



Comparative Study of Removal of Cadmium Ions from Aqueous Solution Using *Dialium indum* and *Santalum album* Fruitshells as Adsorbents

N. A. Jibrin^{1*}, J. Yisa², J. O. Okafor³ and A. Mann²

¹Department of Petroleum Chemistry, Baze University, Abuja, Nigeria.

²Department of Chemistry, Federal University of Technology, Minna, Nigeria.

³Department of Chemical Engineering, Federal University of Technology, Minna, Nigeria.

Authors' contributions

This work was carried out in collaboration between all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/ACSJ/2016/21587

Editor(s):

(1) Gustaaf Schoukens, Department of Textiles, Ghent University, Belgium.

Reviewers:

(1) Jamidu H. Y. Katima, University of Dar es Salaam, Tanzania.

(2) Anonymous, Manipal University, India.

(3) Eduardo Alberto López-Maldonado, Center for Research and Technological Development in Electrochemistry, CIDETEQ, Mexico.

(4) Bahia Meroufel, Bechar University, Algeria.

(5) Ahmed H. A. Dabwan, TATI University College, Malaysia.

Complete Peer review History: <http://sciencedomain.org/review-history/12370>

Original Research Article

Received 25th August 2015
Accepted 26th October 2015
Published 21st November 2015

ABSTRACT

Sandal and velvet tamarind fruit shells were screened for their metal uptake capacities for Cd(II) ions, at various initial concentrations of 10, 20, 30, 40 and 50 mg/L at variable contact period of between 30 and 180 minutes and different adsorbent dosages of 0.2 – 1 g/l, pH(1 to 6) using batch adsorption mode. The adsorbents were characterized using standard methods. The amount of adsorbate adsorbed by the developed adsorbents for Cd²⁺ increased with increase in initial metal concentration, contact time, adsorbent dosage and pH until equilibrium point was reached. At equilibrium; the optimum contact time for this operation was 90 minutes for velvet tamarind and 120 minutes for sandal fruit, effective pH for both adsorbents is 5.5, the equilibrium initial metal concentrations are 42 and 41 mg/l for sandal fruit and velvet tamarind shell respectively. The data conformed to Freundlich isotherm with $r^2 > 0.95$ for both adsorbents and were also fit for second-order kinetics. The surface areas of the sandal fruit and velvet tamarind shell as determined were

*Corresponding author: E-mail: noakoj@yahoo.com;

615 and 570 m²/g. The results obtained from this study indicated a high adsorption ability of the adsorbents for Cd(II) with sandal fruit having better performance.

Keywords: Adsorption; removal; *Santalum album*; *dialium indum*; cadmium; modeling; characterization.

1. INTRODUCTION

The recent advances in industrial and technological advancements at the global level have brought about the introduction of different pollutants into water bodies. This tremendous increase in the use of heavy metals over the past few decades has inevitably resulted in an increased flux of metallic substances in the aquatic environment [1]. These metals are of special concern because of their persistency and health impacts. Industrial waste constitutes a major source of various kinds of metal pollution in natural water [2]. These toxic metals of interest include; Cd, Zn, Pb and Ni. These toxic heavy metals enter into the water bodies through waste waters from metal plating industries and industries of Cd Ni batteries, phosphate fertilizers, mining activities, pigments and stabilizer alloys etc Cd²⁺, Zn²⁺, Ni²⁺ and Pb²⁺ ions exist in the soil-water system as micronutrients for plants and animals' use. According to Dermibas [3], Cd²⁺ ion poisoning in humans are very serious, among which high blood pressures, kidney damage and destruction of testicular tissues and red blood vessels are prevalent [4]. US EPA standards for cadmium in drinking water is 0.005 mg/l.

A number of technologies have been developed over the years to remove toxic metals from water. The most important technologies include chemical precipitations, electro flotation, ion exchange, reverse osmosis, and adsorption on commercial activated carbon [5]. These methods are not cost effective and have prompted a search for cheaper substitutes. Low cost and non conventional adsorbents include the use of agricultural wastes like natural compost, Irish peanut, peanut shell, coconut shell, bone char and biomass such as *Aspergillus tereus* and *Mucus remanianus*, polymerized onion skin with formaldehyde and EDTA, modified cellulosic material, natural materials such as hair and cat tails (Thypha Plant) [6], waste wool [7], peanut skin [8], modified barks [9], barely straw [10], low rank coal [11], waste tyre rubber and tea leaves [12]. The present investigation attempts have been made to assess and compare the efficiency of low cost adsorbent materials prepared from

sandal and velvet tamarind fruit shells for removal of Cd(II) from wastewater. The influence of various factors, such as adsorbent loadings, initial pH, temperature and initial metal ion concentration on the adsorption capacity were studied. The Langmuir and Freundlich models were used to analyze the adsorption equilibrium.

2. MATERIALS AND METHODS

2.1 Sample Collection and Preparation

The sandal and velvet tamarind fruit shells biomasses were collected from agricultural wastes which are present in abundance for wastewater treatment and washed several times with distilled water, and then dried in an oven at 80°C for 24 h. The dried biomasses were ground with a mortar and pestle. The particles were separated by using a US standard testing sieve (No. 100). The material after sieving were soaked for twenty four hours in a solution prepared from sulphuric acid, then placed in a crucible and positioned at the center of a muffle furnace for 15 mins at 500°C, to produce the activated carbon (AC) which is cooled in desiccators [13].

2.2 Preparation of Adsorbate Stock Solution

Standard Cadmium stock solution (1000 mg/L) was prepared by dissolving 2.10 g of Cd(NO₃)₂ in deionized water then diluted to 1000 ml. pH adjustment of solution was made with dilute NaOH and HCl solutions. The reagent was of analytical grade and stored in polypropylene containers. Plastics were cleaned in dilute HNO₃ (10% v/v) and dried at 60°C after rinsing with deionized water.

2.3 Characterization of Adsorbents

The iodine number was determined based on [14] by using the sodium thiosulphate volumetric method. The specific surface area of the activated carbon was estimated using Sear's method [15,16]. Bulk density and Particle density were determined using method used by ([17]. Porosity was determined from the values of bulk density and particles density. Functional groups

presence in the adsorbents was determined by a Fourier-transform infrared spectrophotometer (FTIR; Shimadzu 8400s). The X-ray diffraction (XRD) of the adsorbents was determined in a Randi-con MD 10 model, while scanning electron microscopy (SEM; Hitachi S4800) was used to assess the morphology of the adsorbents.

2.4 Batch Adsorption Studies

The experiments were carried out in a batch mode for the measurements of adsorption capacities, and to generate adsorption kinetics [18]. The effect of pH (3, 4, 5, 6, 7 and 8), contact time (30-180 mins), adsorbent dose (0.2-1g/l) and initial metal ion concentration (10-50mg/l) were determined at room temperature using stopper bottles. The initial pH of solution was adjusted by using 0.05 M HCl or 0.05 M NaOH without changing the volume of the sample. After agitating the sample for the required contact time, the contents were centrifuged and filtered through Whatman No.41 filter paper and unreacted cadmium in the filtrate was analyzed by atomic absorption spectrophotometer.

