



Status and Speciation of Silicon and Its Interaction with Physico-Chemical Properties

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Authors' contributions

This work was carried out in collaboration among all authors. Field sampling and experimental designed was initiated by authors ERO and OJI. The Chemical & Laboratory analysis (sequential extraction of Si & soil measurement was co-ordinated by author OJI and assisted by author MAE, data analysis done by authors IUE and BJ. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJEE/2021/v14i230199

Editor(s):

(1) Prof. Angelo Paone, Liceo Scientifico Ettore Majorana, Italy.

Reviewers:

(1) Champa Lal Khatik, Skn Agriculture University, India.

(2) Paul Benyamin Timotiwu, University of Lampung, Indonesia.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/64293>

Original Research Article

Received 17 November 2020

Accepted 22 January 2021

Published 11 February 2021

ABSTRACT

The effectiveness of three different extractants soil mixtures—HCl, HCl + H₂SO₄, and DTPA-TEA, in order to determine Si from soil and the forms of Silicon as influenced by different parent materials under acidic medium.

Seven forms of Silicon; namely water soluble, specifically adsorbed, oxides bound, organic matter bound, exchangeable, residual, total viz sequential fractionation.

Extractable Si value established in this study was (50.0 mg kg⁻¹), indicating negative effect on plant physiology. The physico-chemical properties decreased significantly with increase in soil depth vs soil parent materials. In addition, the forms of Si in the parent materials decreased in the pattern RES, bound residual fractions > EXC, soluble & exchangeable fractions > OM, organic matter fraction.

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Among the properties the silt fraction, pH & OM significantly and positively correlated with the forms of silicon, with negative correlation vs clay which maybe due to silicon adsorption by clayey fraction of the soil (redox). Therefore the soil maybe be maintained and conserved for farming activities.

Keywords: Silicon (Si); adsorption; soil; chelates; speciation.

1. INTRODUCTION

Parent materials mainly control soil type, total and available micronutrient viz soils [1]. The contents of micronutrients differ by prevailing environmental conditions in soils [2]. Mineral elements plays vital role in plants enzymatic activities and other plant functions [3]. The application of small quantities of these nutrients in soils with micronutrient deficiencies, enhance crop production [4], while large quantities added to the soil alters grain size and harmful to human health. There is a small window of concentration of these elements, where plant growth is optimal. Silicon an inorganic micronutrient exhibit a mechanism to buffer plant nutrient uptake and biofortification [5,6,7&8], which enhances optimum soil-plant processes. Berkhout et al. [9] revealed that there is a significant positive correlations between soil micronutrient contents vs malnutrition. And the deficiency of one micronutrients alters crop productivity [10].

Silicon adsorption/desorption of soil solution from weathering materials constitute the uptake and assimilation by vegetation and microorganisms [11].

The different forms of silicon present in the soils are categorize into: water soluble, non-specially and specially adsorbed, Fe-Al amorphous/crystalline occluded, Si and Ni oxide occluded, organically bond and residuals associated with soil silicates [12].

The utilization of silicon mainly in household, industrial applications, and construction is of importance to human activities [13]. The status of Si forms gives a better knowledge of it chemical reactions in soils. Majority of crops such as rice, groundnut, maize, yam, tomatoes, sugarcane etc. consumed in mid-west agro-ecological zone require adequate macro and micro nutrients for maximum growth and yield. According to [14], the concentration of silicon in plants depends primarily on the phylogenetic position of the plant, compared with the environmental effects that encompass silicon concentrations in the soil and soil solution and the pH for sustainability of these crops production, there is the need to examine and know silicon status of these soils.

The understanding of silicon concentrations of these areas will help to improve the soil type & fertility. Therefore, this study was aimed at assessing the status of Silicon in soils derived from different soil type in mid – western agro-ecological zones of Nigeria via HCl, HCl + H₂SO₄, and DTPA on mineral nutrients and relationship between the forms of Silicon and some physiochemical status of the soils investigated.

2. MATERIALS AND METHODS

2.1 Study Area

The soil type collected for this study were from four locations of Mid-western, Nigeria: Sobe (basement complex, latitude 06° 05' 59" N and Longitude 05° 46' N), koko (alluvium, latitude 06° 00' 04" N and longitude 05° 28' 28.03" E) , Uhonmora (Rhodic tropudalf –shale, latitude 06° 30' 45" N and longitude 05° 50' 26") and Iyanomo (coaster plain sand, latitude 6° 05' and 6° 25' N and longitude 5° 35' E), Mid-western is in southern Nigeria.

