-P937 Pyrazolequinazoline</p> 	ATP-competitive Inhibitor. ⁴³⁻⁴⁵	20 nM	More than 100 cell lines and 200 protein kinases have been tested Shows prolonged M phase and induce apoptosis Active in Xenograft tumor model IC ₅₀ < 100 nm on solid tumor
<p>ZK-Thiazolidinone TAL</p> 	ATP-competitive Inhibitor. ⁴⁶	19 ± 12 nM	Induced arrest in prometaphase-like arrest and finally cytokinesis failure and multinucleation IC ₅₀ = 0.2–1.3 μM on human and mouse tumor cell lines
<p>MLN0905 (benzolactam derivative)</p> 	ATP-competitive inhibitor. ⁴⁷	2 nM	Mitotic arrest on tumor growth inhibition

Table 1 Continued.

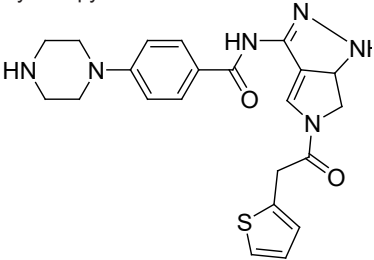
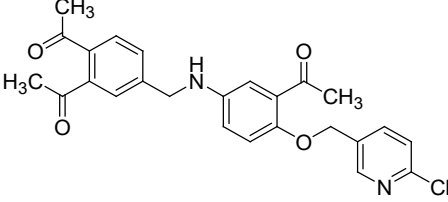
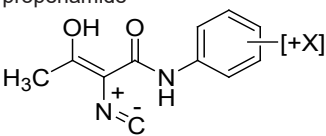
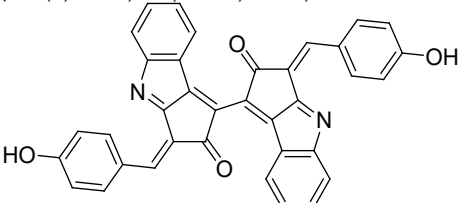
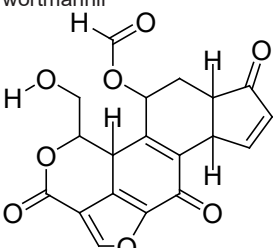
<p>PHA-680626 Pyrrolo-pyrazole derivative</p> 	ND	0.53 nM	<p>PLK-2 ($IC_{50} = 0.07 \mu M$) PLK-3 ($IC_{50} = 1.61 \mu M$) Weaker inhibition was detected on few kinases.⁴⁸</p>
<p>SBE13 Vanillin derivative</p> 	Non ATP- competitive inhibitor. ⁴⁹	12–39 μM (EC_{50})	Shows 1000-fold selectivity within the PLK family
<p>LFM-A13 α-cyano-β-hydroxy-β-methyl-<i>N</i>-(2,5-dibromophenyl)-propanamide</p> 		Plx1 32.5 μM Using GST-CDC 25 as a substrate	<p>PLK-3 $IC_{50} = 61 \mu M$. Also inhibits human BTK with an IC_{50} of 17.2 $\pm 0.81 \mu M$ The activity is 3–15 fold greater against a panel of protein kinases.⁵⁰⁻⁵²</p>
<p>Scytonemin Subunit derived from tryptophan and Phenylpropanoid isolated from many strains of cyanobacteria. (3,3-Bis((4- hydroxyphenyl)methylene)-(1,1- bicyclo-pent(b)indole) 2,2(3H,3H)-dione)</p> 	ATP-competitive Inhibitor. ^{53,54}	2.0 $\pm 1 \mu M$	Also inhibits the transcriptional factor MYT1 CDK-1, Chk-1, and PKC. Does not directly inhibit PLK1 up to 3-4 μM
<p>Wortmannin Steroidal furanoids, originally isolated from Penicillium wortmannii</p> 	ATP-competitive Inhibitor. ⁵⁵⁻⁵⁷	24 nM	Also inhibits the other member of PLK family and interacts with similar binding affinity Inhibits the PI3K

Table 1 Continued.

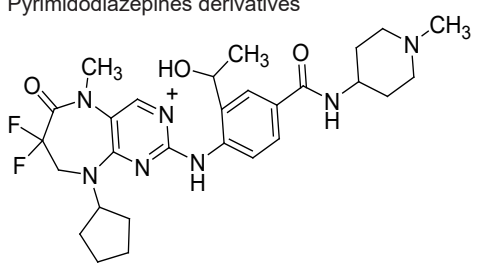
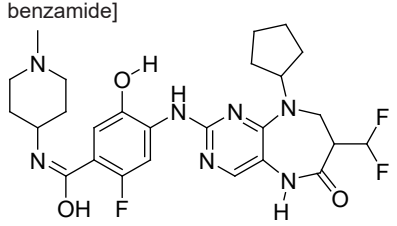
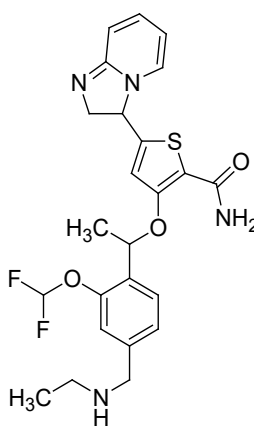
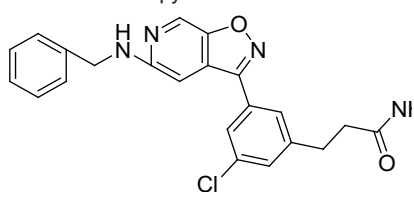
<p>RO3280</p> <p>Pyrimidodiazepines derivatives</p> 	ATP-competitive Inhibitor. ⁵⁸	0.09 nM	318 wild type and mutants protein kinases tested More than 85% protein kinases inhibits at 1mM
<p>TAK-960</p> <p>[4-[(9-cyclopentyl-7,7-difluoro-5-methyl-6-oxo-6,7,8,9-tetrahydro-5H pyrimido[4,5-b][1,4]diazepin-2-yl)amino]-2-fluoro-5-methoxy-N-(1-methylpiperidin-4-yl)benzamide]</p> 	ATP-competitive Inhibitor. ⁵⁹	0.8 nM	No inhibitory activity against 282 protein kinases Anti-tumor activity against <i>TP53</i> , <i>KRAS</i> , <i>MDR</i> mutated cell lines Monopolar spindle and G2/M phase arrest
<p>Compound 36</p> <p>Imidazopyridine derivative</p> 	ATP-competitive Inhibitor. ^{60,61}	9.8 nM	No inhibitory activity against 212 protein kinases at 1 μM. Tolerated toxicity Observed against WBC
<p>Compound 15</p> <p>2-Aminoisoxazopyridine</p> 	ATP-competitive Inhibitor. ⁶²	0.051 μM	Treated cells showed monopolar phenotype and mitotic arrest in colorectal carcinoma cell lines

Table 1 Continued.

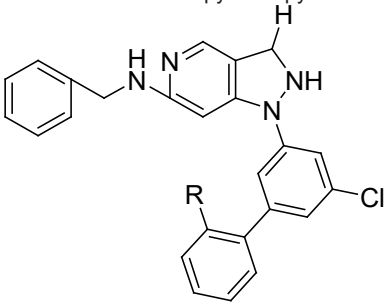
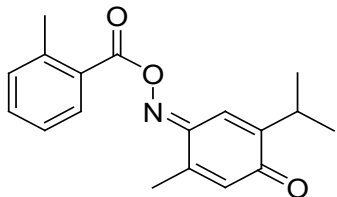
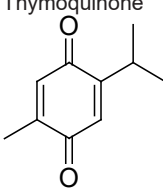
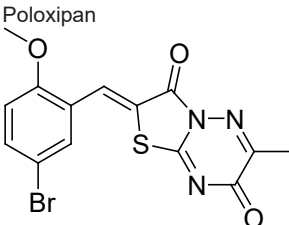
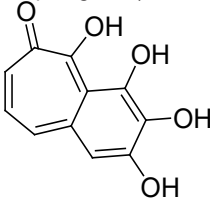
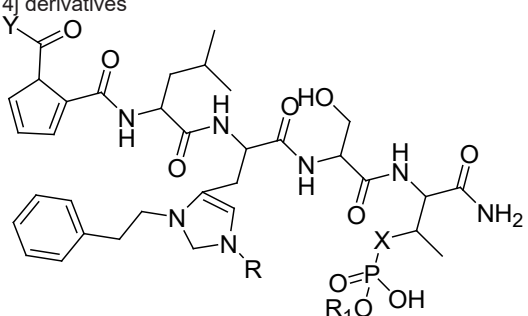
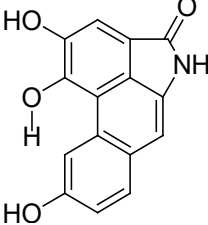
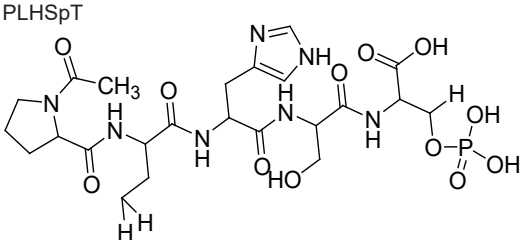
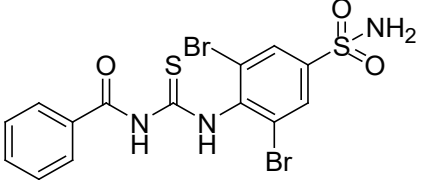
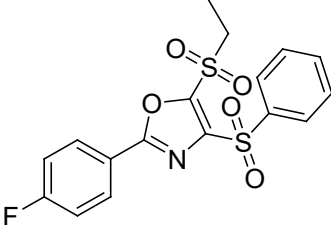
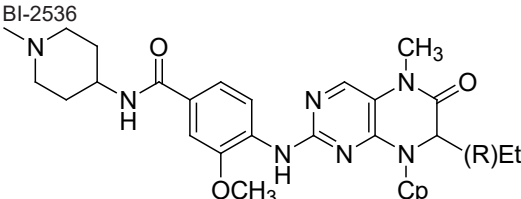
<p>Compound 38</p> <p>Derivative of 2-aminopyrazolopyridines</p> 	ATP-competitive Inhibitor. ⁶³	0.042 μ M	HCT116 colorectal cancer cell lines showed G2/M arrest and induced apoptosis
<p>Poloxin (thymoquinone derivative)</p> 	PBD site Targeting. ⁶⁴	4.8 \pm 1.3 μ M	Interferes with PLK1-PBD functions <i>in vitro</i> and <i>in vivo</i>
<p>Thymoquinone</p> 	PBD site Targeting. ⁶⁴	1.14 \pm 0.04 mM	Inhibits PBD dependent binding and subcellular localization
<p>Poloxipan</p> 	PBD site Targeting. ⁶⁵	3.2 \pm 0.3 mM	Pan inhibitor of Plk1–3 PBDs
<p>Purpurogallin (benzotropolone- containing compound)</p> 	PBD site Targeting. ⁶⁶	< 0.3 mM	Inhibits PBD-dependent binding <i>in vitro</i> and <i>in vivo</i>
<p>4j derivatives</p> 	PBD site Targeting. ^{67,68}	3 nM (320 mM)	Inhibits PBD-dependent binding <i>in vitro</i>