The adsorption capacity, q_e , was calculated as [19].

$$q_e = \frac{(C_0 - C_e)}{m} V \quad (1)$$

Where C_0 and C_e are the initial and equilibrium adsorbate concentrations in solution (mg/L), respectively, V is a known volume of synthetic wastewater (L), and m is a known mass of dry adsorbent (g).

2.5 Adsorption Isotherms

2.5.1 Freundlich adsorption isotherm

Freundlich Isotherm is represented by the following equation [20]:

$$q_e = K_F C_e^{1/n} \quad (2)$$

Where;

q_e = the amount of adsorbate adsorbed per unit mass of adsorbent (mg adsorbate/g adsorbent)

K_f = adsorption capacity

n = adsorption intensity are the empirical constants.

C_e = equilibrium concentration of adsorbate (mg/l)

2.5.2 Langmuir adsorption isotherm

Langmuir Isotherm is represented by the following equation [21]:

$$q_e = \frac{q_{\max} b C_e}{1 + b C_e} \quad (3)$$

q_{\max} and b are Langmuir constants related to adsorption capacity (maximum specific uptake corresponding to the site saturation) and energy (intensity) of adsorption (L of adsorbent/mg of adsorbate) respectively.

2.5.3 Adsorption kinetics of Cd(II)

To describe the adsorption kinetics, the linear form of the pseudo-first order (Lagergren) equation reported in the literature was applied in the following form [22]:

$$\log(q_e - q_t) = \log q_e - \left(\frac{K_1 t}{2.303}\right) \quad (4)$$

where K_1 (min^{-1}) is the rate constant, and q_t and q_e are the amounts of metal ion adsorbed at time t and equilibrium, respectively.

To describe the adsorption kinetics, a pseudo-second order rate model reported in the literature was also applied in the following form [23]:

$$\frac{t}{q_t} = \frac{1}{h_0} + \frac{1}{q_e} t \quad (5)$$

Where,

h_0 = the initial adsorption rate (mg/g min)

q_e = the amount of metal ion adsorbed at equilibrium (mg/g)

q_t = the amount adsorbed at time t (mg/g)

The initial adsorption rate, h_0 , as $t' \rightarrow 0$ is defined as:

$$h_0 = K_2 q_e^2 \quad (6)$$

Where, K_2 is the pseudo second order rate constant for the adsorption process (g/mg min). The initial adsorption rate h_0 , the equilibrium adsorption capacity, and the rate constant K_2 were determined from the slope and intercept of the plot of t/q_t against t .

3. RESULTS AND DISCUSSION

The adsorbents characteristics were determined and the results are summarized, on Table 1.

Table 1. Physicochemical properties of adsorbents

Parameter	Adsorbents	
	SF	VT
Bulk density (g/cm ³)	0.92	0.151
Ash (%)	4.45	7.2
Iodine number (mg/g)	649.5	614.7
Surface area (m ² /g)	615	570
Particle density (g/cm ³)	1.25	0.72
Porosity (%)	26.4	79

3.1 XRD and SEM Analysis

The Figs. 1 and 2 represent the SEM images and XRD spectra showing the surface morphologies and XRD spectra peaks of Velvet tamarind and sandal fruit.

The surface morphology of the velvet tamarind and sandal fruit examined by SEM as shown in clearly revealed a fragmented and porous surface texture, which indicated the adsorbent was highly porous, with a high adsorption capacity [24]. The porous nature of adsorbent signifies sorption potential for metal ions, such as physical adsorption will play a major role in over all sorption process. The specific surface area of the adsorbents of 615 m²/g for sandal fruit and 570 m²/g for velvet tamarind further confirmed their porous nature. According to [25], an adsorbent with a surface area of 144 m²/g and above can be classified as mesoporous. The XRD spectra of Sandal fruit and Velvet tamarind are shown in Fig. 3. XRD gave useful information about crystalline and amorphous changes of an adsorbent. The presence of sharp and weak peaks suggests an amorphous adsorbent [26] and the amorphous nature of the adsorbents indicates that Cd(II) ions in solution can easily penetrate its surface for efficient adsorption [27].

3.2 FTIR Analysis

Figs. 5, 6 shows the FTIR spectral of sandal fruit and velvet tamarind adsorbents.

The FTIR spectral of adsorbents sandal fruit and velvet tamarind were used to determine the vibration frequency changes in the functional groups on the surface which facilitates the adsorption of metal ions. The spectra of adsorbents were measured within the range of 400 – 4000 cm⁻¹ wave number. The FTIR analysis results (Figs. 5 and 6) suggested the presence of such functional groups as the carboxylic acid or alcoholic O-H bond stretching

which may overlap with amine (N-H) bond stretching at peaks between 3250-3400 cm⁻¹; possible C=O bond of carbonyl or amide groups within 1640-1670 cm⁻¹; C-O and O-H bond stretching of alcohol and ethers at 1000-1260 cm⁻¹ of the finger-print region [28]. The FTIR spectroscopic analysis indicated broad bands at 3412 cm⁻¹, representing bonded –OH groups [29].

3.3 Effect of Contact Time

In adsorption system, the contact time plays a vital role irrespective of the other experimental parameters, affecting the adsorption kinetics. The effect of contact time on the adsorption of Cd(II) ions onto velvet tamarind and sandal fruit is shown in Fig. 7.

An initial increase in amount adsorbed with increased contact time was observed, and adsorption became fairly stable with time. Equilibrium removal was achieved around 90 min for velvet tamarind and 120 min for sandal fruit, after which further increases in contact time did not result in significant adsorption. These times were used in all experiments to ensure maximum removal. The rapid adsorption during the initial stages might be due to the presence of abundant active sites on the surface of both adsorbents which become saturated with time. Initially, sorption is controlled mainly by diffusion from the bulk to the surface of the adsorbents, whereas, later, sorption is probably an attachment-controlled process due to the presence of fewer active sites [30]. The adsorption began to slow down due to slow movement of Cd ions into the interior bulk of the biosorbent. Considering the effective adsorption time of 90 minutes for velvet tamarind which is lower than 120 minutes for sandal fruit, it can be said that it has less active sites which become saturated earlier than sandal fruit. Velvet tamarind has a surface area of 570 m²/g and sandal fruit has the surface of 670 m²/g which correspond to the rate of metal uptake by each of them. The trend observed for the metal ions in the two substrates under the same experimental conditions revealed that the pattern of adsorption is a function of the substrate [31].

3.4 Effect of PH

The effects of initial pH on adsorption process are presented at the Fig. 8. Importance of pH on heavy metal adsorption tests were undertaken with different initial pH values of the metal solutions, using a constant amount of sandal fruit and velvet tamarind adsorbents.