2.2 (BCR) European Community Bureau of Reference Procedures for Metals

BCR method adopted [15], Seven forms of Silicon; namely (F1), water soluble, (F2), specifically adsorbed, (F3), oxides bound, (F4), organically bound, (F4), (F5), exchangeable, (F6), residual and (F7), total were determined by sequential fractionation, with a reference soil (SRM14513) for quality control. The Certified Reference Material was averagely 98% respectively.

STEP 1. (Soluble silicon)

One gram soil was measured into a conical flask containing 10 ml of distilled water and shaken overnight (16 hours). The sample was filtered to 100ml. Silicon was determined using the atomic absorption spectrophotometer.

STEP 2. (Exchangeable silicon)

Ten mls (10 ml) of 1M MgCl₂ was added to the residue from the soluble Silicon and shakes

overnight for 10 hours, the sample was filtered to 100ml. Silicon was determined using the atomic absorption spectrophotometer.

STEP 3. (Specifically adsorbed silicon)

Eight mls (8 ml) of 1M sodium acetate was added to the residue from the exchangeable phase and equal to pH 5.0 with acetic acid and was agitated in a mechanical shaker for 5 hours, filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

STEP 4. (Oxide bound silicon)

Forty mls (40 ml) of hydroxylamine hydrochloride (0.5M $\text{OHNH}_2\cdot\text{HCl}$) was added to the residue from the specifically adsorbed Silicon and shake for 6hours and occasionally agitated , filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

STEP 5. (Organic bound silicon)

Three mls (3ml) of 0.02 M HNO_3 and 5ml of 30% H_2O_2 (pH 2.0) were added to the residue of oxide bound and heated for 2 hours, and then 3ml of 30% H_2O_2 was added and heated again for another 3 hours, The sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

STEP 6. (Residual silicon)

The residue from the organic bound was digested with HF-HClO_4 until metals were dissolved and the sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

STEP 7. (Total silicon)

One gram (1 g) of soil sample was weighed, 15 ml of concentrated nitric acid (HNO_3) was added into a 250ml flask, the mixture allowed to stand for 1 hour. 5 ml of concentrated perchloric acid (HClO_4) was carefully added and heated until solution became clear and reduced to about 5 ml. 5 ml of 2M HCl was added, the sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

2.2.1 Physico-chemical analysis

Soil pH (1:1) soil:water [15]. The sand, silt and clay fraction distribution [16] as modified by [17]. Organic carbon via wet oxidation methods [18]. The exchangeable cations acidity was

determined by titration method as reported by [17]. Exchangeable cations via ammonium acetate solution (1N NH_4OAc) buffered pH 7.0. Ca and Mg were determined from the extract of 0.01M EDTA (ethylenediaminetetra-acetic acid) titration method as described by [17] , while K and Na were determined using photometer [19]. Total nitrogen and available phosphorous was determined by [20]; [21].

2.2.2 Single acid extraction method (0.1M HCl)

Five grams (5 g) of soil was weighed into 250 ml flask , and 0.1 M HCl was added and shaken for 30 minutes. The sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

2.2.3 Double acid extraction method (0.05 HCl IN 0.125 H_2SO_4)

Five gram (5 g) soil was weighed into a 250 ml flask; 25 ml of the extract reagent (0.05 HCl + 0.125M H_2SO_4) were added and shaken for 15 minutes. The sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer [15].

2.2.4 DTPA-TEA Extraction

Ten gram (10 g) of soil was weighed, 20 ml of diethylene triamine penta-acid (DTPA) and triethanol amine (TEA) mixture was added the sample was filtered to 100ml mark. Silicon was determined using the atomic absorption spectrophotometer.

2.3 Statistical Analysis

The data obtained were analyzed by Genstat computer package. The mean significant differences were separated at 5% level, while the relationship between some soil properties vs Silicon was shown.

3. RESULTS AND DISCUSSION

3.1 Determination of Si

Si was extracted with various extractants, the amounts of the metals removed were lowest from the single acid of extraction, and increased with subsequent chelates (Table 2). HCl, HCl + H_2SO_4 , and DTPA were found to be the most effective extractants for determining Si respectively. The single and DTPA extraction methods were higher for Si significantly revealing

that the successive extraction is particularly effective for determining Si, as related to pH of the solution.