Table 1 Continued.

<p>Aristolactam AIIIa CCRIS 2996, Aristolactam-aii, Dibenz(cd,f)indol- 4(5H)-one, 2-hydroxy-1- methoxy</p>	PBD site Targeting. ⁶⁹	10 μ M	Inhibits kinase and PBD domain with different inhibitory concentration
			
<p>MAGPMQSpTPLGAKK (Optimal phosphopeptide sequence)</p>	PBD site Targeting. ⁷⁰⁻⁷³	5 μ M	Polo Boxtide is recognized by pincer grips like pocket PB1 and PB2
<p>LLCSpTPNG and LLCSTPNG Cdc25C-P & Cdc25C (Optimal phosphopeptide derived from Cdc25C protein)</p>	PBD site Targeting. ⁷⁴	1.8 μ M	LLCSpTPNG is recognized by the trp414 residue of PB1
<p>PLHSpT</p> 	PBD site Targeting. ^{75,76}	0.445 μ M	The side-chain of N-terminal Pro docked into a surrounding core of hydrophobic amino acid Trp414, Phe535, Arg516 residue
<p>4-(3-Benzoyl-thioureido)-3,5-dibromo-benzene sulfonamide</p>	Most selective and potent Plk1 PBD inhibitor. ²⁹	39.8 \pm 3.5 μ M	Inhibits PBD-dependent binding in vitro.
			
<p>5-(Ethylsulfonyl)-2-(4-fluorophenyl)-4-(phenylsulfonyl)oxazole (T521)</p>	T521 PBD Plk1 inhibitor. ³¹	NA	
			
<p>BI-2536</p> 	Dual Plk1 Kinase–BRD4 Bromo domain Inhibitor. ⁷⁷	BRD4: 56 \pm 9 And Plk1: 0.22 \pm 0.01	It is shown to induce mitotic arrest and apoptosis in bone marrow precursors from treated patients.

growth inhibition assay. Compound **1** shows potent Plk1 inhibition activity having IC_{50} was $6.83 \pm 0.52 \mu M$ and high sevenfold higher selectivity against some kinases like Plk2 ($IC_{50} = 14.72 \pm 4.07 \mu M$) and Plk3 ($IC_{50} \geq 50 \mu M$). The SAR studies revealed that Plk1 inhibition is greatly affected by the phenyl ring's electron density (Figure 2). The authors also found the mechanism of action, which shows that compound **1** was an ATP-independent and substrate-dependent Plk1 inhibitor.⁷⁶

Liu *et al.*⁷⁷ in 2015, reported the synthesis and biological evaluation of indole-3-carboxylic acids for a small molecular non-ATP-competitive inhibition of Plk1, as shown in Figure 3. They have calculated the inhibitory activity of Plk1 and *in vitro* cell lines growth inhibition of eleven indole-3-carboxylic acid derivatives using thymoquinone, a naturally occurring product reported as a non-ATP inhibitor of the Plk1 as a control. Compound **2** ($IC_{50} = 0.41 \pm 0.09 \mu M$) and **3** ($IC_{50} = 0.13 \pm 0.02 \mu M$) showed significantly more promising Plk1 inhibitory activity compared with thymoquinone. SAR investigation from biological data revealed that the carbon chain's length between the indole nucleus and acetic acid greatly influences the Plk1 inhibition. The compounds are substituted by 4-methyl piperazine and morpholine as the side branch exhibits the same activity as that of thymoquinone and lead CJ054. When the compound was substituted for piperidine (**3**) and pyrrolidine (**4**) as a side chain, it emerged as the most potent chemical in the series. Compound **3** was also investigated for selection of other similar kinases (Plk2 and Plk3) and demonstrated the excellent function of the Plk1

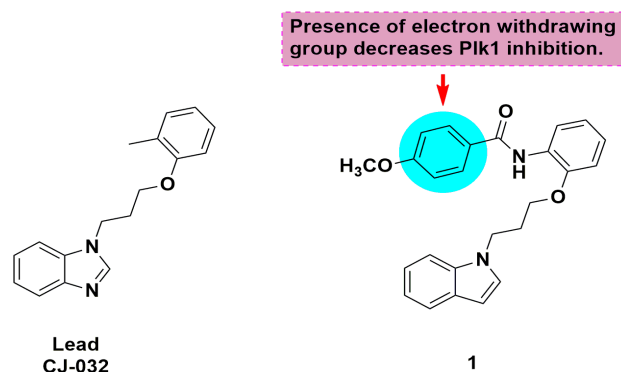
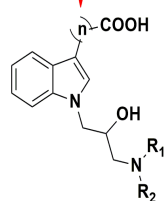


Figure 2. Non-ATP competitive benzimidazole lead CJ-032 and its indole derivatives as Plk1 PBD inhibitors.

Chain length greatly influences the Plk1 inhibitory activity.



	n	NR ₁ R ₂
Compound 2	1	Piperidine
Compound 3	1	Pyrrolidine

Indole-3-carboxylic acids derivatives

Figure 3. Indole-3-carboxylic acids derivatives reported as Plk1 PBD inhibitors.

inhibition specificity towards Plk2 ($IC_{50} > 50 \mu M$) and Plk3 ($IC_{50} > 50 \mu M$).

In 2015, Scharow *et al.*⁷⁸ reported their studies on poloxin analogs as Plk1 PBD inhibitors. Poloxin induces mitotic arrest due to chromosome impairment in Plk1 localization leads to apoptosis, thereby treating tumor cells. Authors focused on synthesizing the polo-box domain of Plk1 inhibitors for good action and specificity. All the compounds tested against the polo-box domain of Plk1-Plk3 were evaluated by the fluorescence polarization assay method. Compound **4** displayed the most significant Plk1 PBD ($IC_{50} = 0.31 \pm 0.02 \mu M$) inhibitory activity and 7-fold and 59-fold specificity with the polo-box domain of Plk2 ($IC_{50} = 2.32 \pm 0.44 \mu M$) and Plk3 ($IC_{50} = 18.3 \pm 1.8 \mu M$). The SAR investigations suggested that the aromatic ring's substitutions significantly affect the Plk1 inhibitory activity by tailoring the ester group's interaction. The presence of an electron-withdrawing group or change in the ortho-substituted methyl substituent increases the action (Figure 4). Simultaneously, the aromatic ring's replacement with aliphatic residue displayed similar potency, showing that the aromatic nucleus is not essential for biological action. Also, the ester group's replacement with acyl hydrazone abolished the activity due to loss of protein acylation, thereby suggesting acylation of protein for the poloxin mechanism of action and its analogs.

In 2016, Long *et al.*⁷⁹ studied newly synthesized heteroaryl styryl sulfone analogs as mechanistic mimetic of rigosertib. This synthesized benzyl styryl sulfone is under clinical trials for Myelodysplastic syndrome (MDS) treatment and

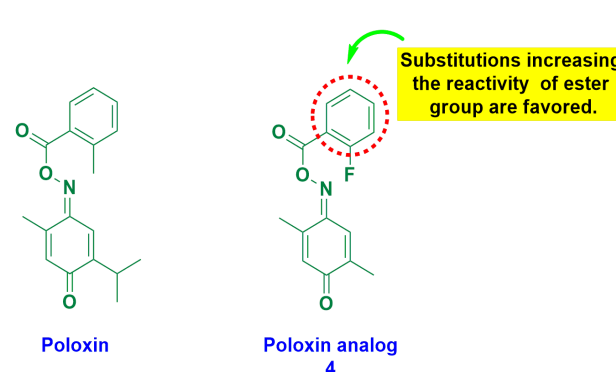


Figure 4. Poloxin and its derivative as Plk1 PBD inhibitor.