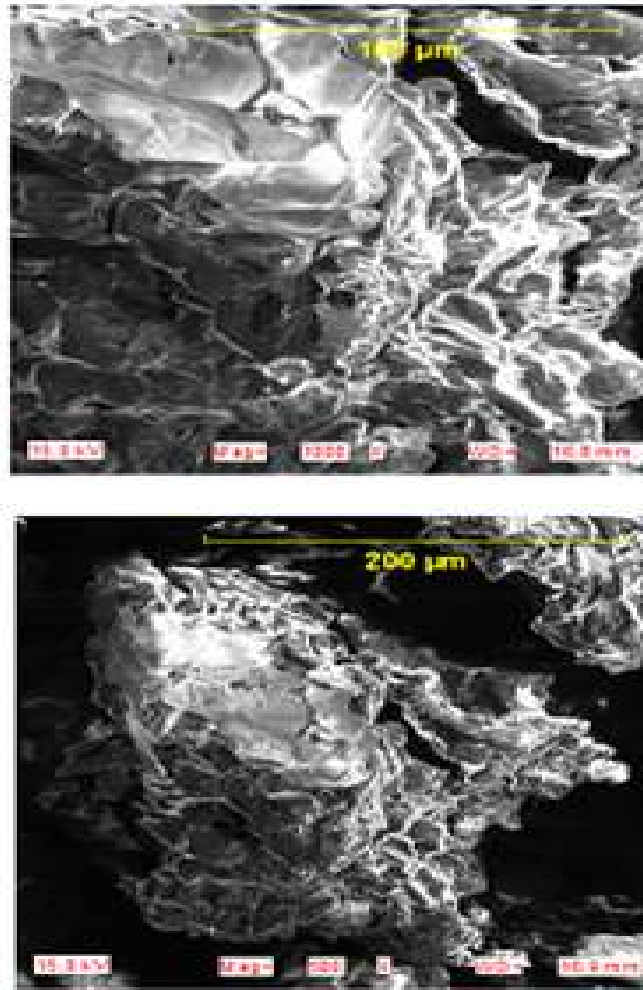


Fig. 1. Scanning electron microscopy (SEM) images of sandal fruit

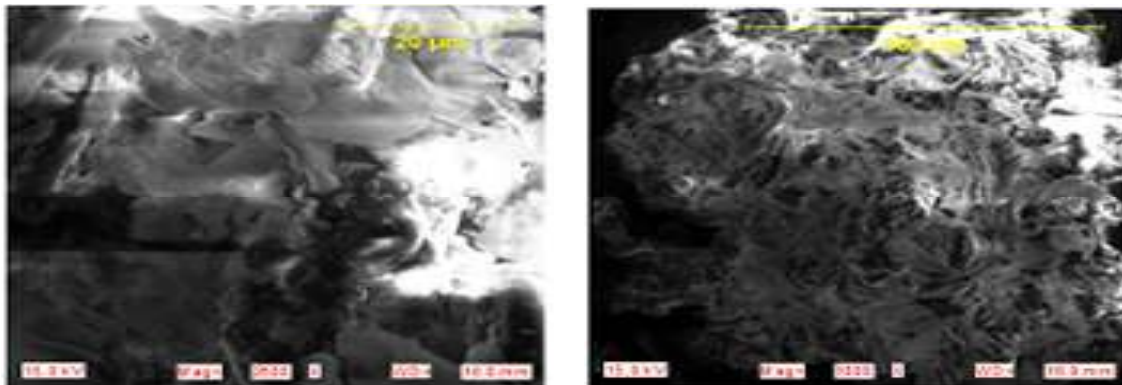


Fig. 2. Scanning electron microscopy (SEM) images of Velvet tamarind at 20 and 100 μm respectively

The adsorptive capacity of velvet tamarind and sandal fruit were dependent on pH of cadmium solution as the amount adsorbed varied with pH. At lower pH values, large number of H⁺ ions

neutralize the negatively charged adsorbent surfaces, thereby creating hindrance to the Cd(II) ions adsorption. At the high pH values, the reduction in adsorption may be due to the abundance of OH⁻ ions causing increased hindrance to diffusion of Cd (II) ions [32]. The adsorbents showed gradual increase in adsorption as the pH increases from 1 up to about 5.5. Both adsorbents performed optimally at pH of 5.5 and decreases with further increase in pH as shown in Fig. 10. The dependence of metal uptake on pH is related to both the surface functional groups present on the biomass and the metal chemistry in solution [33]. At low pH, the surface ligands are closely associated with the hydronium ions (H₃O⁺) and restricted the approach of metal cations as a result of the repulsive force [34,35]. Furthermore, the effects of pH on the metal ions uptake by biomasses can also be justified by the association-dissociation of certain functional groups, such as the carboxyl and hydroxyl groups present on the biomasses [36]. In fact, it is known that at low pH, most of the carboxylic groups is not dissociated and cannot bind the metal ions in solution, although they take part in complexation reactions [37]. The results obtained are in close agreement with previously reported studies [38,39].

3.5 Effect of Adsorbent Dose

Adsorbent dosage is a useful parameter for determining the capacity of an adsorbent for a given initial adsorbate concentration. The effect of modified velvet tamarind and sandal fruit dose on adsorption of Cd(II) ions from solution is shown in Fig. 9.

Adsorbent dose was varied between 0.2-1.2 g/l for adsorbents used. It is clearly seen that the amount of Cd(II) ions adsorbed increases as the adsorbent mass increases mainly due to an increase in the number of available active sites [40,41]. The adsorption of Cd (II) increases at a faster rate initially with increases in adsorbent dose. The result showed that further increase in the adsorbents dose could lead to increase in adsorption until the effective adsorption of 6.1 and 4.64 mg/g is attained for velvet tamarind and sandal fruit respectively. Increasing the dose of modified velvet tamarind from 1 to 1.2 g decreased the equilibrium adsorption capacity from 6.1 to 5.81 mg/g for Cd(II) and increasing

the dose of modified sandal fruit from 1 to 1.2 g decreased the equilibrium adsorption capacity from 4.64 to 4.42. This may be due to a decrease in the total adsorption surface area available, resulting from overlapping or aggregation of adsorption sites [42] or a higher adsorbent dose may provide more active adsorption sites, so that adsorption sites remain unsaturated during adsorption [43].

3.6 Effect of Initial Metal Ion Concentration

The amount of metal ions adsorbed is a function of the initial concentration of the adsorbate (metal ion), making it an important factor in effective adsorption. The effect of initial metal ion concentration (10 to 50 mg/l) on the adsorption of Cd(II) ions onto modified velvet tamarind and sandal fruit is shown in Fig. 10.

It shows that, as the concentration of Cd(II) in solution increases, the amount of Cd(II) adsorbed by the adsorbents increases. The amount of metal ions adsorbed by both adsorbents increased with the metal concentration, as the increasing concentration gradient overcomes the resistance to mass transfer of metal ions between the aqueous phase and the adsorbent [44]. A higher concentration in a solution implies a higher concentration of metal ion to be fixed on the surface of the adsorbent [45]. Modified sandal fruit had a higher adsorption capacity for metal ions than modified velvet tamarind as the initial ions concentration increases. At a concentration of 40 mg/L, 3.04 mg/g of Cd(II) ions were adsorbed by sandal fruit while velvet tamarind adsorbed 2.83 mg/g Cd(II). The higher adsorption by sandal fruit may be due to its larger surface area of 615 m²/g as compared with 570 m²/g for velvet tamarind. The equilibrium concentrations of ions during adsorption are 42 and 41 mg/l for sandal fruit and velvet tamarind respectively which are very close. The decrease in adsorption after that might be due to the agglomeration/ aggregation of the adsorbent particles at a higher concentration. Such aggregation can lead to decrease in the total surface area of the adsorbent available for adsorption and increase in diffusional path length [46]. Similar results were reported by many authors using different types of adsorbents [47,48].