In addition, three successive extractions increased the pH of the mixture. Among the three methods, DTPA formed higher medium (pH = 7.3) and hence removed larger quantities of Si from the soil than the other extractants, vs depth (Table 2).

The DTPA (pH = 7.3) was higher than that of mixtures treated with HCl and HCl +H₂SO₄. Therefore, DTPA removed higher quantities of silicon than the other two extractants. Ion exchange play an important mechanism for the removal of metals by soil chelates, our study show that acidic dissolution, especially the extractant-soil mixtures with acids or acidic extractants, may be a good alternative to ion exchange [22].

3.1.1 Si in soil

Si distribution fractions varied across the seven fractions via the soils depth. Silicon was mainly distributed in residual (F6) fractions, this show higher values of total Si, bound to the residual fractions, revealing that the total Si controls the available pool of silicon for plant uptake and its composition, this is in consonant with an experimental results for oxyanions (W), in the soils-plant system [23], total W controls the bioavailable pool, and a peak value of 93% of total W, bound to the residual fractions, implying that apart from minerals nutrients, this mechanism is similarly associated with metals respectively. The mechanism of Si vs parent materials across the soil depth generally, B>D>C>A, In addition, the forms of Si in the parent materials decreased in the pattern RES, bound residual fractions > EXC, soluble & exchangeable fractions > OM, organic matter fraction. Fractions of distribution is very important in assessing the efficiency of metal extraction through chemical chelates, as a result each fraction varies in the metal that is primarily bound [24]. Exchangeable Si, and Si bound to organic matter, and Fe-Mn oxides are removed by one of the following mechanisms—replacement by H⁺, formation of complexes with DTPA and acidic dissolution.

3.1.2 Forms of silicon vs soil parameters

Organic carbon, sand, silt, calcium, magnesium, potassium, and phosphorus were positively and

significantly correlated with all the forms of Silicon, while clay negatively and significantly correlated with all the forms of Silicon. Also in Soil B (Table 4) the clay content of the soil negatively and significantly correlated with all the forms of silicon, whereas the organic carbon, sand, silt, calcium, magnesium and phosphorus properties of the soils positively and significantly correlated with all the forms of Silicon. Similarly, in soil C the soil pH, organic carbon, sand, silt calcium, magnesium, potassium and phosphorus components positively and significantly corrected with all forms of Silicon while the clay content negatively and significantly corrected with all the forms of Silicon. Also, in Soil D the soil pH positively and significantly correlated with Soluble Silicon, Exchangeable Silicon, Carbonate Silicon, Iron oxide Silicon and Total Silicon. While the potassium content negatively and significantly correlated with Carbonate Silicon, Iron oxide Silicon and Total Silicon. The organic carbon, sand, silt, calcium, and magnesium positively correlated with all the forms of Silicon. The potassium content of the soil was also positively and significantly correlated with soluble Silicon and exchangeable Silicon.

3.2 Discussion

The increase in clay fraction with soil depth can be ascribed to clay illuviation into the subsoil thus suggesting pedogenic processes of clay lessivage which are wide spread occurrence in inter-tropical soils [25].

The soils pH were moderately acidic. The acid nature of the soils could be ascribed to erosion and leaching of exchangeable bases which is characterized by extreme rainfall. These effects however affect soil nutrient as most basic cations are removed by leaching thereby leaving behind Al³⁺ and H⁺ that can cause soil acidity. For instance, Al³⁺ causes acidity by hydrolysis reaction while H⁺ is by direct dissociation [26]. Brady et al., 2007, reported that most micronutrients including Si is more available at low pH, in that they become soluble. In other words, at slight acid levels of soil, micronutrients are more soluble and their availability increase. The high organic matter content as observed in the study might be due to any of these factors: mineralization of organic matter and reduced burning, erosion, thick vegetation cover, cultivation with adequate return of crop residue [27]. Nitrogen a nutrient element has direct relationship with the organic matter which

