



Biochemical Parameters of Female Wistar Rats Fed Shellfish and Cabbage as Potential Nutritive Supplements for Malnutrition

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Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

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ABSTRACT

Inability to meet the nutritional requirements causes malnutrition which further leads to various disorders and makes people susceptible to infections. Malnutrition is a serious problem globally and this research was conducted with an aim to understand the nutritive potentials of Shellfish and Cabbage to help deal with malnutrition. Female Wistar rats in post weaning stage were placed in six different groups on the basis of diets provided for a duration of 20 weeks. Investigations for biochemical parameters such as Total Protein, Albumin, enzymes Aspartate aminotransferase (AST), Alanine aminotransferase (ALT), Alkaline phosphatase (ALP), Bilirubin, Creatinine, Blood Urea Nitrogen, Cholesterol and Triglycerides were done to determine the potentials of these food sources as dietary supplements.

Keywords: Nutritive supplements; cabbage; shellfish; total protein; triglycerides.

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1. INTRODUCTION

Malnutrition is the condition in which a person has deficiency or excess of nutrients. Accordingly, malnutrition includes undernutrition involving deficiency of nutrients and also conditions such as being overweight and obese. Malnutrition can be both, a cause of the disease as well as a consequence of a disease [1]. Women are more likely to suffer from nutritional deficiencies than men are, for reasons including women's reproductive biology, low social status, poverty and lack of education [2]. India is known to have one of the highest number of malnourished children and upto a million children die before one month of age as many young mothers are anemic. Mortality rates of both mothers as well as children due to malnutrition are considerably high [3]. During early age, as the development of brain as well as body as a whole is at its peak, undernutrition in childhood is definitely going to leave a long lasting mark on an individual's development and body functioning [4]. It is known that proper nutrition is essential in achieving a satisfactory level of human development [5].

Malnutrition affects biochemical parameters negatively. Hypoinsulinemia and discrete hypoglycemia were found in adult Wistar rats, due to protein restriction during lactation [6,7]. Significant decreases in total protein concentration in mice have also been observed [7-10]. Changes in blood sugar, insulinemia and leptinemia in Wistar rats, caused by protein restriction during gestation and/or lactation, can be passed transgenerationally to the second generation of pups [7, 11]. Protein malnutrition reduces the content of many proteins in the body, including enzymes and hormonal proteins [12,13]. Physicians more often rely on albumin levels to assess a patient's nutritional status [13,14]. Albumin concentration is a better predictor of underlying malnutrition in the cardiac transplant recipients than the body mass index [13,15]. It is also a better prediction factor in the evaluation of malnutrition in patients undergoing elective orthopaedic surgery [13,16]. A lower concentration of albumin may be also related to the liver function damage [13,17]. Malnutrition is known to affect liver cells and causes liver enzyme imbalances. Several studies have shown that malnutrition can increase the levels of liver enzymes such as ALT and AST in patients. Such changes have been reported in different degrees of malnutrition. The amount of ALP was found to be lower in malnourished children [18-20].

Shellfishes are an important source of high quality animal protein [21]. Shellfishes also have lipids, glycogen and minerals that contribute to the nutritional value of their soft tissues. These molecules along with minerals and minor components of lipophilic and hydrophilic nature, contribute to its organoleptic characteristics and nutritional value [22]. Protein is the most abundant biochemical component in tissues and in some Shellfishes, it may be an alternative energy reserve during gametogenesis [23]. Many studies have suggested that increasing consumption of plant foods like Cabbage decreases the risk of obesity, diabetes, heart disease and overall mortality while promoting a healthy complexion, increased energy and overall lower weight. Cabbage contains beta-carotene, lutein and zeaxanthin, carotenoids that are a large class of natural plant pigments. They have chemo-protective effects, exhibit strong antioxidant properties and may reduce the risk of age related macular degeneration and some types of cancer [24-26]. Finding solutions to deal with the problem of malnutrition is the need of the hour. This paper focuses on the biochemical parameters of Wistar rats fed Shellfish and cabbage as potential nutritive supplements to help overcome malnutrition.