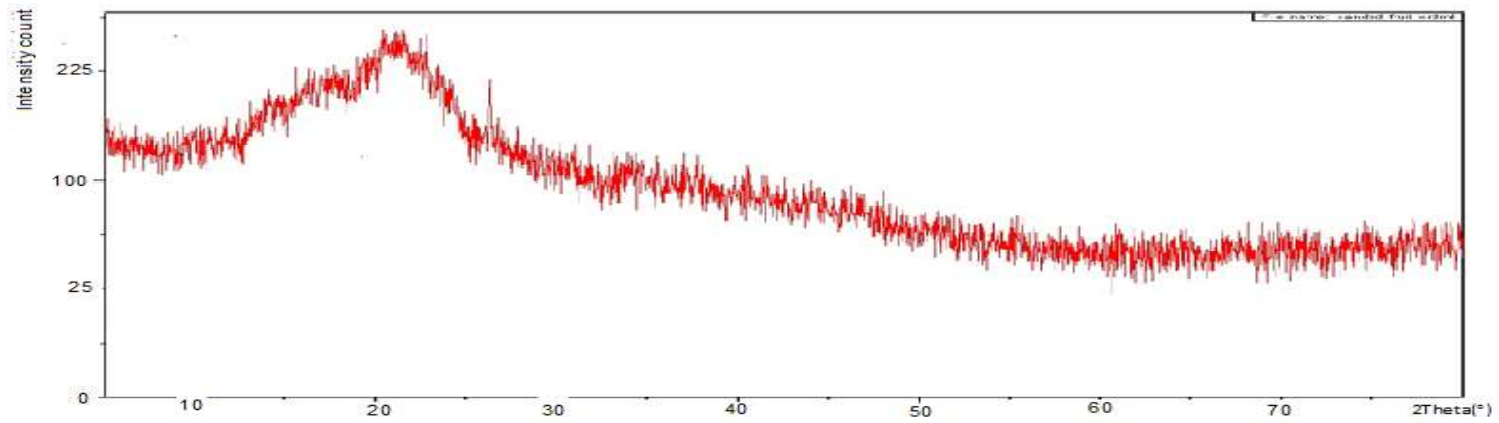


Fig. 3. X-ray diffraction spectrum of sandal fruit biomass

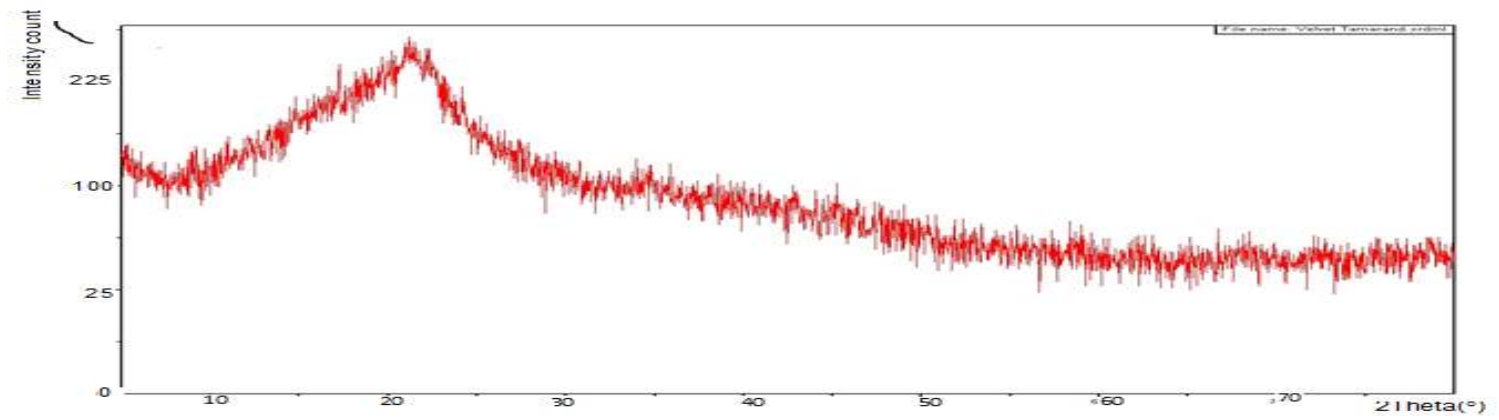


Fig. 4. X-ray diffraction spectrum of velvet tamarind biomass

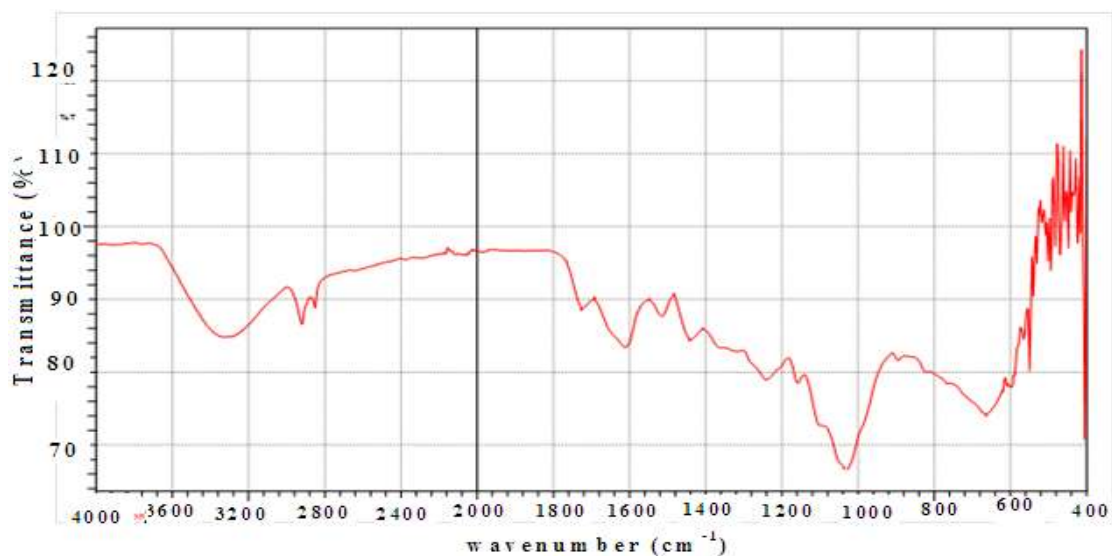


Fig. 5. Fourier transforms infrared (FTIR) spectra of Sandal fruit

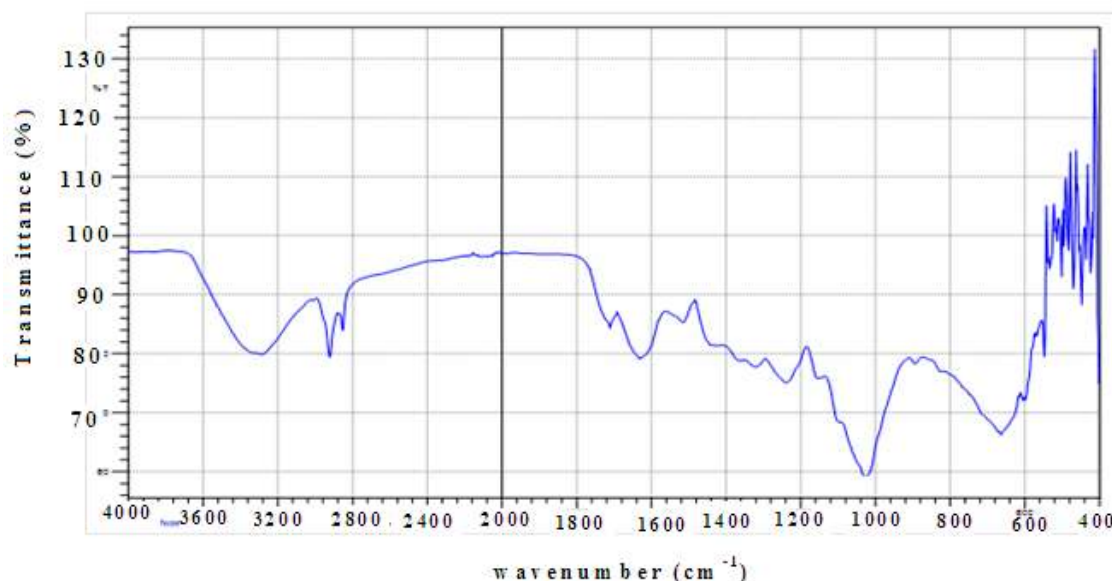


Fig. 6. Fourier transforms infrared (FTIR) spectra of velvet tamarind

3.7 Equilibrium Isotherm Modeling

Equilibrium adsorption isotherms are used to relate the adsorbate concentration in solution and the amount on the adsorbent at equilibrium [49]. These parameters often provide fundamental information on the sorption mechanism, surface properties and the affinity of adsorbents, which helps to determine the applicability of sorption as a unit operation. Therefore, it is important to establish the most suitable correlation of equilibrium curves in order

to optimize the conditions for designing adsorption systems. The most frequently used isotherms, the Langmuir and Freundlich models, were therefore used to analyse the data (Table 2). The Langmuir isotherm is used to describe monolayer adsorption onto the surface of an adsorbent with a finite number of identical adsorption sites and no interaction between sites. The model is expressed as [50]:

$$\frac{C_e}{q_e} = \frac{1}{q_{maxb}} + \frac{C_e}{q_{max}} \quad (6)$$

Where q_{max} (mg/g) is the monolayer adsorption capacity of the adsorbent, and b (L/mg) is the adsorption constant, which reflects the affinity between the adsorbent and adsorbate. Q_{max} and b were determined from the slope and intercept of the plots of C_e/q_e versus C_e . Table 2 shows low correlation coefficients (r^2) for Cd(II)<0.95,

indicating that the adsorption does not follow the Langmuir isotherm closely. The Freundlich isotherm is based on the assumption that sorption takes place on a heterogeneous adsorbent surface, where the sorption energy distribution decreases exponentially and can be expressed as [51]:

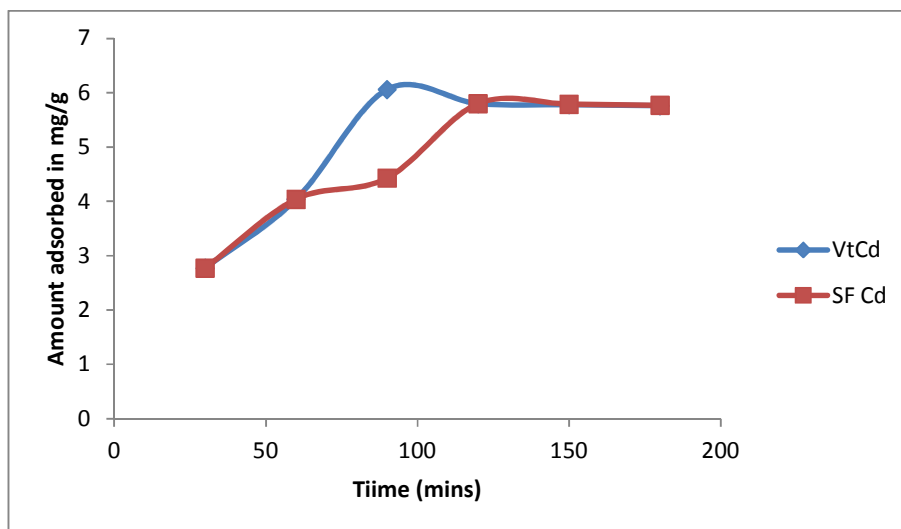


Fig. 7. Plot of amount of Cd(II) ions adsorbed versus Contact Time by Velvet tamarind and Sandal fruit (Temp. $\pm 2^\circ\text{C}$; pH.5.5, Agitation 200 rpm , Initial concentration. 100 mg/l^{-1} , Adsorbent dosage: 1 g/50 ml)

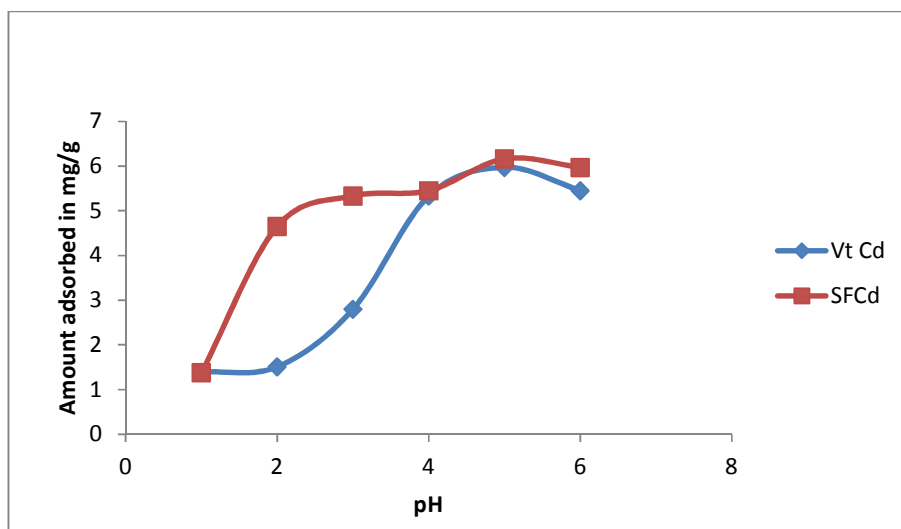


Fig. 8. Plot of amount of Cd adsorbed by Velvet tamarind and Sandal fruit versus pH (Temp. $28\pm 2^\circ\text{C}$; Agitation 200 rpm, Initial metal conc entration; 100 mg/l)