supplies over 80% of soil nitrogen. This implied that low organic matter content will invariably affect the nitrogen content of that soil. The result of this study showed that nitrogen content varied among locations. The top 0 - 60 cm depth of koko, sobe and uhomora had high Nitrogen above the critical level reported by 1.5gkg^{-1} [28]. Phosphorus is another major nutrient apart from nitrogen that is required by all plants irrespective of the age of the plant [29]. The P content of the top soils 0 – 30 cm depth varied among location was above critical level of 8 g/kg^{-1} [30], which depicts that P content of the soils were generally high. The high content of P as observed in the study could be attributed to the high organic matter content. These findings were in agreement with [31], in their studies on P distribution and forms in Nigeria basement complex. The high P content may also be due to the low sorption capacity (fixation arising from high Fe and Al oxides) [32]. The exchangeable bases which comprise Ca, Mg, K and Na which varied among location were outwardly low; $<5.0\text{ cmol kg}^{-1}$ for Ca, $<1.5\text{ cmol kg}^{-1}$ for Mg, $<18\text{ cmol kg}^{-1}$ for K and $<0.24\text{ cmol kg}^{-1}$ for Na [30]. The low content of the basic cation could be as a result of intensive weathering of the environment, leaching which cause losses of colloidal material and low activity clay which were mostly the 1:1 clay mineralogy [27]. In a related study of the same ecological zone [33] reported same low content of basic cations and this low basic cations was attributed to the kaolinitic nature of the soil and low content of clay.

The values for the single acid extractable Silicon obtained in this study varied with location and depths and were generally above the critical level of 50.0 mg kg^{-1} as reported by [33, 34], below which Silicon becomes deficient in soil. The degree of the extraction was more favorably disposed in alluvium soils compared to Shales, Basement complex and Coaster plain sand for Silicon, this may be due to the nature of the parent material.

The specifically adsorbed Silicon varied with depth and across locations and was higher at the upper depth, which is far below the critical level 50.0 mg kg^{-1} and 100.0 mg kg^{-1} as reported by [33,34], The oxides bound which is considered as Silicon occurred in Al and Fe oxides and hydroxide varied with soil depths and across the locations and was higher at the upper layer than the lower depth 50.0 mg kg^{-1} as reported.

Silicon exchangeable varied with soil depths and across the various locations and tends to be

higher at the upper soil depths. This fraction is believed to be part of the most readily available Silicon for plant uptake [35], mainly weakly absorbed by soil components. The lower amount of this fraction of Silicon may be refer to leachable [36].

The residual Silicon fraction constituted the highest proportion of the total Silicon in the soil, they are resistant and may not solubilize under natural condition [37], and falls within the range of 50.0 mg kg^{-1} as reported by [33,34], This fraction is however not related to Silicon available for plant uptake. The range obtained is comparable with that of [38], which ranged from 50.0 mg kg^{-1} and 100.0 mg kg^{-1} as reported by [33,34], and also similar to the findings of [39], (2.03 to 170.85 mgkg^{-1}) in the upper depth. Total Silicon in the soil are relatively high which indicates that Silicon exists in soils largely in unavailable form, and was generally distributed in the soil, this may suggest long term sustainability of crops grown in these soils. The higher content of Silicon in the surface of the soil maybe due to the associations of Silicon with organic matter. The results of the study showed that sand and silt fraction of the soil significantly and positively correlated with the forms of Silicon except for few forms across locations. This might be due to the fact that sandy soils enhances leaching of soluble cation that lead to acidity, with more cation displaced, Silicon gets adsorbed into the exchange complex and thus become more available at such acidic medium. Attractively, clay as a soil fraction which is known to have very strong affinity with micronutrients and is one of the fixation agents responsible for immobilization [26], and was found to have a negative but significant relationship. This was in agreement with the findings of [40], who indicated that fractions of readily available form of Silicon are more associated with the sand than clay and silt fraction. This align with the work of [41 & 42] who reported positive correlation of clay with some extractable Silicon. These differences reported could be attributed to the adoption of varied practices, mechanical implement, different soil management, cropping system and the nature of the parent material [25 & 26].