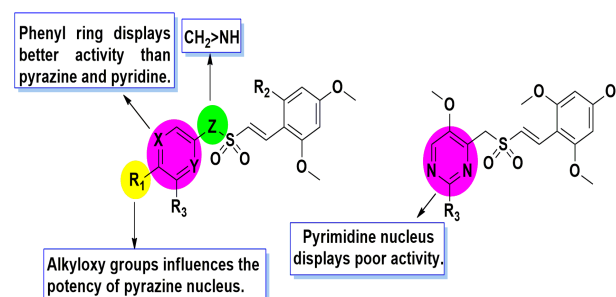


Figure 5. Rigosertib analogs reported as potential anti-cancer agents.

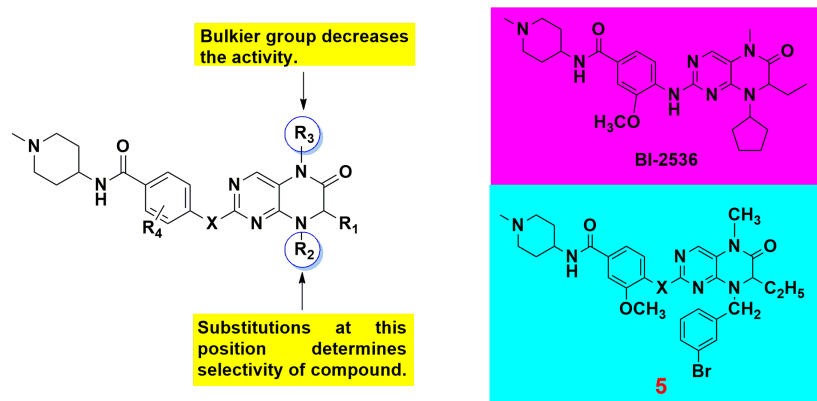


Figure 6. Dual Plk1/BRD4 inhibitors identified for management of cancer.

other cancers. Rigosertib targets Plk1, controlling the cell cycle progression, which helps the cyclin B1 activation and CDC25C phosphatase ATP dependent system. They synthesized novel styryl sulfonyl analogs bearing some nitro-heteroaryl systems. Compounds bearing simple phenyl rings displayed the most promising results by inhibiting CDC25C and demonstrated growth inhibition in various tumor cell lines in the range of 0.01-0.35 μM . It was hypothesized that the Plk1 inhibition modulates CHK2 and p53, which eventually results in a decrease of CDC25C expression. The SAR studies revealed that the compounds bearing pyrazine nucleus as heterocyclic core displayed promising activity than the pyrimidine derivatives, suggesting that the number and position of heteroaryl nitrogen atom significantly affects the antiproliferative activities (Figure 5).

In the same year, Chen *et al.*⁸⁰ reported the synthesis and biological evaluation of Plk1 inhibitor, BI-2536 analogs, as dual Plk1 kinase/BRD4 bromodomain inhibitors (Figure 6). The BRD proteins are known to be 'epigenetic readers' and can identify histones of ϵ -N-acetylated lysines. Due to their crucial role in transcriptional gene regulation involved in tumor enlargement, they have an attractive druggable target for cancer management. Since BRD4 and Plk1 actively participate in mitosis, dual inhibition of both these targets by a single molecule may be a novel strategy to treat cancer. Their study reflects the compound 5 is the most active inhibitor of BRD4, having $K_i = 8.7$ nM, and equally potent Plk1 inhibitors ($K_i = 5.80$ nM). The SAR investigations suggested that the replacement of the NH group of pyrimidine with oxygen atom abolishes the Plk1 inhibitory activity by losing critical hydrogen bonding.

In the same year, Yun *et al.*⁸¹ also published the study in which they synthesized and evaluated acyl thiourea derivatives as Plk1 PBD inhibitors. They are screened an in-house database which resulted in N-((4-sulfamoylphenyl) carbamothioyl)acetamide as a starting lead displaying an excellent binding affinity to the Plk1 PBD *in vitro*. They synthesized series of acyl thiourea derivatives and also studied their binding affinities against Plk1 PBD. A series of twenty-four compounds were synthesized by keeping acyl thiourea as a core fragment, and various substitutions

were done at positions R1 and R2. R1 position of the derivative was replaced with different aromatic, aliphatic or alicyclic hydrocarbons, while various bioisosteric groups of sulfamoylphenyl group explored the R2 position. Their study reported compound 6 as the most selective and potent Plk1 PBD inhibitor having an IC_{50} value of 39.8 ± 3.5 μM (Figure 7). The lead compound's binding affinities were compared to synthesized compounds that revealed that methyl, ethyl and phenyl groups were well tolerated at position R1. In contrast, substitutions with cyclopropyl and benzyl deteriorated the binding affinities. At the same time, halogen substitutions at position R2 improved binding affinity by occupying the empty pocket in the active site of the Plk1 PBD.

In 2016, Scharow *et al.*⁸² reported the synthesis and evaluation of bifunctional Plk1 inhibitors, comprising both Plk1 ATP-competitive ligand and the Plk1 PBD inhibitors. The ATP-competitive inhibitor of Plk1 in clinical trials, BI2536, was incorporated to design both peptidic (Ac-PLHSpT based) and non-peptidic (Poloxin 2 based) Plk1 PBD inhibitors (Figure 8). The compound shown in Figure 9 was found to be the most potent compound displaying IC_{50} of 0.054 ± 0.004 μM for Plk1 PBD and also an IC_{50} of 0.038 ± 0.002 μM against Plk1. It also showed an excellent specificity profile for Plk1 ($K_i = 0.012 \pm 0.001$ μM) over other two kinases, Plk2 ($K_i = 5.5 \pm 0.7$ μM) and Plk3 ($K_i = 3.7 \pm 0.2$ μM). This study provided an excellent opportunity to design Plk1 inhibitors targeting both functional and enzymatic domains to overcome the specificity of ATP-competitive kinase inhibitors.

Chen *et al.*⁸³ also reported 5-(ethylsulfonyl)-2-(4-fluorophenyl)-4-(phenylsulfonyl) oxazole (T521) as PBD

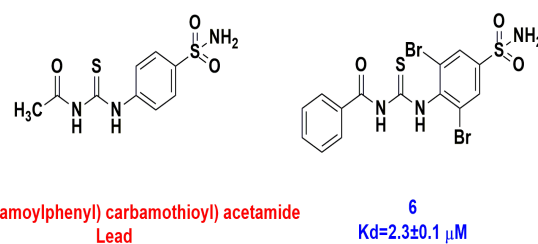


Figure 7. Acylthiourea derivatives as Plk1 PBD inhibitors.

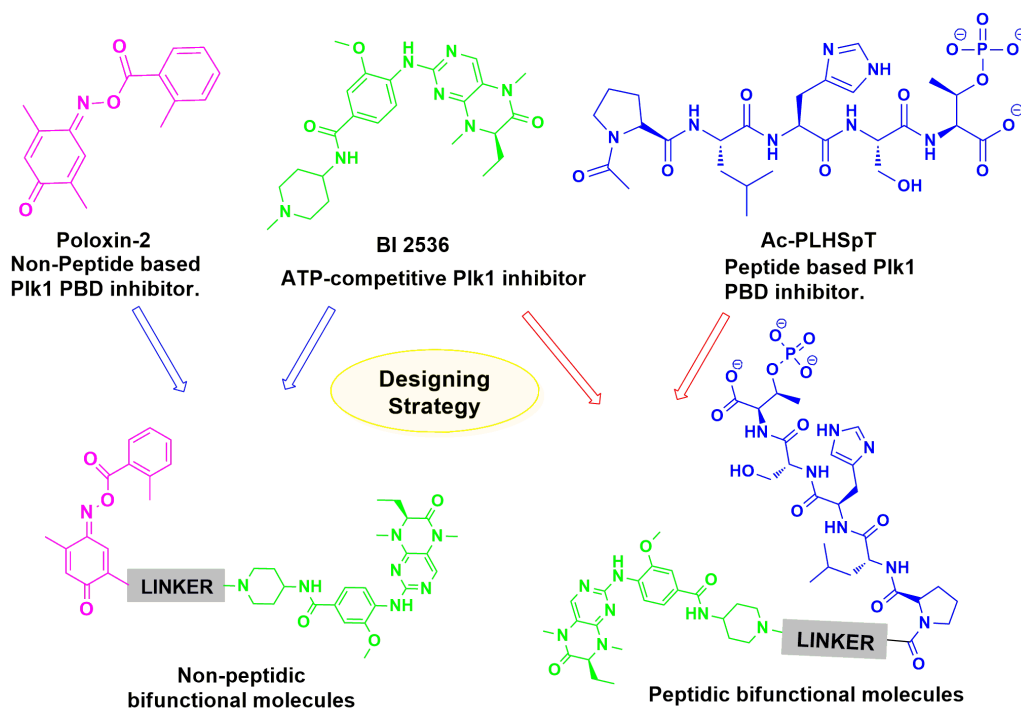


Figure 8. Designing strategy employed to synthesize bifunctional Plk1 inhibitors.