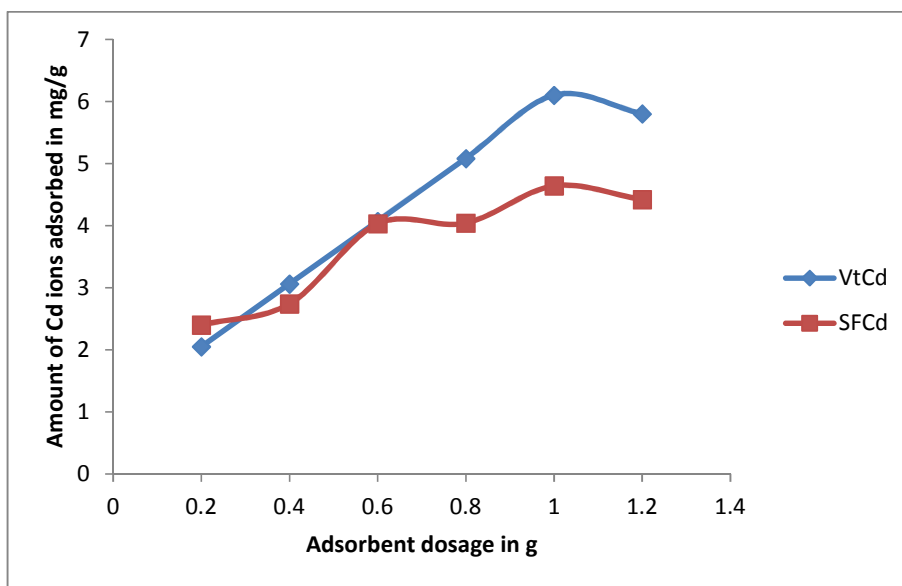


Fig. 9. Plot of amount of Cd ions adsorbed versus Adsorbent dosage by Velvet tamarind and Sandal fruit (Temp.28±2°C; pH.5.5, Agitation 200 rpm, Initial metal concentration; 100 mg/l)

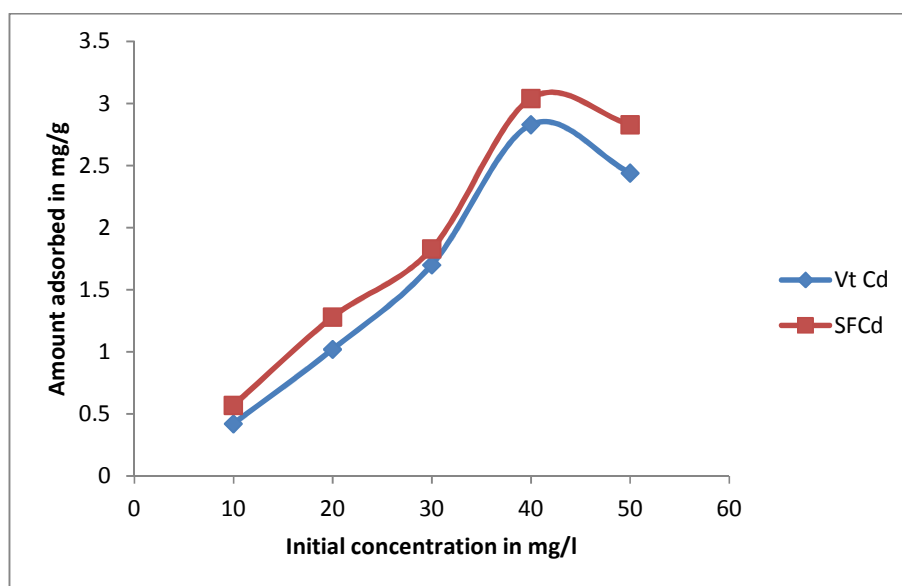


Fig. 10. Plot of amount of Cd(II) ions adsorbed in mg/g versus Initial concentration of Cd(II) ions by Velvet tamarind and Sandal fruit (Temp.28±2°C; pH.5.5, Agitation 200 rpm)

$$\log q_e = \log K_F + \left[\frac{1}{n}\right] \log C_e \quad (7)$$

Where K_F (mg/g)(mg/L) and n are the constants for adsorption capacity and intensity, respectively. A plot of $\log q_e$ versus $\log C_e$ gives a straight line of slope $1/n$ and intercepts $\log K_F$. As can be seen in Table 2, the r^2 values for both adsorbents for adsorption of Cd (II) were high (>0.95), indicating a good fit of the Freundlich

model to the adsorption process. This suggests that the surfaces of the adsorbents are heterogeneous. Furthermore, the values of n obtained for both metal ions lie between 1 and 10, indicating favourable adsorption [52].

3.8 Kinetic Modeling

The mechanism controlling adsorption was investigated with pseudo-first order and pseudo-

second order. The kinetic parameters are shown in Table 3. The pseudo-first order or Lagergren equation is based on the assumption that the rate of adsorption site occupation is proportional to the number of unoccupied sites (Lagergren, 1898). The constant K_1 and $q_{e,cal}$ were obtained from the slope and intercept of the plot of $\log(q_e - qt)$ versus t from equation 4. The R^2 values (Table 3) obtained for the metal ions showed a poor fit.

The pseudo-second order model assumes that the rate of occupation of adsorption sites is proportional to the square of the number of unoccupied sites [53]. The plot of t/qt versus t yields a straight line therefore the pseudo-second order equation is applicable. K_2 and q_e were calculated from the slope and intercept of the plot. This model provided a good fit to the experimental data for both metal ions, as seen from the r^2 values (Table 3):

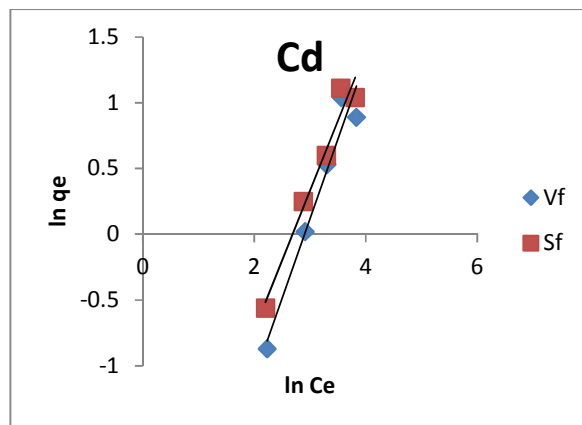


Fig. 11. Freundlich adsorption isotherm of Cadmium (II) ions by velvet tamarind and sandal fruit shells

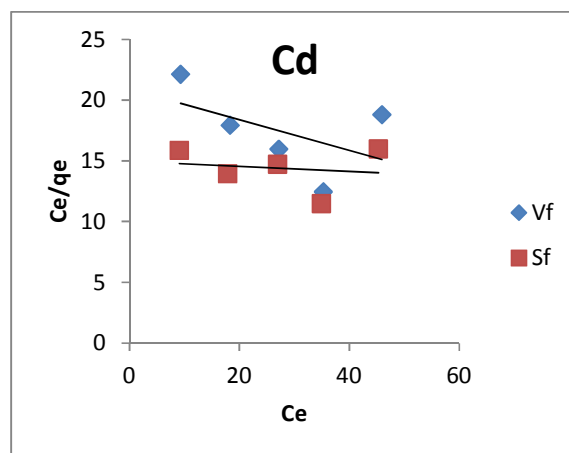


Fig. 12. Langmuir adsorption isotherm of Cadmium (II) ions by velvet tamarind and sandal fruit shells

Table 2. Values of langmuir and freundlich isotherm constants for adsorption of Cd(II)ions by velvet tamarind and sandal fruit

Heavy metal	Adsorbent	Freundlich			Langmuir		
		R^2	K_f	n	R^2	q_{max}	b
Cd	Vt	0.952	0.03	1.827	0.024	7.94	0.006
	S_f	0.964	0.058	1.943	0.253	50	0.001