Soil pH were positively and significantly correlated with forms of Silicon in basement complex except in soluble Silicon which had a negative and non-significant correlation, while other parent materials were positively but not significantly correlated, these positive correlation

Table 1. Major soil physico-chemical properties

Soil ID	pH	OM ^a (gkg ⁻¹)	Total Si (mgkg ⁻¹)	Total Ni (mgkg ⁻¹)	Available P (mgkg ⁻¹)	Ca (cmol(+)kg ⁻¹)	Na (cmol(+)kg ⁻¹)	Mg (cmol(+)kg ⁻¹)	K (cmol(+)kg ⁻¹)
Soil A	5.42	32.41	31.93	23.63	8.51	0.66	0.15	0.24	0.28
Soil B	5.72	26.41	61.9	2.84	8.56	0.42	0.23	0.28	1.23
Soil C	6.49	25.81	50.17	26.24	6.43	1.92	0.66	0.24	0.32
Soil D	6.04	33.88	58.11	16.56	3.63	1.18	0.15	0.27	0.60

^a Organic matter**Table 2. BCR speciation of silicon**

Location/parent material	Depth cm	Sol.Si	Ex.Si	Co ₃ Si	Fe oxide Si	Org. Bond Si mg/kg	Res. Si	Total Si
A	0-30	0.23a	0.29a	0.20a	0.24a	0.34a	33.62a	35.80a
	30-60	0.20a	0.26a	0.16a	0.21b	0.31a	31.52b	33.30b
	60-90	0.17b	0.22b	0.15b	0.20bc	0.28b	29.93c	31.48c
	90-120	0.16b	0.20b	0.13b	0.17c	0.26b	27.75d	29.15d
B	0-30	0.21a	0.44a	0.16a	0.24a	0.35a	68.38a	70.58a
	30-60	0.18b	0.39a	0.14a	0.19b	0.33a	59.30b	64.45b
	60-90	0.15b	0.34b	0.11b	0.17c	0.31a	57.38b	59.58c
	90-120	0.11c	0.30b	0.09b	0.14d	0.29a	51.40b	53.00d
C	0-30	0.28a	0.76a	0.32a	2.40a	0.48a	55.45a	58.60a
	30-60	0.21b	0.59b	0.27b	0.37a	0.44a	50.52b	53.00b
	60-90	0.16c	0.46c	0.20c	0.31a	0.37b	45.38c	47.33c
	90-120	0.12c	0.30d	0.14d	0.22a	0.31c	40.27d	41.77d
D	0-30	0.24a	0.55a	0.25a	0.30a	0.36a	58.90a	66.42a
	30-60	0.19b	0.48b	0.21b	0.29a	0.33ab	57.75a	61.15b
	60-90	0.15c	0.44c	0.16c	0.25b	0.31b	52.92a	54.95c
	90-120	0.13d	0.40d	0.12d	0.22c	0.27c	48.55a	49.92d

Mean value with same letter(s) are not significantly different at 5% level of probability

Table 3. The amount of Si removed with various extractants

Location/parent material	Depth (cm)	Single Acid	Double Acid mg/kg	DPTA
A	0-30	8.93a	19.65a	28.50a
	30-60	8.28ab	18.40ab	26.00b
	60-90	7.49bc	16.40c	22.68c
	90-120	6.74c	13.47c	19.85d
B	0-30	10.77a	27.02a	50.92a
	30-60	8.76b	23.43a	46.70b
	60-90	7.46bc	18.65b	43.25b
	90-120	5.97c	15.57b	38.73c
C	0-30	10.28a	17.48a	31.55a
	30-60	7.97b	13.68b	25.52b
	60-90	5.19c	10.19c	21.27c
	90-120	3.20d	8.45d	17.95d
D	0-30	8.68a	21.85a	43.12a
	30-60	7.90b	18.82b	37.23b
	60-90	6.81c	14.90c	32.65c
	90-120	6.19c	11.29d	26.40d

Mean value with same letter(s) are not significantly different at 5% level of probability

Table 4. Correlation coefficient between forms of silicon and some properties of soils