2. MATERIALS AND METHODS

After obtaining the required approval from Institutional Animal Ethics Committee (IAEC), 36 female Wistar rats in post weaning stage, obtained from Bharat Serums and Vaccines, Thane were brought to the Animal house at Dr. L.H. Hiranandani College of Pharmacy, CHM College Campus, Ulhasnagar. The rats were housed in Polypropylene cages with corn cob bedding. Rats were acclimatized for two weeks. They were provided normal rat feed pellets obtained from Nutrivet Lifesciences, Pune *ad libitum* during the acclimatization period. Water was provided *ad libitum* to all the animals throughout the experiment duration. After two weeks, rats were divided into six groups randomly, with six rats in each group. They were fed group specific diet as: Group 1 rats were fed Normal rat feed pellets *ad libitum* throughout the experiments. Rats in Group 2 were fed Normal rat feed pellets for 75% of their dietary requirements and remaining 25% was Shellfish. Group 3 rats were fed Normal rat feed pellets for 75% of their dietary requirements and remaining 25% was Cabbage. Group 4 rats were malnourished as they were on a restricted diet, being fed just 50% of their dietary requirement of rat feed pellets. Group 5 rats were also

malnourished being fed just 50% of their dietary requirement of rat feed pellets but these were provided with Shellfish as dietary supplement for the remaining 50% of their dietary requirements. Rats in Group 6 were malnourished too, being fed just 50% of their dietary requirement of rat feed pellets and these were provided with Cabbage as dietary supplement for the remaining 50% of their dietary requirements.

Feed consumption of Group 1 rats (*ad libitum*) was noted daily and it served the basis for calculation of feed requirements for the other groups. Shell fish (Squid) and Green Cabbage were obtained from local market, cleaned with water, cut into small pieces, weighed and fed to the rats as per their feeding groups. The feeding experiments were carried out for 20 weeks after which the rats were sacrificed using CO₂ chamber. Blood samples to study biochemical parameters were collected by cardiac puncture method and serum was used for the biochemical tests. The parameters investigated were Bilirubin (Modified Jendrassik and Grof's method), Aspartate aminotransferase (Modified Reitman and Frankel's Colorimetric DNPH method), Alanine aminotransferase (Reitman and Frankel's Colorimetric DNPH method), Alkaline Phosphatase (p-NPP kinetic method), Total protein (Biuret method), Albumin (BCG dye binding method), Globulin (Calculated using Total protein and Albumin values), Blood Urea Nitrogen (calculated using Urea estimated by Berthelot method), Creatinine (Modified Jaffe's method), Random Blood Sugar (GOD POD method), Cholesterol (Trinder CHOD/PAP method), Triglycerides (GPO/PAP method).

2.1 Statistical Analysis

Minitab Software was used to analyze the result by one way analysis of variance (ANOVA) to determine the differences between the mean values at P < 0.05 level followed by Tukey post hoc test. Data presented as Mean ± SD. Graphs were created using Microsoft Excel.

3. RESULTS AND DISCUSSION

The Bilirubin (Total) content did not show any significant difference among any of the groups. With respect to AST (Aspartate aminotransferase) enzyme, Group 5 and 6 showed significant difference compared to group 1 (Normal rat feed). Highest AST levels were shown by group 1 and lowest by group 6. Normal rat feed group 1 had highest while Groups 4 and 6 showed lowest levels of ALT (Alanine aminotransferase) enzyme, however, it was statistically insignificant. Group 6 rats which were malnourished and fed Cabbage showed lowest level of ALP (Alkaline Phosphatase) enzyme and it was significantly different as compared to group 3 rats with normal feed and Cabbage which showed highest value of ALP. Total Protein content in malnourished rats (group 4) was significantly lower to groups 1 and 2 while that of group 5 was significantly higher from group 4. Both the Shellfish fed groups showed higher level of total protein. Albumin content of groups 4 and 6 was significantly lower compared to groups 1, 2 and 3 while that of group 5 malnourished rats being fed Shellfish was