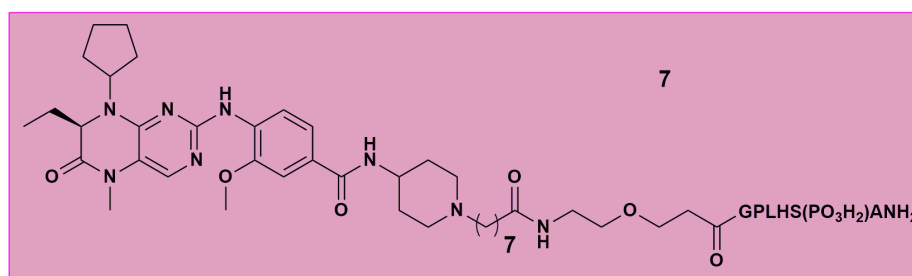


Figure 9. Most potent and selective bifunctional inhibitor of Plk1.

inhibitor of Plk1 that selectively blocks the function of PDB in Plk1. Their study utilized fluorescence polarization assay to screen the library of 20,000 molecules and reported T521 as a specific inhibitor of PDB Plk1 (Figure 10). The inhibitor covalently binds to the lysine residues in the active site of the PDB Plk1, thereby resulting in a significant change in the Plk1 secondary structure. The inhibition assay of cell proliferation studies was also carried out, which demonstrated that T521 hampers the binding of Plk1 to a checkpoint protein of spindle assembly, Bub1 *in vivo*. The compound was also found to suppress the growth of A549 cells by using xenograft mice models and showed marked mitotic defects when treated with HeLa cells. Later in the year 2017, Pan *et al.*⁸⁴ published their study in which they reported Plk1/EEF2K (polo-like kinase 1/ eukaryotic elongation factor 2 kinase) dual inhibition for the management of breast cancer. Their research created pharmacophore models for EEF2K and Plk and used them as a screening tool to retrieve EEF2K/Plk1 dual inhibitors. The ten derivatives hits were then subjected to *in vitro* studies; five derivatives displayed EEF2K and Plk1 inhibition. By analyzing the binding mode of the top

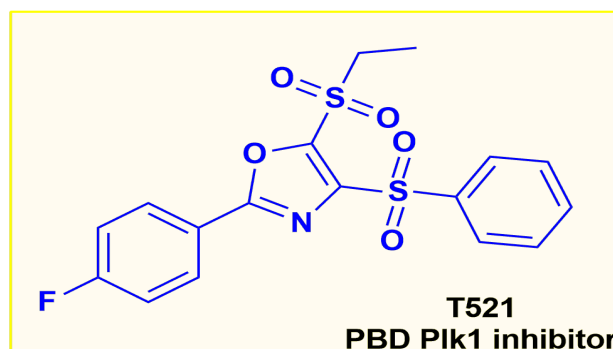


Figure 10. Structure of T521 reported as specific PDB Plk1 inhibitor.

hits, various derivatives were synthesized and evaluated. Compound substituted R1 position with ethyl group, R2 with trifluoromethyl group and R3 position with chlorine substitution, displayed the most potent novel Plk1/EEF2K dual inhibition IC₅₀ values of 0.085 and 0.762 μM respectively. In contrast, compounds containing R1 position substituted with cyclopropyl group or R3 position naphthalene group displayed poor EEF2K inhibition. SAR

analysis revealed that hydrophobic groups are less tolerated at position R3 (Figure 11).

Later in 2017, Nogawa and his research group isolated trachyspic acid 19-butyl ester from the fungus RKGFSF2684 and reported it as Plk1 PBD dependent inhibitor (Figure 12). The isolated compound's inhibitory potential was determined by performing *in vitro* studies using purpurogallin, a potent Plk1 PBD inhibitor as a positive control. The compound displayed the activity with an IC_{50} of 102 μ M, similar to purpurogallin (114 μ M). Thus, the study suggests that microbial fraction library can also be utilized to develop more potent compounds for cancer management of cancer.⁸⁵

Sun *et al.*⁸⁶ also reported a study in which they utilized an *in-silico* modification technique to identify potent Plk1

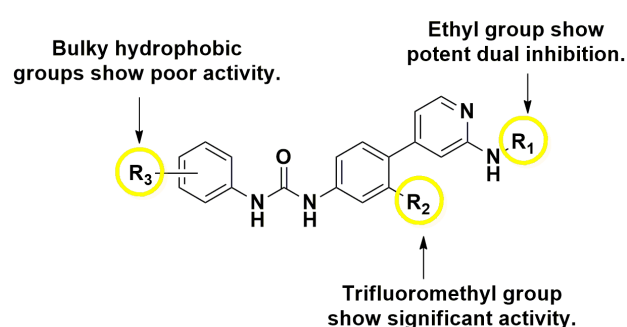


Figure 11. SAR analysis of novel PLK1/EEF2K dual inhibitors.

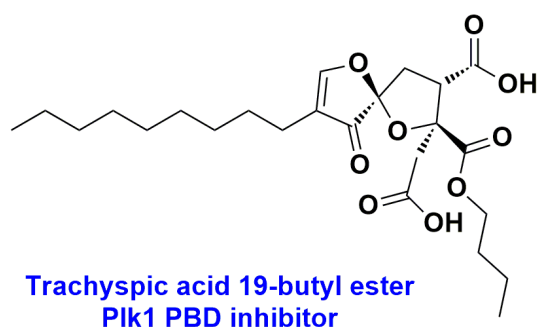
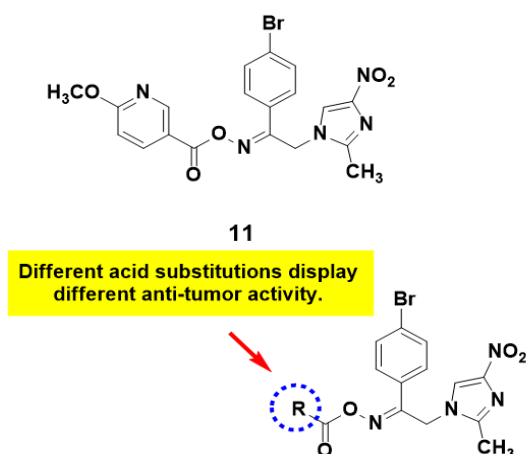


Figure 12. Structure of ester derivative of trachyspic acid reported as Plk1 PBD inhibitor



PBD inhibitors. They employed docking studies and *in vitro* primary screening in combination to identify novel leads as Plk1 PBD inhibitors. The retrieved lead was derivatized, synthesized and evaluated using various bioassays. They synthesized twenty-two nitroimidazole oxime-based derivatives and evaluated them for antiproliferative activity *in-vitro*. The SAR analysis revealed that niacin substitution at the R position resulted in better activity than benzoic acid derivatives. The unsubstituted niacin ring (IC_{50} =0.01 μ g/ml) and isonicotinic ring (IC_{50} =0.02 μ g/ml) displayed better activity than the compounds substituted with niacin ring bearing chloro-group (IC_{50} =0.71-1.28 μ g/ml) against four cancer cell lines (human). The synthesized compounds were also evaluated to affirm their Plk1 inhibitory activity using thymoquinone as a positive control. Compound **11** emerged as the most potent compound with Plk1 PBD inhibitory activity in the range of 0.002 ± 0.02 μ g/ml. The compound also displayed better potency than poloxin against the MGC803 cancer cell line (Figure 13).

To study the binding mode of the synthesized compounds, the most potent **compound 11** was docked in the active site of polo-box domain Plk1 (PDB code:4HCO). The molecular docking results revealed that the compound occupied the active site by forming three hydrogen bonds with Lys540 and Arg 557, one Pi-cation bond with Lys540, and one Pi-Pi bond with Trp 414. Docking simulations showed that the nitroimidazole ring displays good binding with the Lys540-His538 pincer clinches in the electrostatic binding region (EBR) and 6-methoxy pyridine substitution accurately occupies hydrophobic motif (HM) region similar to the previously reported thymoquinone, as Plk1 PBD inhibitor. **Compound 11** also displays some Van der Waals interactions that further contribute to binding affinity.

Lin *et al.*⁸⁷ published a study in which they utilized the Fmoc-solid-phase peptide synthesis (SPPS) strategy to synthesize nine phosphopeptides with non-natural amino acids as Plk1 PBD inhibitors. The synthesized derivatives were subjected to their Plk1 PBD inhibitory potential by determining the binding affinity using fluorescence

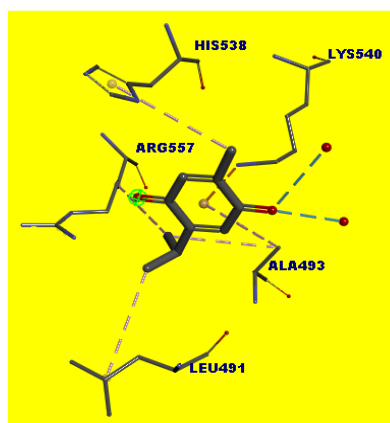


Figure 13. SAR profile of nitroimidazoleoximes as Plk1 PBD inhibitors and the stereo view of binding mode of thymoquinone in the active site of the Plk1 PBD.

polarization (FP) assay. Phosphopeptide, Ac-QTF(3,4-Cl) DPPLHSpTAIYANNH2(**12**) displayed the most significant inhibition of Plk1 PBD (IC_{50} of 38.99 nM) and were highly selective. The compound showed 600 times more selectivity over Plk3 and no binding affinity to Plk2. *In vitro* biological assay was also carried out for the phosphopeptide **12**, which revealed that compound stop the cell cycle progression by inhibiting G2/M phase transit, depending on concentration and time. In addition to this, cell membrane studies were also performed, which revealed that phosphopeptide **12** apoptosis induced by inhibition which is time and attention. According to Western blot analysis, studies demonstrated that phosphopeptide **12** increases protein cleavage of PARP caspase-3, thereby arresting G2/M phase transition of the cell cycle by controlling CyclinB1-CDK1. Also, phosphopeptide **12** penetrates the cell membranes and enters into the cytoplasm and overcomes the shortcomings of peptide-Plk1 inhibitors that cannot penetrate cell membrane.