		Sol.Si	Ex.Si	Co₃ Si	Fe oxide Si	Org. Bond Si	Total Si
A	pH	0.203	0.215	0.212	0.517	215	0.468
	Org.carbon	0.899*	0.854*	0.941*	0.810*	0.967*	0.932*
	Silt	0.919*	0.866*	0.933*	0.904*	0.948*	0.938*
	Clay	-0.883*	-0.821*	-0.917*	-0.872*	-0.948*	-0.923*
	Ca	0.949*	0.894*	0.858*	0.879*	0.894*	0.934*
	Mg	0.934*	0.917*	0.836*	0.853*	0.843*	0.914*
	K	0.860*	0.935*	0.851*	0.891*	0.892*	0.906*
	P	0.901*	0.949*	0.917*	0.859*	0.953*	0.915*
B	pH	-0.182	-0.194	-0.017	-0.283	-0.178	0.878*
	Org.carbon	0.866*	0.957*	0.866*	0.873*	0.818*	0.933*
	Silt	0.912*	0.976*	0.903*	0.882*	0.895*	0.953*
	Clay	-0.822*	-0.93*	-0.961*	-0.849*	0.911*	-0.916*
	Ca	0.854*	0.956*	0.803*	0.886*	0.748*	0.951*
	Mg	0.837*	0.887*	0.903*	0.908*	0.858*	0.965*
	K	0.860*	0.989*	0.756*	0.897*	0.859*	0.950*
	P	0.852*	0.945*	0.857*	0.896*	0.878*	0.945*
C	pH	0.919*	0.908*	0.916*	0.891*	0.886*	0.844*
	Org.carbon	0.919*	0.896*	0.968*	0.974*	0.978*	0.938*
	Silt	0.932*	0.895*	0.953*	0.945*	0.928*	0.923*
	Clay	-0.944*	-0.923*	-0.984*	-0.964*	-0.945*	-0.942*
	Ca	0.832*	0.799*	0.840*	0.828*	0.850*	0.954*
	Mg	0.679*	0.758*	0.766*	0.765*	0.777*	0.667*
	K	0.779*	0.728*	0.693*	0.699*	0.771*	0.653*
	P	0.942*	0.906*	0.952*	0.949*	0.959*	0.918*
D	pH	0.584*	0.642*	0.497	0.618*	0.286	0.790*
	Org.carbon	0.973*	0.912*	0.931*	0.874*	0.692*	0.926*
	Silt	0.951*	0.947*	0.957*	0.911*	0.706*	0.957*
	Clay	-0.840*	-0.839*	-0.906*	-0.809*	-0.841*	-0.753*
	Ca	0.842*	0.864*	0.797*	0.772*	0.797*	0.920*
	Mg	0.817*	0.801*	0.819*	0.933*	0.673*	0.929*
	K	1.000*	0.899*	-0.776*	-0.800*	-0.508*	-0.782*
	P	0.970*	0.884*	0.908*	0.867*	0.811*	0.949*

was in line while the negative correlation obtained may be due to larger solubility of Silicon in acidity, type of parent material, climatic factors and diverse agronomic practices. The positive correlation obtained between organic matter and forms of Silicon suggests a strong association between extractable Silicon, and organic matter, indicating Silicon being a composite part of organic matter. This result was in line with the studies on micronutrients reported by [43 &44], who reported that the positive correlation between extractable Silicon, and organic matter could be tied to the organic constituent of the soil since the organic constituent of a given soil provide more exchange site for boron adsorption in the soil. The positive relationship between N, P, K, Ca, Mg indicates synergistic relationship of these nutrient elements with Silicon.

4. CONCLUSION

The findings of the study showed that the soil were textually sand and sandy loam, ranged from moderately acidic to slightly alkaline, low in organic matter, total N and available P. The basic cations and ECEC indicated low fertility status of the soil. The extractable Silicon content of the soil varied from one location to another and concentrated more at the surface than the lower soil depth. Generally, the extractable Silicon content decreased with increased soil depth. The soil Silicon level could be said to be below the critical level established method for available Silicon. The extractable Silicon seem to be more in terms of the amount extracted at the top soil across the various locations. This was followed by double acid extraction method, while the least was obtained with single acid extraction method. The residual Silicon had the highest content and this showed that silicon exist largely in the unavailable form, due to the bound fractions in soils. The correlation between extractable Silicon, and soil properties (Sand, Silt, Clay, pH, Organic matter, Nitrogen, Phosphorus, Magnesium, and CEC showed their influence on Silicon availability. Generally, the value of extractable Silicon established in this study showed that they were not within the sufficiency range (50.0 mg kg⁻¹) and may have negative effect on crop production but application of organic fertilizers might trigger Silicon from the deficient level to a more sufficient level in these locations. However, the soil must be maintained and conserved.