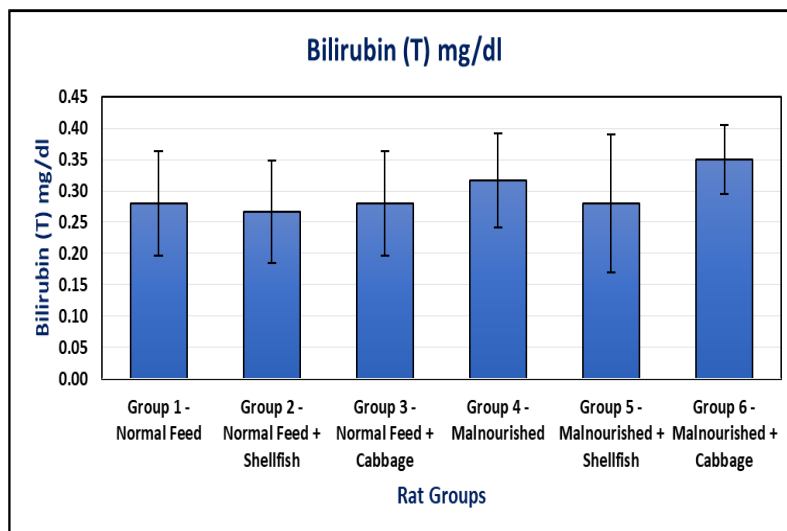


Fig. 1. Graph of Bilirubin (T) (Mean ±SD) vs Rat groups

Table 1. Biochemical parameters

Rat Groups	Bili (T) mg/dl	AST (IU/L)	ALT (IU/L)	ALP (IU/L)	Total Protein (g/dl)	Albumin (g/dl)	Globulin (g/dl)	Creatinine (mg/dl)	RBS (mg/dl)	BUN (mg/dl)	Cholesterol (mg/dl)	Triglycerides (mg/dl)
Group 1 - Normal Feed	0.28 ± 0.08	222.4 ± 37	148.2 ± 28.00	235.80 ± 103.00	7.04 ± 0.21	3.40 ± 0.12	3.64 ± 0.15	0.62 ± 0.08	84.40 ± 14.88	17.24 ± 1.58	51.40 ± 8.26	177.60 ± 20.28
Group 2 - Normal Feed + Shellfish	0.27 ± 0.08	183.80 ± 46.20	135.80 ± 37.60	268.30 ± 82.50	7.15 ± 0.21	3.42 ± 0.16	3.60 ± 0.09	0.65 ± 0.05	95.00 ± 10.55	16.72 ± 1.84	54.50 ± 4.85	191.67 ± 16.17
Group 3 - Normal Feed + Cabbage	0.28 ± 0.08	178.60 ± 22.60	120.40 ± 21.52	377.20 ± 118.30	6.96 ± 0.23	3.42 ± 0.13	3.54 ± 0.18	0.58 ± 0.08	93.80 ± 20.97	16.32 ± 0.73	48.00 ± 4.06	193.80 ± 15.06
Group 4 - Malnourished	0.32 ± 0.08	171.33 ± 23.47	118.83 ± 14.12	286.70 ± 70.50	6.42 ± 0.48*	3.00 ± 0.14*	3.42 ± 0.38	0.57 ± 0.05	80.17 ± 12.86	17.72 ± 1.28	50.17 ± 11.09	153.00 ± 35.50
Group 5 - Malnourished + Shellfish	0.28 ± 0.11	162.00 ± 21.76*	123.20 ± 20.61	250 ± 112.30	7.08 ± 0.41*	3.36 ± 0.21*	3.72 ± 0.41	0.60 ± 0.07	87.20 ± 7.63	17.2 ± 1.26	51.80 ± 10.26	181.60 ± 37.10
Group 6 - Malnourished + Cabbage	0.35 ± 0.05	152.80 ± 28.80*	118.80 ± 29.10	173.70 ± 40.00*	6.72 ± 0.43	2.97 ± 0.27*	3.75 ± 0.34	0.48 ± 0.04*	86.67 ± 9.35	19.77 ± 1.12*	49.00 ± 7.56	135.00 ± 19.41*

Data presented as Mean ± SD; *Means are statistically significant at P<0.05 (One way ANOVA followed by Tukey post hoc test). Bili (T)- Total Bilirubin, AST – Aspartate Aminotransferase, ALT- Alanine aminotransferase, ALP- Alkaline Phosphatase, RBS- Random Blood Sugar, BUN – Blood Urea Nitrogen