Later the same year, Liu *et al.*⁸⁸ employed a structure-based drug design strategy to design and synthesize promising and specific multipurpose polo-like kinase 1 (Plk1)/bromodomain 4 (BRD4) inhibitors. The extra-terminal domain (BET) and bromodomain proteins family play an essential role in recognizing acetylated lysines (KAc) on chromatin. They are the target of therapeutic value in cancer treatment inflammation and viral infectious diseases. The simultaneous inhibition of both BRD4 bromodomain and Plk1 by one molecule is a good strategy for inhibiting Plk1, and BRD4 was applied in his work. The essential structural features were extracted from the BI-2535 crystal structures with Plk1 and BRD4-BD1 were removed, followed by synthesis and evaluation of various BI-2536 analogs. It was observed cyclopentyl group and methylated amide group of BI-2536 are required for the maintenance of the bromodomain binding activity, whereas substitution of bulky groups on the asymmetrical ethyl group on the

dihydropteridine nucleus resulted in compounds with potent BRD4/Plk1 inhibitory activity. Compound **13** was reported as the most active compound displaying the most potent BRD4/Plk1 dual inhibitory activity (BRD4-BD1 IC_{50} = 28 nM and Plk1 IC_{50} = 40 nM) (Figure 14).

In 2018, Hymel *et al.*⁸⁹ reported a series of peptide macrocycles that mimic the phosphopeptide binding site of Plk1 PBD. In their work, they utilized previously reported PLH*SpT (**14**) is a highly attractive ligand for Plk1 PBD as starting material and derivatized C-terminal macrocyclization of **14** employing N(π), N(bis-alkylated histidine residues as ring joint. The resulting compounds were evaluated by performing biochemical assays and displayed high target affinity and improved selectivity for Plk1-PBD. **Compound 15** emerged as an active Plk1-PBD inhibitor and with an IC_{50} of 45 nM. The crystal structure of compound **15** bound to Plk1-PBD was also solved at 1.45 Å resolution. From the binding mode of compound **15**, it was observed that the ring-closing methylene chain aided the C terminal carboxamide to acquire a favorable trans amide configuration, also assisted in retaining significant H bond interactions with the basic amide of Leu491. The polymethylene chain was involved in making hydrophobic interactions with the sidechain of Leu490, which were not observed in the case of the basic nucleus compound (Figure 15).

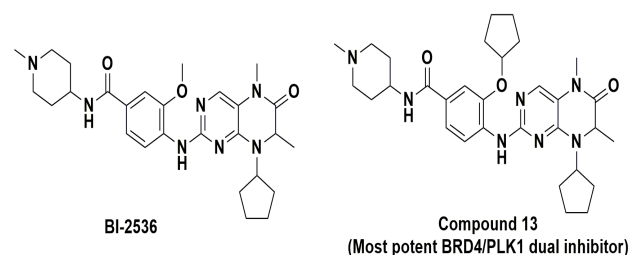


Figure 14. Most potent structural analogue of BI-2536 reported as a potent BRD4/PLK1 dual inhibitor.

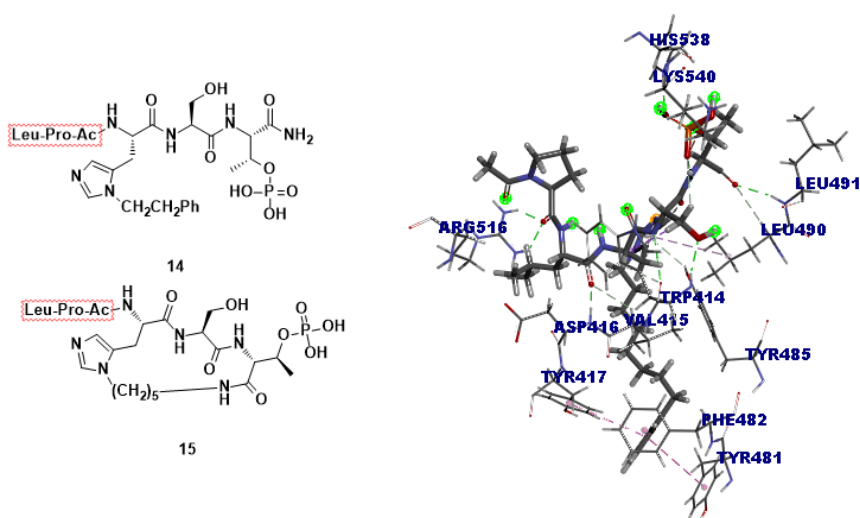


Figure 15. Most potent macrocyclic peptide (**15**), analogue of **14** reported as Plk1-PBD inhibitors and its binding mode in the active site (PDB: 3RQ7)

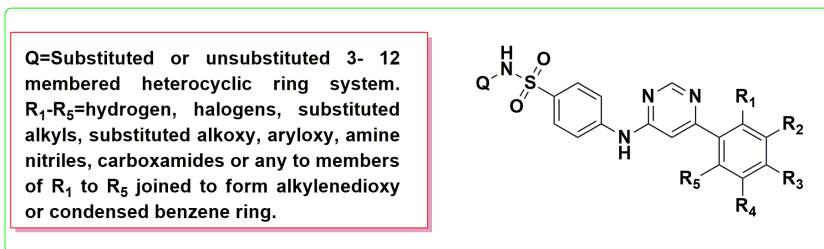


Figure 16. General structure of patented 4-pyrimidinyl amino-benzenesulfonamide derivatives as Plk1. inhibitors.

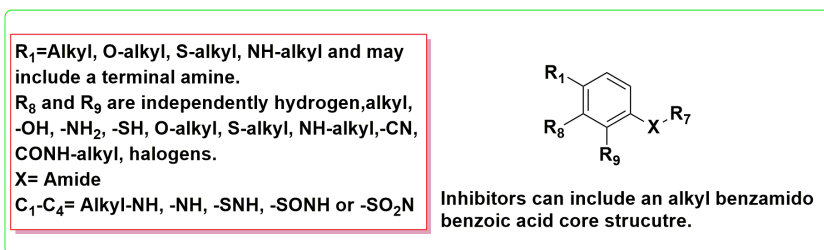


Figure 17. General structure of Plk1-PBD inhibitors.

In 2018, Liu and his research group synthesized and designed pyrrole-imidazole polyamide-Hoechst conjugate, PIP3 DNA sequence targeted Plk1 promoter. It is also responsible for specifically inhibiting cell cycle regulation which inhibits Plk1 expression and consequently retard cancer cell growth. The treatment of cancerous cells with PIP3 resulted in some mitotic misfunctions, which causes apoptosis of cells, while it will not show any effect on a normal cell by PIP3 treatment. The compound displayed the great antitumor effective activity of xenografts along with less toxicity. Hence, this PIP-Hoechst conjugate can serve as an essential lead to further develop more specific and highly selective Plk1 inhibitors.⁹⁰

Plk1 inhibitors patents

In 2014, Vichemchemie Kutato Kft. filed a patent relating to the invention of 4-pyrimidinylamino-benzenesulfonamide derivatives as Plk1 inhibitors (Figure 16). The inhibitors were evaluated by performing *in vitro*, *in vivo* and genotoxicity studies. The inhibitors have also been identified as new drug candidates for the treatment of resistant tuberculosis.⁹¹

In 2015, McInnes *et al.*⁴¹ filed two patents reporting fragment-specific inhibitors are selective against PBD. They developed fragment-specific Plk1 inhibitors, including non-peptide fragmentation that terminal of the inhibitor, the non-peptide fragment shows similar SAR of a peptide fragment selective for the polo-box domain of a Plk1peptide inhibitor. The invention was related to designing non-peptidic PBD inhibitors and the method of development of SAR for peptide fragment inhibitors and then developing non-peptide fragment alternatives for portions of the peptide inhibitors. In this work, the development of protein-protein interaction inhibitors has been applied to create fragment alternatives for portions of existing known peptide inhibitors; in this fragment alteration approach, binding key determinants

are identified for further treatment by understanding the peptide SAR.

In 2017, McInnes *et al.*⁵² filed another patent relating to non-peptide small-molecule inhibitors having the atomic mass of 1000 Daltons or less as selective for polo-like kinase proteins. The molecules were reported to target the PBD of Plk1. The inhibitors synthesized are specific for Plk1 are also much less likely to affect the activity of the Plk3 tumor suppressor, as certain of the Plk1 PBD domain inhibitors have minimal activity against Plk3. The inhibitors can be effectively displayed antitumor activity against cancer cells and can be effectively used against cancer cells that acquire resistance to ATP-based inhibitors. It was also disclosed that inhibitors could combine competitive ATP inhibitors as a synergistic means to target Plk1 clinically. Moreover, by targeting non-catalytic functions, Plk1 can be less likely to obtain resistance to the inhibitors. The general structure inhibitor is shown in Figure 17.

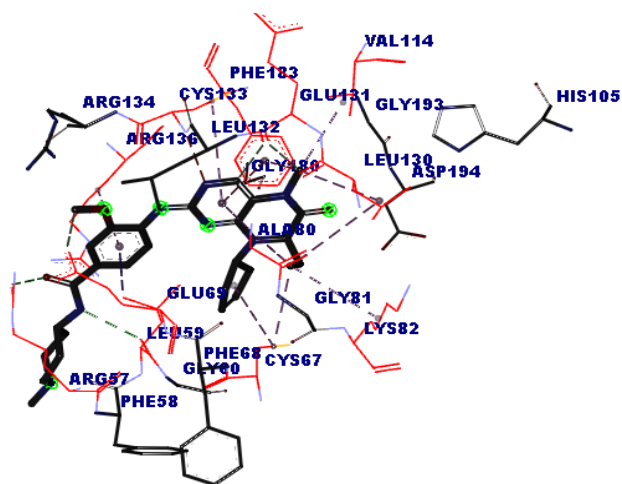


Figure 18. Stereo view of BI 2536 showing the binding mode bound to the active site. (PDB: 3RQ7).