ACKNOWLEDGEMENT

We are indebted to the staff of Central Laboratory Faculty of Agriculture University of Benin for analytical assistance.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Chude VO, Malgwi WB, Amapu IY, Ano AO. Manual on soil fertility assessment. Federal fertilizer department. FAO and national programme on food security, Abuja, Nigeria. CRC Press, Boca Raton, FL. 2011;62.
2. Olowokere FA, Dare MO, Olasantan FO, Akanji MA . Evaluation of animal manures on the yield of solanum macrocarpon L. and chemical properties of soil. In: Achieving Social and Economic Development through Ecological and Organic Agricultural Alternatives, Proceedings of the Scientific Track, 3rd African Organic Conference, Lagos, Nigeri. 2015;43-46.
3. Rangel Z. Heavy metals as essentials nutrients. In: Prasad MNV, hagemeyer J (eds); heavy metal stress in plants: Molecules to ecosystems. Springer-Verlag, Berlin, Heidelberg. 2003;271-294.
4. Mortvedt JJ, Johnson DL and Croissant RL. Fertilizing sunflowers. Colorado state comperative extension fact sheet No. 543. Soil and Crop Sciences. 2003;3:96.
5. Epstein E. The anomaly of silicon in plant biology. Proc. Natl. Acad. Sci. USA 91:1117.1; 1994.
6. Keeping MG, Kvedaras OL , Bruton AG. Epidermal silicon in sugarcane: Cultivar differences and role in resistance to sugarcane borer *Eldana saccharina*. Environ. Exp. Bot. 2009;66:54–60.
7. Meyer JH, Keeping MG. The impact of nitrogen and silicon nutrition on the resistance of sugarcane varieties to *Eldana saccharina* (Lepidoptera: Pyralidae). Proc. S. Afr. Sug. Technol. Ass. 2005;79:363–367.
8. Snyder GH, Martichenkov VV, Datnoff LE. Silicone. In: Handbook of plant nutrition. Barker A.V, Pilbean DJ (eds.). CRC Taylor and Francis, New York, USA. 2007;551–568.

9. Berkhout ED, Malan M, Kram T. Better soils for healthier lives? An econometric assessment of the link between soil nutrients and malnutrition in sub-Saharan Africa. *PLoS. ONE*. 2019;14(1):e0210642. Available:https://doi.org/10.1371/journal.pone.0210642
10. Berkhout E, Malan M, Kram T. Micronutrients for agricultural intensification. Is sub-Saharan Africa at risk? The Hague: PBL Netherlands Environmental Assessment Agency; 2017.
11. Cornelis JT, Delvaux B, Georg RB, Lucas Y, Ranger J, Opfergelt S. Tracing the origin of dissolved silicon transferred from various soil-plant systems towards rivers: A review. *Biogeosciences*. 2011;8:89–112.
12. Tsadilas CD, CS Yassoglou, CS Cosmas, CH Kallianou. The availability of soil boron fractions to olive trees and barley and their relationships to soil properties. *Plant Soil*. 1994;162:211-217.
13. Vasanthi ND, Saleena LM, Raj SA. Silicon in day today life. *World Appl Sci J*. 2012;17:1425–1440.
14. Hodson MJ, White PJ, Mead A.. Phylogenetic variation in the silicon composition of plants. *Ann Bot*. 2005;96:1027–1046
15. Udo EJ, Ibia TO, Ogunwale JA, Ano AO Esu IE, Udoh BT, Ndaeya NU, Harold KO. Fertility status and variability of some soils developed from parent material in the humid region of Nigeria. *Proceedings of the 31st Annual Conference of Soil Science of Nigeria (SSSN), 13th – 17th November, 2006, Ahmadu Bello University, Zaria*. 2006;79 – 85.
16. Bouyoucos CJ. Recalibration of the hydrometer method of making the mechanical analysis of soils. *Agronomy Journal*. 1951;43:434–438.
17. Day PR. Hydrometer method of particle size analysis. In: Black CA., Ed, *methods of soil analysis*. American Society of Agronomy, Madison, Wisconsin Argon. 1965;562-563.
18. Walkley A, Black IA. An examination of the different methods for determining soil organic matter and the proposed modification of the chromic acid wet titration method. *Soil Science of America Journal*. 1934;37:29–38.
19. Jackson ML. *Soil chemical analysis*. Prentice Hall, New York. 1962;263–268.
20. Bremner JM, Mulvaney CS. Nitrogen-total. In: *Methods of soil analysis*. Part 2. Chemical and microbiological properties, Page AL, Miller RH, Keeney DR, Eds. American Society of Agronomy, Soil Science Society of America, Madison, Wisconsin. 1982;595-624.
21. Murphy J, Riley JP. A modified single solution method for determination of phosphate in natural waters. *Anal. Chem. Acta*. 1962;42:31–36.
22. Reed B, Carriere P, Moore R. Flushing of Pb (II) contaminated soil using HC1, EDTA and CaC12. *Environ. Eng*. 1996;122:48–50.
23. James B, Wang G. Accumulation characteristics of tungsten (W) and its potential health risk assessment in the soil-vegetable system under field conditions. *J Soils Sediments*. 2020;20:599–608.
24. Barona A; Aranguiz I; Elías A. Metal associations in soils before and after extractive decontamination: Implications for the effectiveness of further clean-up procedures. *Environmental Pollution*. 2001;113:79-85.
25. Amalu UC, Esu IER, Nwulu MVN. Fertility management of some gleyic luvisols and alisols grown to rice in South East Nigeria: Boron, Copper and Zinc status; 2001.
26. Brady AC, Weil RR. *The nature and properties of soils* (13thed.) Pearson Prentice-Hall Education Inc. 2007;976.
27. Egbuchua CN. (2012). *Pedological characterization of some alluvial complex soils in Delta State, Nigeria*. *Nigerian Journal of Soil and Environmental Research*. 2012;9:65–70.
28. Enwenzor WO, Udo ER, Usoroh NJ, Adepetu JA., Claude VA, Udegbe CL. *Fertilizer use and management for crops in Nigeria*. Series. 1979;2:163.
29. Havlin JL, Beaton JD, Tisddate SL, Nelson WL. *Soil fertility and fertilizer*. An introduction to nutrient management. Prentice Hall New Delhi, Indian. 2005;519.
30. FMANR – Federal Ministry of Agriculture and Natural Resources. *Literature review of soil fertility evaluation in Nigeria (in 5 volumes)* Federal Ministry of Agriculture. Lagos. 1996;281.
31. Hassan AM, Raji BA. Phosphorus distribution and forms in Nigeria basement complex rock bauchi. *Proceedings of the 13th – 17th Nov, 2006*. Ahmadu Bello University, Zaria. 2006;506–511.
32. Tsado PA, Agbenin JO, Osunde AO, Bala A. Phosphate sorption in three cultivated soils of Nigeria guinea savanna as