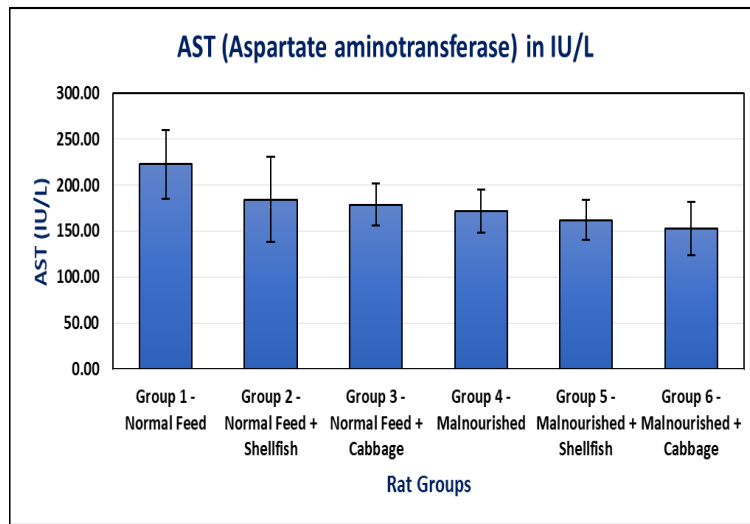


Fig. 2. Graph of AST (Mean \pm SD) vs Rat groups

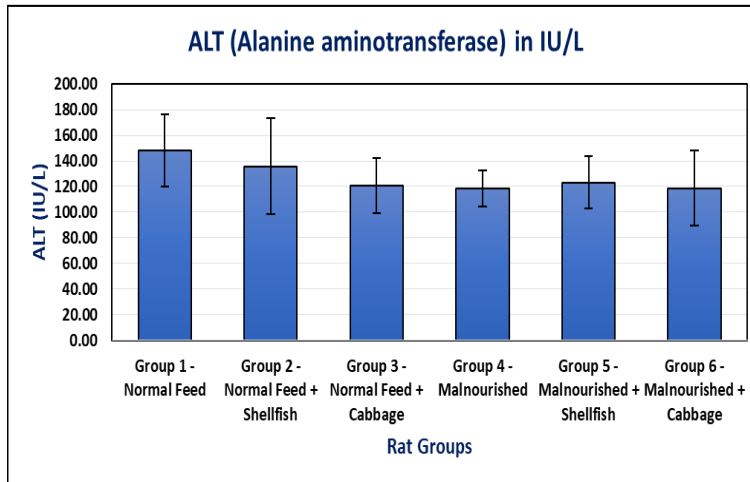


Fig. 3. Graph of ALT (Mean \pm SD) vs Rat groups

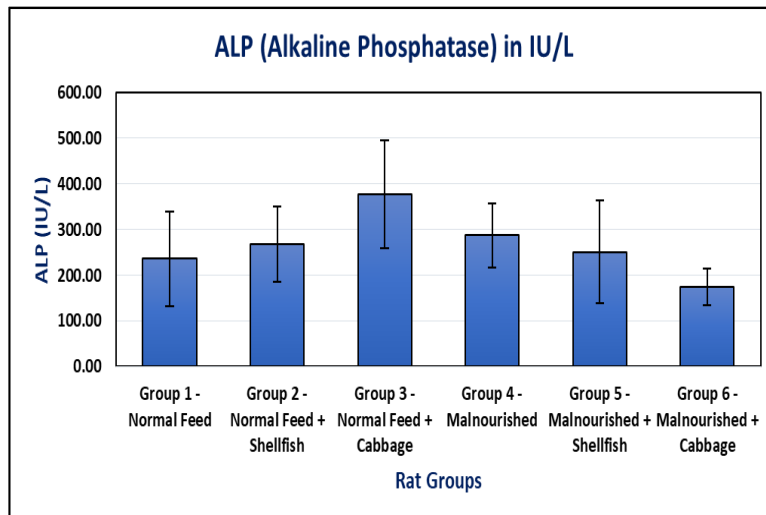


Fig. 4. Graph of ALP (Mean \pm SD) vs Rat groups

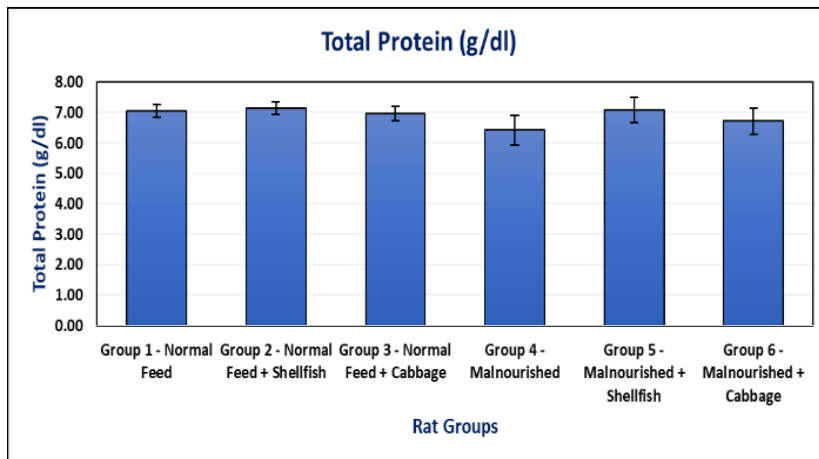


Fig. 5. Graph of Total protein (Mean \pm SD) vs Rat groups