Thus, by studying the binding orientation of different inhibitors of Plk1 reported so far, it can be observed that various amino acid residues other than those in the active site can be explicitly targeted to ensure the specificity of the newly designed derivatives. The binding mode of BI 2536, a selective Plk1 inhibitor reported so far, which is currently in clinical trials, revealed that the hinge region composed of Leu132 and Arg 136 are specific for Plk1 (Figure 18). Thus, designing molecules that will exploit this hinge region of Plk1 will result in more potent and highly selective inhibitors of Plk1 with less off-target effects.

Conclusion

Plk1 is a specific and selective target for the treatment of cancer, as its overexpression promotes tumorigenesis and plays a vital role in regulating the cell cycle, which acts as a specific oncogene for cancer development. Several Plk1 inhibitors have been reported during the past years, and many have been clinically investigated. The Plk1 contains two domains, and they are the highly conserved kinase domain at N-terminal and the PBD containing two polo boxes. So far, the classical ATP-binding site of Plk1 has been investigated to develop Plk1 inhibitors, but they suffer from the problem of selectivity. However, recently, various researchers have exploited the conserved PBD of Plk1 to report more novel and highly specific Plk1 inhibitors that inhibit the tumor suppression activity of Plk2. The efforts should be laid towards improving ATP-competitive inhibitors' specificity and developing new clinically applicable PBD inhibitors by investigating the active site residues of the PBD-binding domain. Understanding the binding mode and molecular mechanism of how the inhibitors interact with the active site of the Plk1 will provide novel insights in developing particular Plk1 inhibitors for future research activities.

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Author Contributions

RLS: Conception or design of the work and revising it critically for important intellectual content. JBW: Language editing and reviewing. RDU: Drafting of the manuscript. GDB: Drafting and revision of the manuscript. All authors have read and agreed to the published version of the manuscript.

Conflict of Interest

Authors declare no conflict of interest.

References

- Nigg E. Polo-like kinases: positive regulators of cell division from start to finish. *Curr Opin Cell Biol.* 1998;10(6):776-83. doi:10.1016/s0955-0674(98)80121-x
- Llamazares S, Moreira A, Tavares A, Girdham C, Spruce B, Gonzalez C, et al. Polo encodes a protein kinase homolog required for mitosis in *Drosophila*. *Gene Dev.* 1991;5(12a):2153-2165. doi:10.1101/gad.5.12a.2153
- Barr F, Sillje H, Nigg E. Polo-like kinases and the orchestration of cell division. *Nat Rev Mol Cell Biol.* 2004;5(6):429-441. doi:10.1038/nrm1401
- Strebhardt K, Ullrich A. Targeting polo-like kinase 1 for cancer therapy. *Nat Rev Cancer.* 2006;6(4):321-30. doi:10.1038/nrc1841
- Spankuch-Schmitt B, Wolf G, Solbach C, Loibl S, Knecht R, Stegmueller M, et al. Down regulation of human polo-like kinase activity by antisense oligonucleotides induces growth inhibition in cancer cells. *Oncogene.* 2002;21(20):3162-71. doi:10.1038/sj.onc.1205412
- Guan R, Tapang P, Levenson J, Albert D, Giranda V, Luo Y. Small interfering RNA-mediated Polo-like kinase 1 depletion preferentially reduces the survival of p53-defective, oncogenic transformed cells and inhibits tumor growth in animals. *Cancer Res.* 2005;65(7):2698-704. doi:10.1158/0008-5472.can-04-2131
- Elez R, Piiper A, Kronenberger B, Kock M, Brendel M, Hermann E, et al. Tumor regression by combination antisense therapy against Plk1 and Bcl-2. *Oncogene.* 2003;22(1):69-80. doi:10.1038/sj.onc.1206038
- Warnke S, Kemmler S, Hames R, Tsai HL, Hoffmann-Rohrer U, Fry A, et al. Polo-like kinase-2 is required for centriole duplication in mammalian cells. *Curr Biol.* 2004;14(13):1200-7. doi:10.1016/j.cub.2004.06.059
- Burns T, Fei P, Scata K, Dicker D, El-Deiry W. Silencing of the novel p53 target gene Snk/Plk2 leads to mitotic catastrophe in paclitaxel (taxol)-exposed cells. *J Mol Cell Biol.* 2003;23(16):5556-71. doi:10.1128/mcb.23.16.5556-5571.2003
- Shimizu-Yoshida Y, Sugiyama K, Rogounovitch T, Ohtsuru A, Namba H, Saenko V, et al. Radiation-inducible hSNK gene is transcriptionally regulated by p53 binding homology element in human thyroid cells. *Biochem Biophys Res Commun.* 2001;289(2):491-8. doi:10.1006/bbrc.2001.5993
- Zimmerman W, Erikson R. Polo-like kinase 3 is required for entry into S phase. *Proc Natl Acad Sci.* 2007;104(6):1847-52. doi:10.1073/pnas.0610856104
- Hudson J, Kozarova A, Cheung P, Macmillan J, Swallow CJ, Cross J, et al. Late mitotic failure in mice lacking Sak, a polo-like kinase. *Curr Biol.* 2001;11(6):441-446. doi:10.1016/s0960-9822(01)00117-8
- Andrzejczyk Z, Bernstein W, Deng L, Myer D, Li Y, Tischfield J, et al. The novel mouse Polo-like kinase 5 responds to DNA damage and localizes in the nucleolus. *Nucleic Acids Res.* 2010;38(9):2931-43. doi:10.1093/nar/gkq01
- Zitouni S, Nabais C, Jana S, Guerrero A, Bettencourt-Dias M. Polo-like kinases: structural variations lead to multiple functions. *Nat Rev Mol Cell Biol.* 2014;15(7):433-52. doi:10.1038/nrm3819

15. Cheng K, Lowe E, Sinclair J, Nigg E, Johnson L. The crystal structure of the human polo-like kinase 1 polo box domain and its phosphopeptide complex. *EMBO J*. 2003;22(21):5757-68. doi:10.1093/emboj/cdg558
16. Jani K, Dalafave D. computational Design of Targeted Inhibitors of polo-Like Kinase 1 (plk1). *Bioinform Biol Insights*. 2012;6:23-31. doi:10.4137/bbi.s8971
17. Weiß L, Efferth T. Polo-like kinase 1 as target for cancer therapy. *Exp Hematol*. 2012;1(1):38. doi:10.1186/2162-3619-1-38
18. Strebhardt K, Becker S, Matthes Y. Thoughts on the current assessment of Polo-like kinase inhibitor drug discovery. *Expert Opin Drug Discov*. 2014;10(1):1-8. doi:10.1517/17460441.2015.962510
19. Reindl W, Yuan J, Krämer A, Strebhardt K, Berg T. Inhibition of polo-like kinase 1 by blocking polo-box domain-dependent protein-protein interactions. *Chem Biol*. 2008;15(5):459-66. doi:10.1016/j.chembiol.2008.03.013
20. Zhang J, Yang P, Gray N. Targeting cancer with small molecule kinase inhibitors. *Nat Rev Cancer*. 2009;9(1):28-39. doi:10.1038/nrc2559
21. Singh P, Singh H, Silakari O. Kinases inhibitors in lung cancer: from benchside to bedside. *Biochim Biophys Acta*. 2016;1866(1):128-40. doi:10.1016/j.bbcan.2016.07.002
22. Chahrour O, Cairns D, Omran Z. Small molecule kinase inhibitors as anti-cancer therapeutics. *Mini Rev Med Chem*. 2012;12(5):399-411. doi:10.2174/138955712800493915
23. Berg A, Berg T. Inhibitors of the Polo Box Domain of Polo-like kinase 1. *Chem Bio Chem*. 2016;17(8):650-656. doi:10.1002/cbic.201500580
24. Strebhardt K. Multifaceted polo-like kinases: drug targets and antitargets for cancer therapy. *Nat Rev Drug Discov*. 2010;9(8):643-660. doi:10.1038/nrd3184
25. Gumireddy K, Reddy M, Cosenza S. ON01910, a non-ATP-competitive small molecule inhibitor of Plk1, is a potent anticancer agent. *Cancer Cell*. 2005;7(3):275-86. doi:10.1016/j.ccr.2005.02.009
26. Kothe M, Kohls D, Low S, Coli R, Cheng A, Jacques S, et al. Structure of the catalytic domain of human polo-like kinase 1. *Biochemistry*. 2007;46(20):5960-71. doi:10.1021/bi602474j
27. Jimeno A, Chan A, Cusatis G, Zhang X, Wheelhouse J, Solomon A, et al. Evaluation of the novel mitotic modulator ON 01910. Na in pancreatic cancer and preclinical development of an ex vivo predictive assay. *Oncogene*. 2008;28(4):610-618. doi:10.1038/onc.2008.424
28. Lenart P, Petronczki M, Steegmaier M, Fiore Di, Lipp B, Hoffmann J, et al. The small molecule inhibitor BI 2536 reveals novel insights into mitotic roles of polo-like kinase 1. *Curr Biol*. 2007;17(4):304-15. doi:10.1016/j.cub.2006.12.046
29. Steegmaier M, Hoffmann M, Baum A, Lenart P, Petronczki M, Krssak M, et al. BI 2536, a potent and selective inhibitor of polo-like kinase 1, inhibits tumor growth in vivo. *Curr Biol*. 2007;17(4):316-22. doi:10.1016/j.cub.2006.12.037
30. Kothe M, Kohls D, Low S, Coli R, Rennie G, Feru F, et al. Selectivity-determining residues in Plk1. *Chem Biol Drug Des*. 2007;70(6):540-6. doi:10.1111/j.1747-0285.2007.00594.x
31. Rudolph D, Steegmaier M, Hoffmann M, Grauert M, Baum A, Quant J, et al. BI 6727, a Polo-like kinase inhibitor with improved pharmacokinetic profile and broad antitumor activity. *Clin Cancer Res*. 2009;15(9):3094-102. doi:10.1158/1078-0432.ccr-08-2445
32. Schoffski P, Awada A, Dumez H, Gil T, Bartholomeus S, Wolter P, et al. A phase I, dose-escalation study of the novel Polo-like kinase inhibitor volasertib (BI 6727) in patients with advanced solid tumours. *Eur J Cancer*. 2012;48(2):179-186. doi:10.1016/j.ejca.2011.11.001
33. Emmitte K, Adjabeng GM, Andrews C, Alberti J, Bambal R, Chamberlain S, et al. Design of potent thiophene inhibitors of polo-like kinase 1 with improved solubility and reduced protein binding. *Bioorg Med Chem Lett*. 2009;19(6):1694-7. doi:10.1016/j.bmcl.2009.01.094
34. Emmitte K, Andrews C, Badiang J, Davis-Ward R, Dickson H, Drewry D, et al. Discovery of thiophene inhibitors of polo-like kinase. *Bioorg Med Chem Lett*. 2009;19(3):1018-21. doi:10.1016/j.bmcl.2008.11.041
35. Gilmartin A, Bleam M, Richter M, Erskine S, Kruger R, Madden L, et al. Distinct concentration-dependent effects of the polo-like kinase 1- specific inhibitor GSK461364A, including differential effect on apoptosis. *Cancer Res*. 2009;69(17):6969-77. doi:10.1158/0008-5472.can-09-0945
36. Murugan R, Park J, Kim E, Shin S, Cheong C, Lee K, et al. Plk1-targeted small molecule inhibitors: molecular basis for their potency and specificity. *Mol Cells*. 2011;32(3):209-20. doi:10.1007/s10059-011-0126-3
37. Didier C, Cavelier C, Quaranta M, Demur C, Ducommun B. Evaluation of Polo-like kinase 1 inhibition on the G2/M check point in Acute Myelocytic Leukaemia. *Eur J Pharmacol*. 2008;591(1-3):102-5. doi:10.1016/j.ejphar.2008.06.062
38. Yuan J, Horlin A, Hock B, Stutte H, Rubsamens-Waigmann H, Strebhardt K. Polo-like kinase, a novel marker for cellular proliferation. *Am J Clin Pathol*. 1997;150(4):1165-72.
39. Garland L, Taylor C, Pilkington D, Cohen J, von-Hoff D. A phase I pharmacokinetic study of HMN-214, a novel oral stilbene derivative with polo-like kinase-1-interacting properties, in patients with advanced solid tumors. *Clin Cancer Res*. 2006;12(17):5182-9. doi:10.1158/1078-0432.ccr-06-0214
40. Tanaka H, Ohshima N, Ikenoya M, Komori K, Katoh F, Hidaka H. HMN-176, an active metabolite of the synthetic antitumor agent HMN-214, restores chemosensitivity to multidrug-resistant cells by targeting the transcription factor NF- κ B. *Cancer Res*.