- influenced by the addition of citric acid. Proceedings of the 31st Annual Conference of Soil Science Society of Nigeria (SSSN), 13th – 17th November, 2006. Ahmadu Bello University, Zaria. 2006;177–482.
33. Udoh BT, TO Ibia, BU Udo, SO Edem. Assessment of micronutrient status of inland depression and floodplain (wetland) soils in Akwa Ibom State Southeastern Nigeria; 2008.
 34. Alloway BJ, Heavy metals in soils. Blackie Academic and Professional, Glasgow, Scotland. 1995;368.
 35. Keren R, Bingham FT, Rhoades JD. Plant uptake of boron as affected by boron distribution between liquid and solid phases in soil. Soil Sci. Soc. Am. J. 1985;48:297-302.
 36. Rhoades JD, Ingvaision RD, Hatcher JT. Laboratory determination of leachable soil boron. Soil Science America Journal. 1970;34:871–875.
 37. Tessier A, Campbell PG, Bishop M. Sequential extraction procedure for the specialization of particular trace elements. Analytical Chemistry. 1979;51:844–851.
 38. Mengel, Kirkby EA. Principles of plant nutrition. 4th Edn. Panima Publishing Corporation, New Delhi, India; 1987.
 39. Raji A. Profile distribution of total and available sulphur and boron in sandy soils of Nigerian semi-arid savanna. Nigerian Journal of Soil and Environmental Research.2008;8:42-49.
 40. VOG Chude, Obigbesan O, Egbe NE. Micronutrient status of selected cacao-growing soils in South-Western Nigeria: Profile distribution of total and available Zn. Can. J. Soil Sci. 1985;65:21-24.
 41. Fagbemi A. Soil of Bendel state. Federal department of agriculture, final report of Dr. Fagbemi. Department of Agronomy, University of Ibadan.1985;1–77.
 42. Aghimien AE. Micronutrient status of hydromorphic soils for southern Nigeria. Nigeria Journals for Palms and Oil Seed. 1985;9:10–36.1985
 43. Katyal JC, Sharma BD. DTPA-extractable and total Zn, Cu, Mn and Fe in Indian soils and their association with some soil properties. Geoderma. 1991;49:165-179.
 44. Elrashidi MA, O'Connor GA. Boron sorption and desorption in soils. Soil Science Society of America Journal. 1982;46:27–31.

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