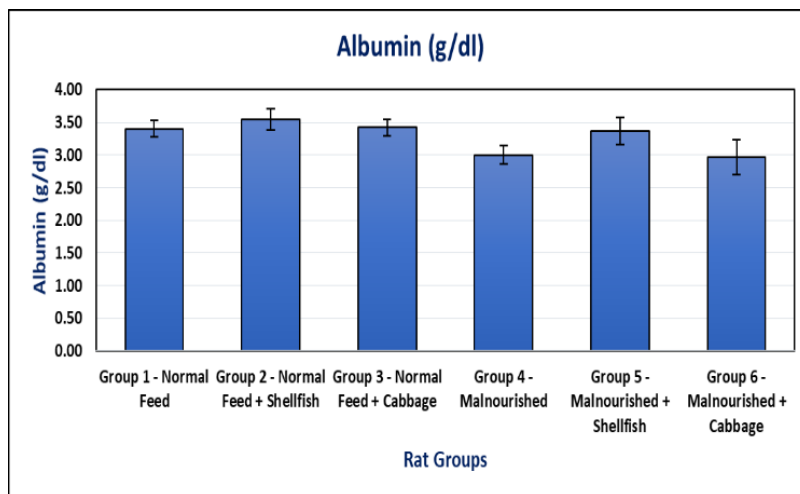


Fig. 6. Graph of Albumin (Mean \pm SD) vs Rat groups

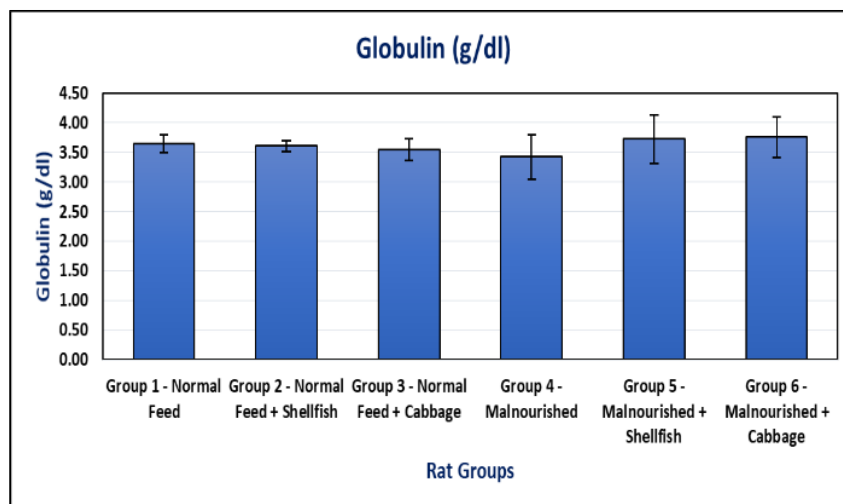


Fig. 7. Graph of Globulin (Mean \pm SD) vs Rat groups

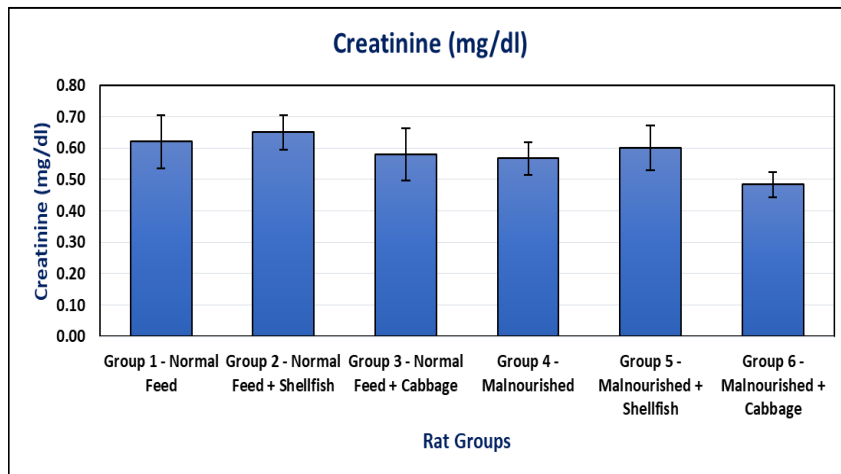


Fig. 8. Graph of Creatinine (Mean \pm SD) vs Rat Groups

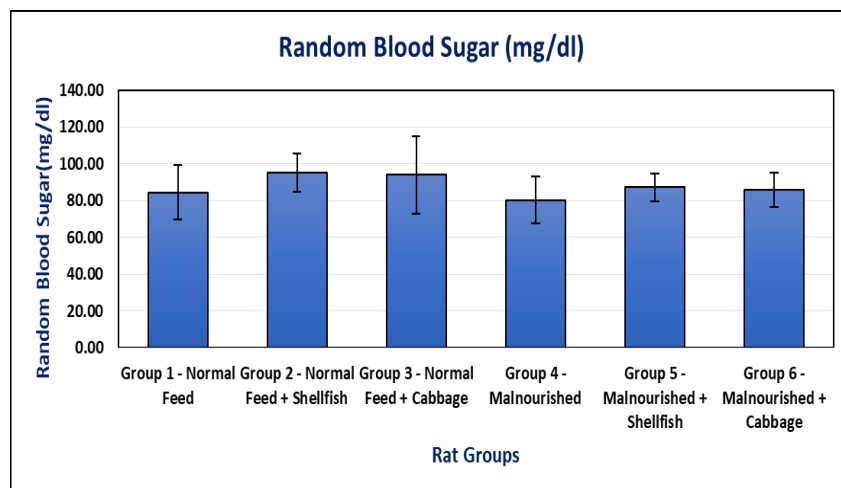


Fig. 9. Graph of random blood sugar (Mean \pm SD)