- 2003;63(20): 6942-7.
41. McInnes C, Mazumdar A, Mezna M, Meades C, Midgley C, Scaerou F, et al. Inhibitors of Polo-like kinase reveal roles in spindle-pole maintenance. *Nat Chem Biol.* 2006;2(11):608-17. doi:10.1038/nchembio825
42. Peters U, Cherian J, Kim J, Kwok B, Kapoor T. Probing cell-division phenotype space and Polo-like kinase function using small molecules. *Nat Chem Biol.* 2006;2(11):618-26. doi:10.1038/nchembio826
43. Beria I, Ballinari D, Bertrand J, Borghi D, Bossi R, Brasca M, et al. Identification of 4,5-dihydro-1H-pyrazolo[4,3 h] quinazoline derivatives as a new class of orally and selective Polo-like kinase 1 inhibitors. *J Med Chem.* 2010;53(9):3532-51. doi:10.1021/jm901713n
44. Beria I, Bossi R, Brasca M, Caruso M, Ceccarelli W, Fachin G, et al. NMS-P937, a 4,5-dihydro-1H-pyrazolo[4,3h] quinazoline derivative as potent and selective Polo-like kinase 1 inhibitor. *Bioorg Med Chem Lett.* 2011;21(10):2969-74. doi:10.1016/j.bmcl.2011.03.054
45. Valsasina B, Beria I, Alli C, Alzani R, Avanzi N, Ballinari D, et al. NMS-P937, an orally available, specific small-molecule polo-like kinase 1 inhibitor with antitumor activity in solid and hematologic malignancies. *Mol Cancer Ther.* 2012;11(4):1006-16. doi:10.1158/1535-7163.mct-11-0765
46. Santamaria A, Neef R, Eberspacher U, Eis K, Husemann M, Mumberg D, et al. Use of the novel Plk1 inhibitor ZK-thiazolidinone to elucidate functions of Plk1 in early and late stages of mitosis. *Mol Biol Cell.* 2007;18(10):4024-36. doi:10.1091/mbc.e07-05-0517
47. Kothe M, Kohls D, Low S, Coli R, Cheng AC, Jacques S, et al. Structure of the catalytic domain of human polo-like kinase 1. *Biochemistry.* 2007;46(20):5960-71. doi:10.1021/bi602474j
48. Keppner S, Proschak E, Schneider G, Spankuch B. Identification and validation of a potent type II inhibitor of inactive polo-like kinase 1. *Chem Med Chem.* 2009;4(11):1806-9. doi:10.1002/cmdc.200900338
49. Uckun F, Dibirdik I, Qazi S, Vassilev A, Ma H, Mao C, et al. Anti-breast cancer activity of LFM-A13, a potent inhibitor of Po-lo-like kinase (PLK). *Bioorg Med Chem.* 2007;15(2):800-14. doi:10.1016/j.bmc.2006.10.050
50. Uckun F, Dibirdik I, Sarkissian A, Qazi S. In vitro and in vivo chemosensitizing activity of LFM-A13, a dual-function inhibitor of Bruton's tyrosine kinase and polo-like kinases, against human leukemic B-cell precursors. *Arzneimittel-forsch.* 2011;61(4):252-9. doi:10.1055/s-0031-1296196
51. Mahajan S, Ghosh S, Sudbeck E, Zheng Y, Downs S, Hupke M, et al. Rational design and synthesis of a novel anti-leukemic agent targeting Bruton's tyrosine kinase (BTK), LFM-A13 [α -cyano- β -hydroxy- β -methyl-N-(2, 5-dibromophenyl)propanamide]. *J Biol Chem.* 1999;274(14):9587-99. doi:10.1074/jbc.274.14.9587
52. McInnes C, Mezna M, Fischer P. Progress in the discovery of Polo-like kinase inhibitors. *Curr Top Med Chem.* 2005;5(2):181-197. doi:10.2174/1568026053507660
53. Zhang G, Zhang Z, Liu Z. Scytonemin inhibits cell proliferation and arrests cell cycle through down regulating Plk1 activity in multiple myeloma cells. *Tumor Biol.* 2013;34(4):2241-7. doi:10.1007/s13277-013-0764-5
54. Elling R, Fucini R, Romanowski M. Structures of the wild-type and activated catalytic domains of *Brachydanio rerio* Polo-like kinase 1 (Plk1): changes in the active-site conformation and interactions with ligands. *Acta Crystallogr D.* 2008;64(9):909-18. doi:10.1107/s0907444908019513
55. Liu Y, Shreder K, Gai W, Corral S, Ferris D, Rosenblum J. Wortmannin, a widely used phosphoinositide 3-kinase inhibitor, also potently inhibits mammalian polo-like kinase. *Chem Biol.* 2005;12(1):99-107. doi:10.1016/j.chembiol.2004.11.009
56. Johnson E, Stewart K, Woods K, Giranda V, Luo Y. Pharmacological and functional comparison of the polo-like kinase family: insight into inhibitor and substrate specificity. *Biochemistry.* 2007;46(33):9551-63. doi:10.1021/bi7008745
57. Chen S, Bartkovitz D, Cai J, Chen Y, Chen Z, Chu X, et al. Identification of novel, potent and selective inhibitors of Polo-like kinase 1. *Bioorg Med Chem Lett.* 2012;22(2):1247-50. doi:10.1016/j.bmcl.2011.11.052
58. Hikichi Y, Honda K, Hikami K. TAK-960, a novel, orally available, selective inhibitor of polo-like kinase 1, shows broad spectrum preclinical antitumor activity in multiple dosing regimens. *Mol Cancer Ther.* 2012;11(3):700-9. doi:10.1158/1535-7163.MCT-11-0762
59. Murugan R, Park J, Kim E, Shin S, Cheong C, Lee K, et al. Plk1-targeted Small molecule inhibitors: molecular basis for their potency and specificity. *Mol Cells.* 2011;32(3):209-20. doi:10.1007/s10059-011-0126-3
60. Sato Y, Onozaki Y, Sugimoto T, Kurihara H, Kamijo K, Kadowaki C, et al. Imidazopyridine derivatives as potent and selective Polo-like kinase (PLK) inhibitors. *Bioorg Med Chem Lett.* 2009;19(16):4673-8. doi:10.1016/j.bmcl.2009.06.084
61. Hanan E, Fucini R, Romanowski M, Elling R, Lew W, Purkey H, et al. Design and synthesis of 2-aminoisoxazopyridines as Polo-like kinase inhibitors. *Bioorg Med Chem Lett.* 2008;18(19):5186-9. doi:10.1016/j.bmcl.2008.08.091
62. Fucini R, Hanan E, Romanowski M, Elling R, Lew W, Barr K, et al. Design and synthesis of 2-amino-pyrazolopyridines as Polo-like kinase 1 inhibitors. *Bioorg Med Chem Lett.* 2008;18(20):5648-52. doi:10.1016/j.bmcl.2008.08.095
63. Reindl W, Yuan J, Kramer A, Strebhardt K, Berg T. Inhibition of polo-like kinase 1 by blocking polo-box domain-dependent protein-protein interactions. *Chem Biol.* 2008;15(5):459-66. doi:10.1016/j.chembiol.2008.03.013