Vt : Velvet tamarind (*Dialium indum*); Sf: Sandal fruit (*Santalum album*)

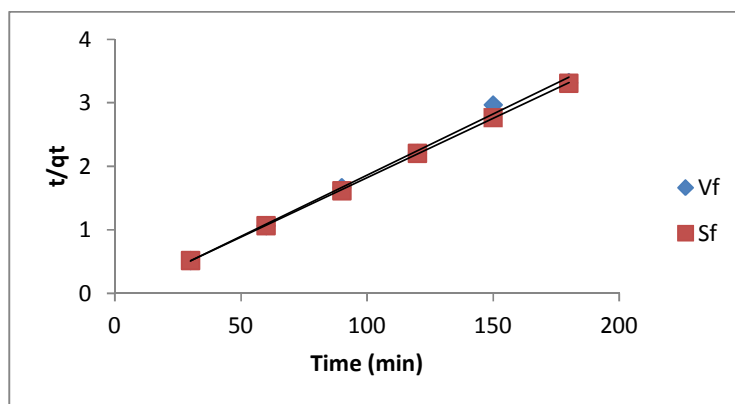


Fig. 13. Pseudo second order kinetic plot for Cadmium (II) ions adsorption by velvet tamarind and sandal fruits shell

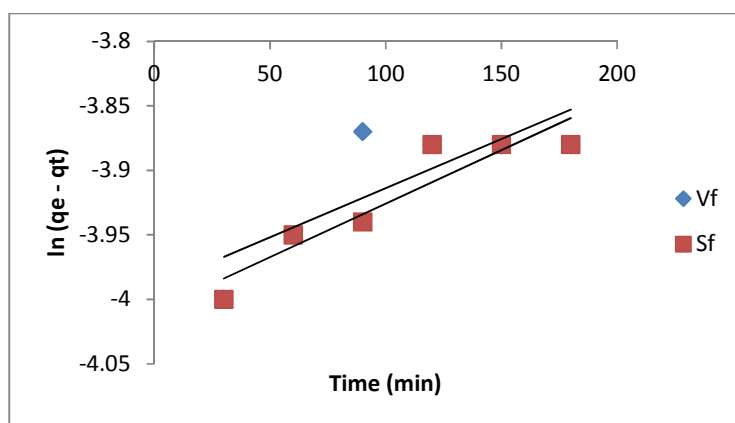


Fig. 14. Pseudo first order order kinetic plot for Cadmium (II) ions adsorption by velvet tamarind and sandal fruits shell

Table 3. The values of first- order and second order parameters obtained from the plot

Heavy metal	Adsorbent	First - order			Second-order		
		R ²	K ₁	q _e	R ²	q _e	K ₂
Cd	Vt	0.653	0.001	0.019	0.994	52.632	0.0004
	S _f	0.866	0.001	0.018	0.999	55.555	0.0003

4. CONCLUSION

Based upon the experimental results carried out in this work, the following conclusion can be drawn;

1. The FTIR spectra showed that certain functional groups OH, CO could be responsible for binding of metal ions from the solution by both adsorbents.
2. The heterogeneous natures of both adsorbents were revealed by the good fit of data to the Freundlich model indicating multilayer adsorption onto the adsorbents
3. Both adsorbents have a reasonably high surface area that can be used for adsorption
4. Extent of removal depends on the process parameters e.g Cd ion concentration, contact time, adsorbent dose and pH.
5. Both adsorbent can be applied as a cheap adsorbent for the removal of Cd(II) ions.
6. The adsorption capacity of treated sandal fruits for Cd(II) ions is more than that of velvet tamarind

ETHICAL APPROVAL

We confirm that this manuscript has not been published elsewhere and is not under consideration by another journal. Each author confirms the manuscript represents honest work. All authors have approved the manuscript. Each author agrees with the order in which his name appears on the title page.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Kumar A, Chandel D, Bala I, Muwalia A, Mankotiya L. Managing water pollution all the way through well-designed environmental biotechnology. *Journal of Colloid and Interface Science*. 2010;4(2): 45-56.
- Koby M, Demirabis E, Senturk E, Ince M. Adsorption of heavy metal ions from aqueous solution by activated carbon prepared from apricot stone. *Biores. Technol*. 2005;96:1518–1521.
- Amuda OS, Amoo IA. Coagulation/flocculation process and sludge conditioning in beverage industrial wastewater treatment. *J. Haz. Mater*. 2006;747:778-783.
- Demirbas E. Heavy metal adsorption onto agro-based waste materials: A review. *Journal of Hazard Materials*. 2008; 157(52):220-229.
- Egila JN, Dauda BEN, Jimoh T. Biosorptive removal of Cobalt (II) ions from aqueous solution by *Amaranthus* hydrides L. Stalk wastes. *African Journal of Biotechnology*. 2010;9(48):8192-8198.
- Erdem E, Karapinar N, Donat R. The removal of heavy metal cations by natural zeolites. *Journal of Colloid and Interface Science*. 2004;280(4):309–314.
- Kumar PD, Dara SS. Binding heavy metals ions with polymerized onion skin. *Journal of Polymer Science*. 2001;19(10):397-402.
- Okiemen FE, Ogbeifun DE, Navala GN, Kumsah CA. Binding of copper, cadmium and lead by modified cellulosic materials. *EJEAFChe*. 2005;3(4):702-709.
- Elifantz H, Tel-or E. Heavy metal biosorption by plant biomass of the macrophyte *Ludwigia stolonifera*. *Water Air Soil Pollution*. 2002;141:207- 218.
- Franca AS, Oliveira LS, Ferreira ME. Kinetics and equilibrium studies of methylene blue adsorption by spent coffee grounds. *Desalination*. 2009;249:267–272.
- Galadima A, Garba ZN. Heavy metals pollution in Nigeria: Causes and consequences. *Elixir Pollution*. 2012;45: 7917-7922.
- Gao H, Liu Y, Zeng G, Xu W, Li T, Xia W. Characterization of Cr(IV) removal from aqueous solution by a surplus agricultural waste- Rice-husk-straw. *J. Hazard, Mater*. 2007;150(2):446-452
- Karthika C, Vinnilamani N, Pattabhi S, Sekar M. Utilization of Sago Waste as an adsorbent for the removal of Pb(II) from aqueous solution: Kinetics and isotherm studies. *International Journal of Engineering Science and Technology*. 2010;2(6):1867–1879.
- American Society for Testing and Materials. Standard test method for determination of iodine number of activated carbon. Philadelphia, PA: ASTM Committee on Standards;1986
- Al-Qodah Z, Shawabkah R. Production and characterization of granular activated carbon from activated sludge. *Braz. J. Chem. Eng*. 2009;26(1):6-10.
- Alzaydian AS. Adsorption of methylene blue from aqueous solution onto a low – cost natural Jordanian Tripoli. *Am. J. Applied Sci*. 2009;6(6):1047-1058.
- Lasaka HK. Chromium (VI) removal in a semi-continues process of hollow fiber membrane with organic extractants. *Korean J. Chem. Eng*. 2005;22(6):894-898.
- Gil RA, Cerutti S, G'asquez JA, Olsina RA, Martinez LD. Preconcentration and speciation of chromium in drinking water samples by coupling of online sorption on activated carbon to ETAAS determination. *Talanta*. 2006;1(68):1065–1068.
- Pehlivan E, Altun T, Cetin S, Bhangher MI. Lead sorption by waste biomass of hazelnut and almond shell. *J. Hazard. Mater*; 2008.
DOI: 10.1016/j.jhazmat.2009.01.126
- Sun Q, Yang L. The adsorption of basic dyes from aqueous solutions on Modified peat– resin particle. *Water Research*. 2003;37:1535–1544.
- Tamura H, Hamaguchi T, Tokura S. Destruction of rigid crystalline structure to prepare chitin solution. *Advances in Chitin Science*. 2003;7:84–87.