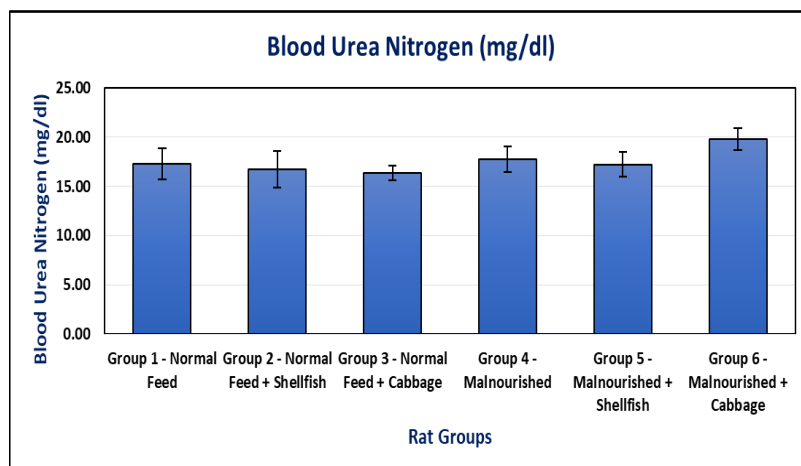


Fig. 10. Graph of blood urea nitrogen (Mean vs Rat groups \pm SD) vs Rat Groups

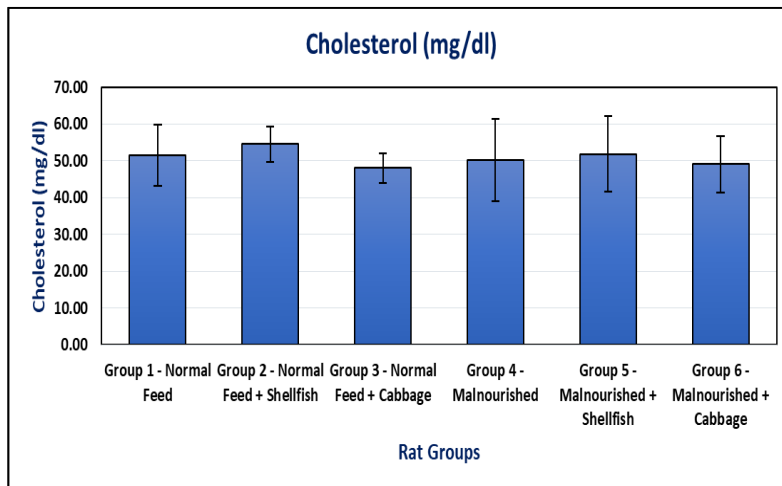


Fig. 11. Graph of Cholesterol (Mean ±SD) vs Rat groups

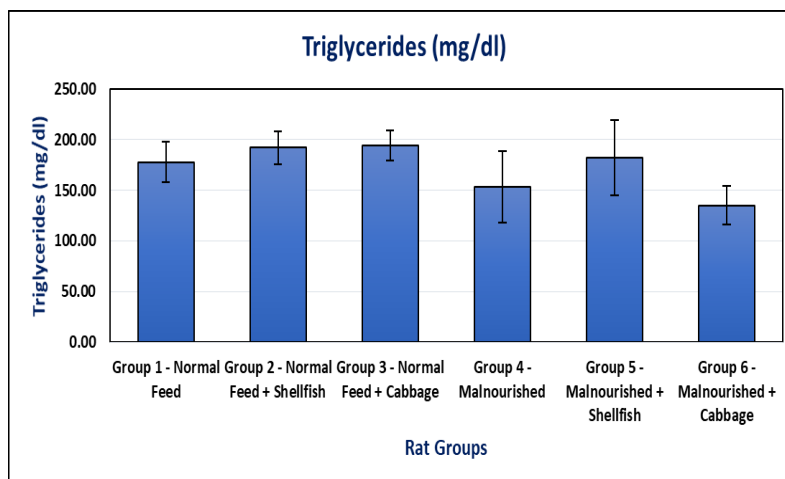


Fig. 12. Graph of Triglycerides (Mean ±SD) vs Rat groups

significantly higher compared to group 4. No statistically significant result observed with respect to Globulin. Creatinine levels of Group 6 were lowest and significantly different from groups 1 and 2. Random Blood sugar was highest for group 2 and lowest for group 4, no statistically significant difference was observed. Blood Urea Nitrogen level was highest in group 6 and statistically significant in comparison to groups 1, 2 and 3. It was lowest in group 3. Both Shellfish fed groups i.e. group 2 and 5 showed higher Cholesterol levels compared to normal feed group but no statistically significant difference was observed. Triglyceride levels were lowest in group 6 and showed significant difference compared to groups 1, 2 and 3. Highest level of Triglycerides was observed in group 3.

4. CONCLUSION

In this study, Shellfish and Cabbage reduced the levels of enzymes Aspartate aminotransferase and Alkaline Phosphatase when given as nutritive supplements to malnourished rats. Shellfish diet for malnourished rats significantly increased the Total protein as well as Albumin content. Cabbage diet given to malnourished rats was found to significantly increase Blood Urea nitrogen and significantly lowered Creatinine and Triglyceride levels.

ETHICAL APPROVAL

Animal Ethic committee approval has been collected and preserved by the author(s).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Saunders J, Smith T. Malnutrition: Causes and consequences. *Clinical medicine* (London, England). 2010;10(6):624–627. Available:<https://doi.org/10.7861/clinmedicine.10-6-624>
2. Elizabeth I, Ransom, Leslie K. Elder nutrition of women and adolescent girls: why it matters? Population Reference Bureau: Publications, Article; 2003.
3. Narayan, Jitendra, John, Denny, Karanth, Dr. Nirupama. Malnutrition in India: Status and government initiatives. *Journal of Public Health Policy*. 40. DOI: 10.1057/s41271-018-0149-5
4. Lisa C. Smith, Lawrence Haddad. Reducing child undernutrition: Past Drivers and Priorities for the Post-MDG Era, *World Development*. 2015;68:180-204, ISSN 0305-750X, Available:<https://doi.org/10.1016/j.worlddev.2014.11.014>.
5. Humbwavali JB, Giugliani C, Nunes LN. et al. Malnutrition and its associated factors: A cross-sectional study with children under 2 years in a suburban area in Angola. *BMC Public Health*. 2019;19:220. Available:<https://doi.org/10.1186/s12889-019-6543-5>