64. Reindl W, Yuan J, Kramer A, Strebhardt K, Berg T. A pan-specific inhibitor of the polo-box domains of polo-like kinases arrests cancer cells in mitosis. *Chem biochem.* 2009;10(7):1145-8. doi:10.1002/cbic.200900059
65. Watanabe N, Sekine T, Takagi M, Iwasaki J, Imamoto N, Kawasaki H, et al. Deficiency in chromosome congression by the inhibition of Plk1 polo box domain-dependent recognition. *J Biol Chem.* 2009;284(4):2344-53. doi:10.1074/jbc.m805308200
66. Liu F, Park J, Qian W, Lim D, Graber M, Berg T, et al. Serendipitous alkylation of a Plk1 ligand uncovers a new binding channel. *Nat Chem Biol.* 2011;7(9):595-601. doi:10.1038/nchembio.614
67. Qian W, Park J, Lim D, Lai C, Kelley J, Park S, et al. Mono-anionic phosphopeptides produced by unexpected histidine alkylation exhibit high plk1 polo-box domain-binding affinities and enhanced antiproliferative effects in hela cells. *Biopolymers.* 2014;102:444-55. doi:10.1002/bip.22569
68. Li L, Wang X, Chen J, Ding H, Zhang H, Hu T, et al. The natural product Aristolactam AIIIa as a new ligand targeting the polo-box domain of pololike kinase 1 potently inhibits cancer cell proliferation. *Acta Pharmacol Sin.* 2009;30(10):1443-53. doi:10.1038/aps.2009.141
69. Lee K, Grenfell T, Yarm F, Erikson R. Mutation of the polo-box disrupts localization and mitotic functions of the mammalian polo kinase Plk. *Proc Natl Acad Sci.* 1998;95(16):9301-06. doi:10.1073/pnas.95.16.9301
70. Elia A, Rellos P, Haire L, Chao J, Ivins F, Hoepker K, et al. The molecular basis for phosphodependent substrate targeting and regulation of Plks by the Polo-box domain. *Cell J.* 2003;115(1):83-95. doi:10.1016/s0092-8674(03)00725-6
71. Elia A, Cantley L, Yaffe M. Proteomic screen finds pSer/pThr-binding domain localizing Plk1 to mitotic substrates. *Science.* 2003; 299(5610):1228-31. doi:10.1126/science.1079079
72. Cheng K, Lowe E, Sinclair J, Nigg E, Johnson L. The crystal structure of the human polo-like kinase-1 polo box domain and its phospho-peptide complex. *EMBO J.* 2003;22(21):5757-68. doi:10.1093/emboj/cdg558
73. Garcia-Alvarez B, de Carcer G, Ibanez S, Bragado- Nilsson E, Montoya G. Molecular and structural basis of polo-like kinase 1 substrate recognition: implications in centrosomal localization. *Proc Natl Acad Sci.* 2007;104(9):3107-12. doi:10.1073/pnas.0609131104
74. Kang M, Yang G, Place R, Charisse K, Epstein-Barash H, Manoharan M, et al. Intravesical delivery of small activating RNA formulated into lipid nanoparticles inhibits orthotopic bladder tumor growth. *Cancer Res.* 2012;72(19):5069-79. doi:10.1158/0008-5472.can-12-1871
75. Yun S, Moulaei T, Lim D, Bang J, Park J, Shenoy S, et al. Structural and functional analyses of minimal phosphopeptides targeting the polo-box domain of polo-like kinase 1. *Nat Struct Mol Biol.* 2009;16(8):876-82. doi:10.1038/nsmb.1628
76. Chen D, Huang J, Liu M, Xu Y, Jiang C. Design, Synthesis, and Evaluation of Non ATP Competitive Small Molecule Polo-like kinase 1 (Plk1) Inhibitors. *Arch Pharm (Weinheim).* 2015;348(1):2-9. doi:10.1002/ardp.201400294
77. Liu M, Huang J, Chen D, Jiang C. Identification of indole-3-carboxylic acids as non-ATP-competitive Polo-like kinase 1 (Plk1) inhibitors. *Bioorg Med Chem Lett.* 2015;25(3):431-4. doi:10.1016/j.bmcl.2014.12.060
78. Scharow A, Raab M, Saxena K, Sreeramulu S, Kudlinzki D, Gande S, et al. Optimized Plk1 PBD inhibitors based on poloxin induce mitotic arrest and apoptosis in tumor cells. *ACS Chem Biol.* 2015;10(11):2570-9. doi:10.1021/acscchembio.5b00565
79. Long Y, Yu M, Li P, Islam S, Goh A, Kumarasiri M, et al. Synthesis and biological evaluation of heteroarylstyrylsulfone derivatives as anticancer agents. *Bioorg Med Chem Lett.* 2016;26(23):5674-8. doi:10.1016/j.bmcl.2016.10.062
80. Chen L, Yap J, Yoshioka M, Lanning M, Fountain R, Rajee M, et al. BRD4 Structure-Activity Relationships of Dual PLK1 Kinase/BRD4 Bromodomain Inhibitor BI-2536. *ACS Med Chem Lett.* 2015;6(7):764-9. doi:10.1021/acsmchemlett.5b00084
81. Yun T, Qin T, Liu Y, Lai L. Identification of acylthiourea derivatives as potent Plk1 PBD inhibitors. *Eur J Med Chem.* 2016;124:229-36. doi:10.1016/j.ejmech.2016.08.043
82. Scharow A, Knappe D, Reindl W, Hoffmann R, Berg T. Development of Bifunctional Inhibitors of Polo-Like Kinase 1 with Low Nanomolar Activities Against the Polo Box Domain. *Chem Bio Chem.* 2015;17(8):759-67. doi:10.1002/cbic.201500535
83. Chen Y, Zhang J, Li D, Jiang J, Wang Y, Si S. Identification of a novel Polo-like kinase 1 inhibitor that specifically blocks the functions of Polo-Box domain. *Oncotarget.* 2017;8(1):1234-46. doi:10.18632/oncotarget.13603
84. Pan Z, Chen Y, Liu J, Jiang Q, Yang S, Guo L, et al. Design, synthesis, and biological evaluation of polo-like kinase 1/eukaryotic elongation factor 2 kinase (PLK1/EEF2K) dual inhibitors for regulating breast cancer cells apoptosis and autophagy. *Eur J Med Chem.* 2018;144:517-28. doi:10.1016/j.ejmech.2017.12.046
85. Nogawa T, Ogita N, Futamura Y, Negishi S, Watanabe N, Osada H. Trachyspic acid 19-butyl ester, a new inhibitor of Plk1 polo box domain-dependent recognition from uncharacterized fungus RKGS-F2684. *J Antibiot Res.* 2017;70(5):705-7. doi:10.1038/ja.2016.167
86. Sun J, Liu H, Xu R, Zhu H. Identification of nitroimidazole-oxime derivatives targeting the polo-box domain of polo-like kinase 1. *Bioorgan Med Chem.* 2017;25(24):6581-8. doi:10.1016/j.bmc.2017.10.035
87. Lin T, Min H, Jiang C, Niu M, Yan F, Xu L, et al. Design, synthesis and biological evaluation of phosphopeptides as Polo-like kinase 1 Polo-box domain inhibitors. *Bioorgan Med Chem.* 2018;26(12):3429-37.

- doi:10.1016/j.bmc.2018.05.014
88. Liu S, Yosief HO, Dai L, Huang H, Dhawan G, Zhang X, et al. Structure-Guided Design and Development of Potent and Selective Dual Bromodomain 4 (BRD4)/Polo-like Kinase 1 (PLK1) Inhibitors. *J Med Chem.* 2018;61:7785-95. doi:10.1021/acs.jmedchem.8b00765
89. Hymel D, Grant R, Tsuji K, Yaffe M, Burke T. Histidine N (τ)-cyclized macrocycles as a new genre of polo-like kinase 1 polo-box domain-binding inhibitors. *Bioorg Med Chem Lett.* 2018;28(19):3202-5. doi:10.1016/j.bmcl.2018.08.018
90. Liu K, Fang L, Sun H, Pan Z, Zhang J, Chen J, et al. Targeting Polo-like Kinase 1 by a Novel Pyrrole-Imidazole Polyamide–Hoechst Conjugate Suppresses Tumor Growth In Vivo. *Mol Cancer Ther.* 2018;17(5):988-1002. doi:10.1158/1535-7163.mct-17-0747
91. Greff Z, Varga Z, Keri G, Orfi L, Pato J, Banhegyi P, et al, inventors; Ecole Polytechnique Federale de Lausanne EPFL Ecole Polytechnique Vichem Chemie Kutato Kft, assignee. 4-Pyrimidinylamino-benzenesulfonamide derivatives and their use for the inhibition of polo-like kinase 1 (PLK1) for the treatment of cancer and their use for the treatment of bacterial infections. European Patent Office EP2941428A1. 11 November 2015