22. Rafeah W, Ngaini Z, Usun JV. Removal of mercury, lead and copper from aqueous solution by activated carbon of palm oil empty fruit bunch. *World Appl. Sci. J.* 2009;5:84-91.
23. Hanif MA, Nadeem R, Bhatt HN, Ahmad NR, Ansari TM. Ni(II) biosorption by *Cassia fistula* (Golden Shower) biomass. *J. Hazard. Mater.* 2007;139(2):345-355.
24. Liu Y, Liu YJ. Biosorption isotherms, kinetics and thermodynamics. *Sep. Purif. Technol.* 2008;61:229-242.
25. Vinod VTP, Sashidhar RB, Sukumar AA. Competitive adsorption of toxic heavy metal contaminants by gum kondagogu: Anatural hydrocolloid. *Colloid Surf.* 2010; 75:490-495.
26. Ahalya M, Hussain KA, Ahmad S, Ahmad A. Role of sawdust in the removal of copper(II) from industrial wastes. *Wat. Res.* 2003;32(10):3085-3091.
27. Gimba CE, Olayemi JY, Ifijeh DOH, Kagbu JA. Adsorption of dyes by powdered and granulated activated carbon from coconut shell. *J. Chem. Soc. Niger.* 2001;26(1):23-27.
28. Hashem A, Akasha RA, Ghith A, Hussein DA. Adsorbent based on agricultural wastes for heavy metal and dye removal: A review. *Energy Edu. Sci. Technol.* 2007; 19:69-86.
29. Khaled AMM. A Comparative study for distribution of some heavy metals in aquatic organisms fished from Alexandria region. PhD Thesis, Chemistry Department, Faculty of Science. Alexandria University, Egypt; 1998.
30. Sud D, Mahajan G, Kaur NP. Agricultural waste materials as potential adsorbent for sequestering heavy metal ions form aqueous solutions – A review. *Bioresour. Technol.* 2008;99:6017- 6027.
31. Igwe JC, Ogunewe DN, Abia AA. Competitive adsorption of Zn(II), Cd (II) and Pb (II) ions from aqueous and non-aqueous solution by maize cob and husk. *Afr. J. Biotechnol.* 2005;4(10):1113-1116.
32. Aydin H, Bulut Y, Yerlikaya C. Removal of Cu(II) ion from aqueous solution by adsorption onto low-cost adsorbents. *J. Environ. Manage.* 2008;87:37-45.
33. Malakootian M, Nouri J, Hossaini H. Removal of heavy metals from paint industry's wastewater using Leca as an available adsorbent. *Int. J. Environ. Sci. Tech.* 2009;6(2):183-190,
34. Meunier N, Blais JF, Tyagi RD. Removal of heavy metals from acid soil leachate using cocoa shells in a batch counter-current sorption process. *Hydrometallurgy.* 2004; 73:225-235.
35. Pagnanelli F, Esposito A, Vegliò F. Multi-metallic modelling for biosorption of binary systems. *Water Res.* 2002;36: 4095-4105.
36. Okuo MJ, Oviawe AP. Selective sorption of mixed heavy metal ions using Cassava fibre modified with citric acid. *J. Chem. Soc. Niger.* 2007;32(2):247-253.
37. Aggarwal GS, Bhuptawat HK, Chaudhari S. Biosorption of aqueous chromium (VI) by *Tamarindus indica* seeds. *Bioresource Technology.* 2006;97:949-956.
38. Barkat M, Nibou D, Chegrouche S, Mellah A. Kinetics and thermodynamics studies ofchromium (VI) ions adsorption onto activated carbon from aqueous solutions. *Chemical Engineering and Processing: Process Intensification.* 2009;48(1):38-47.
39. Guler UA, Sarioglu M. Single and binary biosorption of Cu(II),Ni(II) and methylene blue by raw and pretreated *Spirogyria* sp: Equilibrium and kinetic modeling. *J. Environ. Chem. Eng.* 2013;1:369-377.
40. Mittal A, Krishnan L, Gupta VK. Removal and recovery of malachite green from wastewater using an agricultural waste material. *Sep. Purif. Technol.* 2005; 43(2):125-133.
41. Macht F, Eusterhues K, Pronk GJ, Totsche KU. Specific surface area of clay minerals: Comparison between atomic force microscopy measurement and bulk-gas (N₂) and liquid (EGME) adsorption methods. *Appl. Clay Sci.* 2001;53:20-26.
42. Raji C, Anirudhan TS. Kinetics of Pb(II) adsorption by poly-acrylamide grafted sawdust. *Indian J. Chem. Technol.* 1997; 4:157-162.
43. Ayranci E, Duman O. Adsorption behaviors of some phenolic compounds onto high specific area activated carbon cloth. *J. Hazard. Mater.* 2005;124:125-132.
44. Reddad Z, Gérente C, Andrès Y, Cloirec P. Adsorption of several metal ions onto a low- cost biosorbent: Kinetic and equilibrium studies. *Environ. Sci. Technol.* 2002;36:2067-2073.
45. Temkin MJ, Pyzhev V. Kinetics of ammonia synthesis on promoted iron

- catalyst. Acta Physicochim. URSS. 1940;12: 217–256.
46. Acharya J, Sahu JN, Mohanty CR, Meikap BC. Removal of lead (II) from wastewater by activated carbon developed from tamarind wood by zinc chloride activation. Chemical Engineering Journal. 2009;149: 249-262.
47. Khezami L, Capart R. Removal of chromium (VI) from aqueous solution by activated carbons: Kinetics and equilibrium studies. Journal of Hazardous Materials B. 2005;123:223-231.
48. Langmuir I. The adsorption of gases on plane surfaces of glass, mica and platinum. J. Am. Chem. Soc. 1918;40: 1361–1368.
49. Freundlich HMF. Over the adsorption in solution. J. Phys. Chem. 1906;57:385–471.
50. Patil S, Bhole A, Natrajan G. Scavenging of Ni(II) Metal Ions by Adsorption on PAC and Babhul Bark. J. Environ. Sci. Eng. 2006;48(3):203-208.
51. International Programme on chemical safety. Inorganic chromium (III) compounds, Draft. Concise International Chemical Assessment document. WHO. Geneva; 2006.
52. Lagergren S. About the theory of so-called adsorption of soluble substances, K. Sven. Ventenskapsakad. Handl. Band 1898;24:1–39.
53. Ho YS. Review of second order models for adsorption systems. J. Hazard. Mater. 2006;136:681–689.

© 2016 Jibrin et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://sciencedomain.org/review-history/12370>