6. Fagundes AT, Moura EG, Passos MC, Oliveira E, Toste FP, Bonomo IT, Trevenzoli IH, Garcia RM, Lisboa PC. Maternal low-protein diet during lactation programmes body composition and glucose homeostasis in the adult rat offspring. *Brit J Nutr*. 2007;98(5):922-928.
7. Estrela DC, Lemes CGC AT. B. Guimarães G, Malafaia. Effects of short-term malnutrition in rats. *Scientia Plena*. 2014;10(07).
8. Xavier JG, Favero ME, Vinolo MAR, Rogero MM, Dagli MLZ, Aranha-Chavez VE, Borojevic R, Borelli P. Protein-energy malnutrition alters histological and ultrastructural characteristics of the bone marrow and decreases haematopoiesis in adult mice. *Histol Histopathol*. 2007;22(6):651- 660.
9. Borelli P, Blatt S, Pereira J, Maurino BB, Tsujita M, Souza AC, Xavier JG, Fock RA. Reduction of erythroid progenitors in protein-energy malnutrition. *Brit J Nutr*. 2007;97(2):307-314.
10. Malafaia G, Martins RF, Silva ME. Avaliação dos efeitos, em curto prazo, da deficiência proteica nos parâmetros físicos e bioquímicos de camundongos Swiss. *SaBios: Rev Saúde e Biol*. Jul/Dez 2009;4(2):21-33.
11. Pinheiro AR, Salvucci ID, Aguila MB, Mandarin-de-Lacerda CA. Protein restriction during gestation and/or lactation causes adverse transgenerational effects on biometry and glucose metabolism in F1 and F2 progenies of rats. *Clin Sci*. 2008; 114(5):381-392.
12. Wykes LJ, Fiorotto M, Burrin DG, Del Rosario M, Frazer ME, Pond WG, Jahoor, F. Chronic low protein intake reduces tissue protein synthesis in a pig model of protein malnutrition. *Journal of Nutrition*.1996;126(5):1481–1488.
13. Sławomir Lewicki , Monika Leśniak , Jerzy Bertrandt, Bolesław Kalicki, Jacek Z. Kubiak, Aneta Lewicka. The long-term effect of a protein-deficient-diet enriched with vitamin B6 on the blood parameters in unexercised and exercised rats. *Food And Agricultural Immunology*. 2018;29(1):722–734 Available:<https://doi.org/10.1080/09540105.2018.1439900>
14. Bharadwaj S, Ginoya S, Tandon P, Gohel TD, Guirguis J, Vallabh H, Hanouneh I. Malnutrition: Laboratory markers vs nutritional assessment. *Gastroenterology Report*. 2016;4(4):272–280.
15. Prenner G, Wasler A, Fahrleinter-Pammer, A, Werkgartner G, Mischinger HJ, Koter S., Wagner D. The role of serum albumin in the prediction of malnutrition in patients at least five year after heart transplantation. *Clinical Transplantation*. 2014;28:737–742
16. Cross MB, Yi PH, Thomas CF, Garcia J, Della Valle CJ. Evaluation of malnutrition in orthopaedic surgery. *Journal of the American Academy of Orthopaedic Surgeons*. 2014;22(3):193–199

17. Lee JS. Albumin for end-stage liver disease. The Korean Journal of Internal Medicine. 2012;27(1):13–19.
18. Taher A, Nahar N, Haque M, Rahman A, Chowdhury MSI. Serum Aspartate Aminotransferase (AST) and Alanine Aminotransferase (ALT) Levels in Different Grades of Protein Energy Malnutrition. J Bangladesh Society Physiologist. 1970;2: 17–9.
DOI: 10.3329/jbsp.v2i0.978.19
19. Islam MN, Karmacharya K. Role of serum alanine aminotransferase aspartate aminotransferase and alkaline phosphatase in early detection of protein energy malnutrition. J Nepal Pediatr Society. 1970;27(2):68–72.
DOI: 10.3126/jnps.v27i2.1412
20. Karajibani, Mansour, Montazerifar, Farzaneh, Hosseini, Razieh, Suni, Fatemeh, Dashipour, Ali, Fadaeimokhtarkanlo, Mahshid. The Relationship Between Malnutrition and Liver Enzymes in Hospitalized Children in Zahedan: A case-Control Study. Zahedan Journal of Research in Medical Sciences. In Press; 1994.
DOI: 10.5812/zjrms.102994
21. Karsoon Tan, Leongseng Lim, Ya Peng, Kit-Leong Cheong. Effects of food processing on the lipid nutritional quality of commercially important fish and shellfish, Food Chemistry: X, 2023;20. ,101034,ISSN 2590-1575,
Available:https://doi.org/10.1016/j.fochx.2023.101034.
22. Orban E, Di Lena G, Navigato T, Casini I, Caproni R, Santaroni G, Giulini G. Nutritional and commercial quality of the striped Venus clam, *Chamelea gallina*, from the Adriatic sea. Food Chem. 2007; 101:1063- 1070.
23. Galap C, Leboulenger F, Grillot JP. Seasonal variations in biochemical constituents during the reproductive cycle of the female dog cockle *Glycymeris glycymeris*. Mar. Biol. 1997; 129:625-634.
24. Michael W, Pius J, Beverley H, Detmar B. Molecular and cellular mechanisms of cadmium carcinogenesis. Toxicol. 1999; 192(2-3):95-117.
25. Lynn A, Collins A, Fuller Z, Hillman K, Radcliffe B. Cruciferous vegetables and colo-rectal cancer. Proc. Nutr. Soc. 2006; 65:135-144.
26. Uuh-Narvaez JJ, Segura-Campos M. R. Cabbage (*Brassica oleracea* var. *capitata*): A food with functional properties aimed to type 2 diabetes prevention and management. J Food Sci. 2021;86:4775–4798.
Available:https://doi.org/10.1111/1750-3841.15939

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